

REVIEW

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# Nanomaterial-related hemoglobin-based oxygen carriers, with emphasis on liposome and nano-capsules, for biomedical applications: current status and future perspectives

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## Abstract

Oxygen is necessary for life and plays a key pivotal in maintaining normal physiological functions and treat of diseases. Hemoglobin-based oxygen carriers (HBOCs) have been studied and developed as a replacement for red blood cells (RBCs) in oxygen transport due to their similar oxygen-carrying capacities. However, applications of HBOCs are hindered by vasoactivity, oxidative toxicity, and a relatively short circulatory half-life. With advancements in nanotechnology, Hb encapsulation, absorption, bioconjugation, entrapment, and attachment to nanomaterials have been used to prepare nanomaterial-related HBOCs to address these challenges and pend their application in several biomedical and therapeutic contexts. This review focuses on the progress of this class of nanomaterial-related HBOCs in the fields of hemorrhagic shock, ischemic stroke, cancer, and wound healing, and speculates on future research directions. The advancements in nanomaterial-related HBOCs are expected to lead significant breakthroughs in blood substitutes, enabling their widespread use in the treatment of clinical diseases.

**Keywords** Oxygen transport, Hemoglobin-based oxygen carriers (HBOCs), Nanomaterials, Hemorrhagic shock, Cancer

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## Introduction

Oxygen (O<sub>2</sub>) is essential for life [1] and physiological processes such as cell growth, differentiation, metabolism, and tissue homeostasis [2, 3]. Factors such as trauma [4], blood loss [5], pernicious anemia [6] and ischemic cerebrovascular disease [7, 8] can decrease tissue oxygen content, lead to hypoxia [9]. This, in turn, can induce organ dysfunction, particularly in the brain [10], heart [11] and kidney [12] and leading to irreversible damage and/ or death [13]. Besides, the hypoxic microenvironment of tumor tissues can decrease the sensitivity of malignant tumors to antitumor drugs and enhance the invasion and metastasis abilities of tumor cells [14–16]. Thus, maintaining an adequate O<sub>2</sub> supply to tissues and organs, is essential for maintaining the physiological functions of the body, and it is critical for disease treatment [17].

As natural oxygen carriers, red blood cells (RBCs) can bind to O<sub>2</sub> in the lungs and deliver throughout to the body. This process is essential to meet the metabolic demands of the body [18, 19] and ensure normal organ function [20]. Hemoglobin (Hb), the main O<sub>2</sub> carrier in RBCs, facilitates O<sub>2</sub> delivery via its well-known oxygen-carrying function [21]. In adults, Hb consist of four subunits ( $\alpha$ 1,  $\beta$ 1,  $\alpha$ 2, and  $\beta$ 2) that form a tetrameric protein containing a central ferrous heme, enabling it to transport O<sub>2</sub> by binding reversibly to O<sub>2</sub> [22, 23].

Numerous factors, such as trauma, blood loss, and pernicious anemia [24, 25], can potentially cause decrease in RBCs and Hb levels in the bloodstream. Timely transfusion of RBCs, restoration of blood volume in the body, and ensuring adequate of O<sub>2</sub> supply are essential for saving lives [26–28]. Indeed, these measures are widely practiced in clinical and military rescue scenarios [29]. However, the demand for natural blood often outpaces supply [30, 31]. Natural blood has a limited preservation period (<42 d), requires special storage and transportation conditions [32], and is susceptible to damage and adverse reactions when infused [33, 34]. Natural blood transfusion also carries risk of infectious diseases, requires associated, time-consuming, cross-matching [35, 36] and may conflict with religious beliefs [37, 38]. Therefore, blood substitutes that can fully and completely or partially replace the oxygen-carrying functions of RBCs have emerged. These substitutes can be classified into perfluorocarbons (PFCs) and hemoglobin-based oxygen carriers (HBOCs) [39, 40].

Perfluorocarbon emulsions, which physically solubilize O<sub>2</sub> and CO<sub>2</sub>, were the first blood substitutes to be tested as oxygen carriers [41]. However, the biosafety and stability concerns of PFCs limit their potential for clinical use. The first-generation products, Fluosol-DA<sup>®</sup>, was originally approved by the United States of America Food and

Drug Administration (FDA) for coronary transluminal angioplasty; however, it was withdrawn from the market due to O<sub>2</sub> delivery capacity, poor stability, and complement activation [42]. The second-generation product, Oxygent<sup>™</sup>, was also discontinued during phase III clinical trials in the USA because of an increased incidence of instroke and heart diseases in patients undergoing coronary artery bypass surgery [42–44].

HBOCs are a class of blood substitutes that are based on natural hemoglobin obtained through polymerization, cross-linking, and modification with polymer. HBOCs have gradually become the primary research focus for artificial oxygen carriers because they closely resemble the natural oxygen-carrying/-releasing characteristics of Hb [40, 45]. HBOCs development progressed after decades of laboratory and clinical research. However, only a few HBOCs have received worldwide approved for clinical use. Glutaraldehyde-polymerized bovine hemoglobin (HBOC-201, BioPure company), was approved for the treatment of acute pernicious anemia in South Africa and Russia in 2001 and 2010, respectively [46, 47]. PEGylated carboxyhemoglobin bovine (Sanguinate, Prolong Pharmaceuticals, USA), was registered in 2015 under the FDA classification as an “orphan drug” for the treatment of sickle cell anemia; it is currently in phase III clinical trials [48]. Despite these advancement, HBOCs development is also complicated by issues such as vasoactivity caused by nitric oxide (NO) scavenging [49], nephrotoxicity from dissociated dimers [50], oxidative toxicity of hemoglobin [51], and short circulation time [52], resulting in many HBOCs varieties being discontinued or excluded from further research due to safety concerns. Thus far, researchers have focused on addressing the safety concerns of HBOCs by improving strategies such as the cross-linking technique, and the preparation process of Hb, as well as infusion methods. For instance, preventing the dissociation of hemoglobin tetramer and reducing nephrotoxicity [53, 54] by optimizing the molecular weight of the polymer [55]. Hb oxidation is reduced by co-cross-linking Hb with internal biological enzymes such as superoxide dismutase (SOD) and catalase (CAT) [56–58]. While these technical strategies have reduced the safety concerns of HBOCs to extent; they are still in the preclinical research stage.

The role of nanomaterials in drug delivery and disease treatment has received increased research attention due to the rapid development of nanotechnology [59, 60], which provides new opportunities for developing HBOCs. Nanomaterial-related HBOCs (Nano-HBOCs) are prepared by organically conjugating of nanomaterials with hemoglobin through encapsulation, self-assembly, bioconjugation, entrapment, and attachment [61–63]. This approach not only closely mimics the physiological

structure of natural RBCs but also offers more advantages, such as reduced the vasoactivity, improved circulation time [64–66], and enhanced biological safety [67] (Table 1).

Although some excellent reviews have discussed Nano-HBOCs [35, 45, 65–68], there is a lack of detailed and systematic introductions, which, coupled with the rapid progress in the research of Nano-HBOCs applications, has resulted in a lack of reviews on the latest advances in the preparation and application of Nano-HBOCs. In this review, we introduce the specific research on Nano-HBOCs in biomedical fields over decade. We focus on the applications of Nano-HBOCs in hemorrhagic shock, ischemic stroke, cancer therapy, and wound healing (Fig. 1). Additionally, we systematically summarize the research progress, current challenges and future prospects.

### Nano-HBOCs for hemorrhagic shock therapy

Hemorrhagic shock (HS) is a complex condition caused by a reduction in the effective circulating blood volume [87, 88], resulting in irreversible tissue hypoxia [89], eventually triggers circulatory failure [90]. Increasing the oxygen-carrying capacity to restore O<sub>2</sub> supply to the

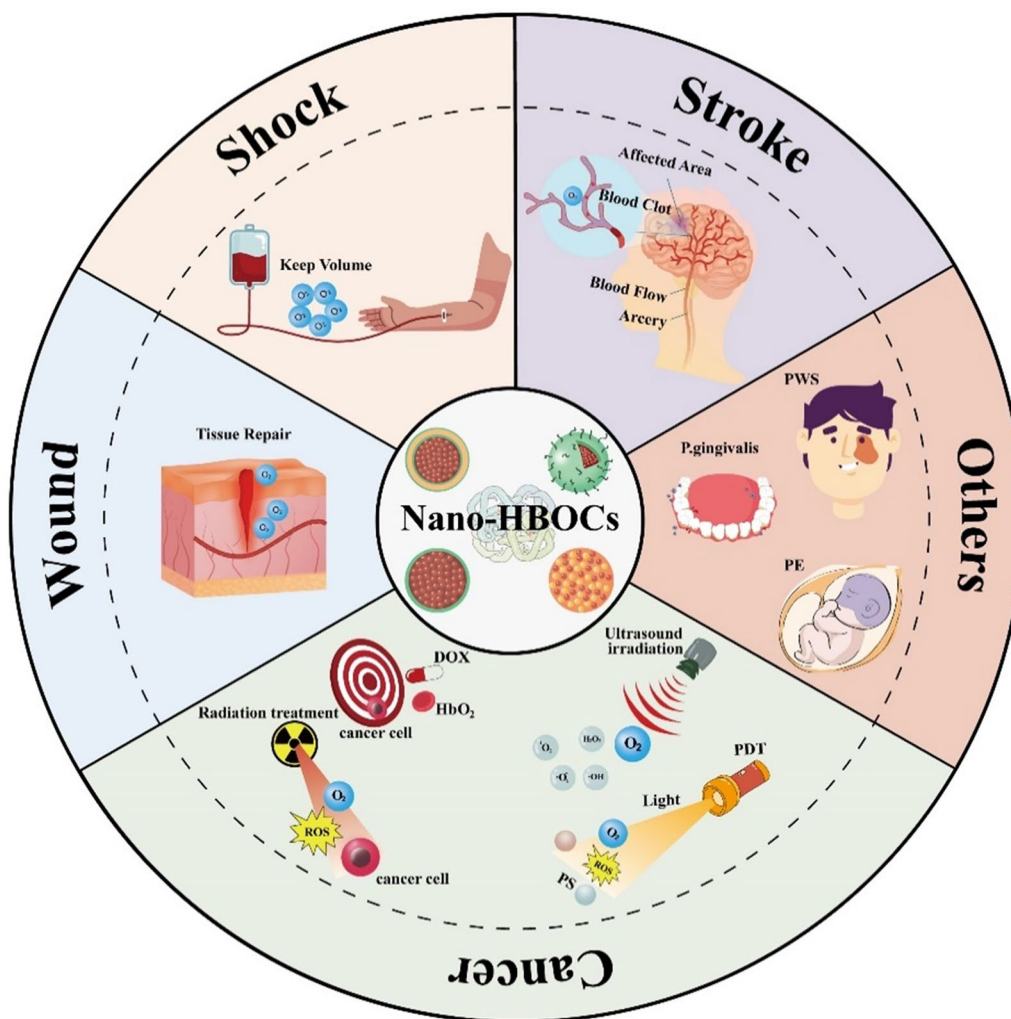
tissue is a necessary approach to treating HS [88, 91] and perverting circulatory failure.

### Restoring oxygen supply

Liposome-encapsulated human hemoglobin (LEH) is a type of HBOCs prepared by encapsulating intact Hb molecules in liposomes, also known to as hemoglobin vesicles (HbV) [65]. LEH has been extensively researched as a potential treatment for HS in animal models [92–95]. Studies have shown that LEH has excellent biocompatibility [96], high biological safety [97–99], easy metabolism and excretion [100, 101], and low vasoactivity [102]. Researchers at the Terumo Research and Development Center [69] (Terumo Co., Tokyo, Japan) used a high-speed emulsification method to encapsulate purified human Hb in liposomes that mimic RBCs membranes. They modified the lipid membrane surface with PEG to form a nanoscale liposomal oxygen carrier (namely TRM-645). In a mouse hypohemoglobinemic shock model, the intravenous infusion of TRM-645 rapidly restored mean arterial pressure (MPA) from 30 to 70 mmHg; its effect on restoring and maintaining MAP was comparable to that of native RBCs. The intracavitary infusion of TRM-645 significantly improved overall survival [103] in a

**Table 1** Several advantages of Nano-HBOCs, classified into seven categories, with information related to their formulations, Hb source, and references

Properties	Categories	Formulations	Hb source	References
Reducing the vasoactivity	TRM-645	Encapsulating purified Hb in liposomes mimicking the RBCs membrane	Human	[69]
Raising biological safety	LEH HbPs HbTcMs O <sub>2</sub> @Hb@ZIF-8	Employment of liposomes to encapsulate intact Hb molecules Hb encapsulated mPEG-PLGA using the double emulsion technique Hemoglobin-polymer conjugate formation, completion of Hb and PS co-loading Hb encapsulated in ZIF-8	Human Bovine Bovine Human	[65, 70–72]
Improving blood circulation time	PDA – LtEc ZIF-8@Hb Hb@AuNCs PFRT-RBCS	<i>Lumbricus terrestris</i> with PDA surface-coating Hb encapsulated in MOFs Hb and Au nanoparticles, combined and incorporated into the reported MOFs A large amount of ZnF16Pc tethered to the RBCs surface	Worm Bovine Bovine Human	[73–76]
Enhancing antioxidant capacity	Hb-PDA PtNP MXene@PDA NSs	Combination of Hb and PDA using a template-based co-precipitation technique Platinum nanoparticles with SOD and CAT-mimetic enzyme activities combined with HSA on the outer layer Coupling of hyaluronic acid and PDA catalyzed by H <sub>2</sub> O <sub>2</sub> /HbO <sub>2</sub> to encapsulated Ti <sub>3</sub> C <sub>2</sub> MXene nanosheets	Bovine Human	[77–79]
Mimicking physiology of RBCs	ErythroMer	Hb Encapsulated in deformable, hybrid peptidic-lipid nanoparticles	Human	[80]
Empowering O <sub>2</sub> targetingly	CPTK@PMH HCM DHCNPNS V(Hb)@DOX Au-Hb@ PLT NPs	Introducing PDA to noncovalently engage Hb and methoxatin to form PMH, and connecting to a specific fibrin-binding peptide Linking GLUT1, and bounding to the micelles via a condensation reaction to form Hb-conjugated micelles Hb and DOX encapsulated in PLGA Utilizing Hb via coupling ε-Caprolactone self assembles to form hollow V(Hb), and loading DOX into V(Hb) Coating Au-Hb NPs with PLT membrane	Human Bovine Human Bovine Human	[81–85]
Releasing O <sub>2</sub> controllably	MN	Loading with black phosphorus and Hb	Bovine	[86]



**Fig. 1** Schematic representation of the application of Nano-HBOCs in hemorrhagic shock, ischemic stroke, cancer, wound healing, and other disease treatment

murine of HS induced by sustained massive hemorrhage, with a better treatment effect RBCs. TRM-645, due to its smaller size, is more likely to enter the circulatory system from the bone marrow cavity, rendering it an effective method for critical scenarios and pre-hospital emergency care. Vivek et al. [104] utilized non-phospholipid anionic lipid conjugated PEG containing hexadecyl carbamoyl methyl hexadecanoate (HDAS) to modify the surface of LEH. This modification improved the host's tolerance to the immune response against LEH [105, 106], delayed its phagocytic clearance by monocytes, and increased the mean residence time in circulation. In a rat model of HS, LEH infusion restored blood volume, improved tissue oxygenation capacity, and ameliorated HS-induced systemic inflammation and multiple organ failure, suggesting that this modified LEH is a potential treatment for HS.

To address the critical need for a transportable, and temperature-stable blood substitute, a bio-synthetic, nano-artificial erythrocyte called Erythromer (EM) [107] has been developed. EM encapsulates human Hb within deformable, hybrid peptidic-lipid nanoparticles. This design, allows for a true physiological correlation between  $O_2$  affinity and tissue respiration. EM employs a novel shuttle, a small molecule allosteric effector, which adjusts  $P_{50}$  in response to pH changes, influencing  $Hb \sim O_2$  affinity. The use of EM allows for the mimicry of key physiological properties of natural RBCs, offering a promising solution to address the aforementioned critical need. Compared to the use of hydroxyethyl starch for resuscitation, EM maintains hemodynamics stability, increases arterial oxygen partial pressure, and improves acidosis in the HS rat and hemodilution mouse models [80]. Okamoto et al. [108] demonstrated



that the combination of bovine hemoglobin (bHb) and human serum albumin (HSA) forms a core-shell protein called HbX-HSA<sub>3</sub>, which has high oxygen affinity. HbX-HSA<sub>3</sub> facilitates O<sub>2</sub> delivery to hypoxic tissues during HS. In the rat HS model, HbX-HSA<sub>3</sub> effectively restores and maintains basic life indicators such as blood pressure, blood oxygen, respiration, and body temperature. It also reduces body lactate levels; and improve the survival rate of HS. Lu et al. [70] used methoxy polyethylene glycol-poly (D, L-lactide-co-glycolide) and mPEG-PLGA to encapsulate bHb using the double emulsion (w/o/w) technique for preparing hemoglobin-loaded nanoparticles (HbPs). This was based on the good biodegradability and biocompatibility characteristics of PLGA, which had a uniform particle size with stable oxygen-carrying function and excellent blood compatibility. In a controlled hemorrhage mouse model, HbPs maintained MAP, improved venous oxygen partial pressure, and restored tissue oxygen supply, demonstrating a promising application potential.

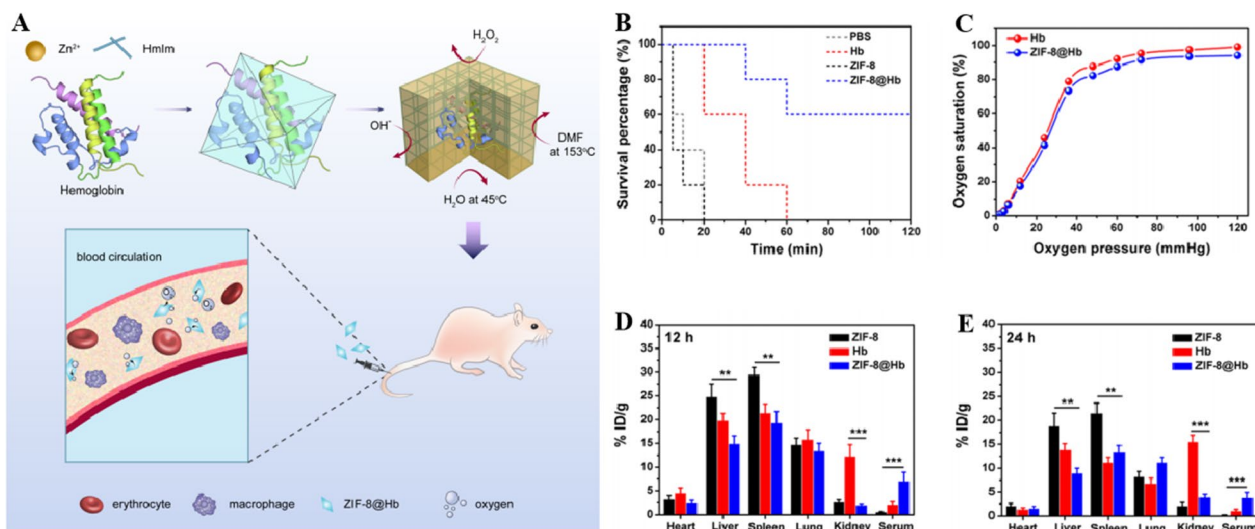
Polydopamine (PDA) has excellent biocompatibility and antioxidant capacity [109]. It can easily and efficiently adhere to the surface of various substrate materials [110], suggesting that PDA has the potential to serve as a universal modification platform for Hb. This can help address the issue of oxidative toxicity in HBOCs. Wang et al. [77] harnessed the unique characteristics of PDA to modify bHb and prepare Hb-PDA, which does not cause platelet aggregation or significant hemolysis. These novel Nano-HBOCs demonstrated outstanding oxygen affinity (P<sub>50</sub>=13.86 mmHg). Hu et al. [78] combined a template-based co-precipitation technique to develop Nano-oxygen carriers, (named Hb-PDA), which displayed uniform size and high biocompatibility. In vitro and in vivo experiments showed that Hb-PDA enhanced the antioxidant properties of Hb, alleviated the toxicity of free Hb, and maintained the O<sub>2</sub> delivery capacity. Further, the resuscitation efficacy of Hb-PDA was investigated using in rat HS model. The results revealed that Hb-PDA maintained the stability of the blood pressure for a longer time relative to using saline alone. Baidukova et al. [111] modified bHb with PDA to synthesize morphologically homogeneous PD-HbMPs. These novel HBOCs showed antioxidant activity and enhanced Hb oxygen-binding capacity. Further, PD-HbMPs exhibited a high ability to scavenge free radical including H<sub>2</sub>O<sub>2</sub>, indicating their potential as practical HBOCs. Ethan et al. [73] proposed a photocatalytic method to synthesize the oxygen therapy agent PDA-LtEc using cell-free hemoglobin (Ec) from the PDA surface-coated worm *Lumbricus terrestris* (Lt). This method increased the oxidative protection of Ec during circulation, thereby making PDA-LtEc a promising oxygen therapy.

Recently, metal-organic frameworks (MOFs; ZIF-8) have been widely used as protective coatings for functional biomacromolecules [112, 113] (such as DNA, proteins, and enzymes). This is because of their flexible topology and favorable physicochemical properties that improve stability during storage and manipulation [114–119]. Peng et al. [74] used MOFs to encapsulate bHb and created an oxygen-carrier platform with nanoporous coatings, ZIF-8@Hb. In vitro studies showed that the neutral charge state of the surface of ZIF-8@Hb surface significantly enhanced stability in alkaline, oxidative, elevated temperature, or enzymatic environments, which is favorable for long-term blood circulation (as the study found that the blood circulation half-life (t<sub>1/2</sub>) of ZIF-8@Hb was 13.9 h, compared to 1.4 and 5.1 h for ZIF-8 and Hb, respectively). Animal studies showed that intravenous injection of ZIF-8@Hb significantly prolonged the survival time of mice in an HS model. This study provides a highly stable and long-circulating oxygen-carrier platform for Nano-HBOCs (Fig. 2).

HS is an important application scenario for Nano-HBOCs. It is worth noting that HS treatment often requires large doses, and multiple intravenous infusion of Nano-HBOCs. Therefore, the safety requirements for Nano-HBOCs are extremely high. However, most current HS studies tend to focus only on the therapeutic effect and do not adequately evaluating biocompatibility, especially in the case of multiple and massive administrations. In this regard, a research group, from Denmark, developed the concept of “stealth properties” [120] of Nano-HBOCs, which may provide new ideas for the treatment of HS using Nano-HBOCs.

### Nano-HBOCs for ischemic stroke therapy

Ischemic stroke is a clinical syndrome that refers to the disruption of blood supply in the brain caused by different cerebrovascular diseases, which eventually lead to local brain tissue ischemia, hypoxic necrosis, and corresponding neurological deficits [121]. Ischemic strokes are typically associated with varying degrees of cerebral infarcts and, in recent years, have become one of the leading causes of human mortality [122, 123] due to their increasing incidence and lethality. Ischemia and hypoxia in brain tissue cause impaired oxygen metabolism, which rapidly triggers brain tissue edema, focal neuronal cell function defects, and even necrosis [124, 125]. Following ischemia stroke, the blood flow in the tissue is typically restored within a short period, however, an excess of reactive oxygen species (ROS) and secondary inflammatory responses are generated in the body during ischemia stroke [126, 127]. These molecules cause tissue organ reperfusion injury [128], further exacerbating the ischemic stroke condition. Currently, several studies have



**Fig. 2** **A** Nanoparticle preparation, stability behavior, blood circulation, and oxygen supplying of ZIF-8@Hb are shown in figure. **B** Survival curves of mice with hemorrhagic shock following intravenous injection of PBS, Hb, ZIF-8, and ZIF-8@Hb. **C** Oxygen dissociation curves of Hb and ZIF-8@Hb. **D** Biodistribution of Hb, ZIF-8, and ZIF-8@Hb after intravenous injection for 12 h. **E** Biodistribution of Hb, ZIF-8, and ZIF-8@Hb after intravenous injection for 24 h. **A–E** Reproduced with permission [76]. Copyright 2019, American Chemical Society

indicated that early improvement in the hypoxic state of the infarct site can benefit ischemia stroke treatment [129, 130].

### Relieving cerebral infarction

HBOCs possess adjustable oxygen-carrying/-releasing capabilities and have a small particle size. These characteristics, which can enhance microvascular perfusion and collateral blood flow, lead to continuous improvement in oxygen supply to the ischemic area. As a result, they can effectively reduce cerebral infarct size [131, 132], providing a new choice for acute ischemic stroke treatment. Besides, Nano-HBOCs can inhibit glycolysis and lactic acid accumulation, which offers obvious advantages in reducing ischemic stroke injury and a significant protective effect on ischemic stroke tissue organs [133–136]. Komatsu et al. [137] established a rat middle cerebral artery occlusion (MCAO) model and an arachidonic acid (AA)-induced stroke models to evaluate the therapeutic effect of HbV. HbV treatment immediately after MCAO significantly reduced the infarct size (34.7%) in rats; the effect, was comparable to that of intravenous thrombolysis with a tissue plasminogen activator (tPA) in a thromboembolic model (34% reduction). The intravenous infusion of HbV reduced brain edema in an AA-induced thromboembolic stroke model. Kawaguchi [138, 139] et al. found that encapsulating Hb in liposomes prevented extravasation in rat stroke models; LEH significantly improved oxygen supply to the ischemic area and reduced cortical infarct size. Daiki Tomita [140]

et al. covalently combined purified bHb and HSA to design a completely new oxygen-carrier platform with a core-shell structure to minimize the vascular leakage of HBOCs and increase circulation time in vivo (possibly through the effect of net surface charge). This platform, known as HbX-HSA<sub>m</sub>, exhibits a high O<sub>2</sub> affinity (P<sub>50</sub> = 11.3 mmHg). Gekka [141] et al. covalently conjugated one Hb molecule with three human serum albumin (Hb1-HSA<sub>3</sub>) form HemoAct with a core-shell structure. In rat model of a transient MCAO (tMCAO), a significant increase in microvascular perfusion in the cortical penumbra and tissue oxygen partial pressure in the cortical penumbra were observed. Additionally, the treatment reduced liposome peroxidation, edema, and cerebral infarct size; while exerting potent neuroprotective effects in the treatment of transient ischemic injury encephalopathy via intravenous infusion. These studies suggest that improvement in the oxygen supply at the infarct site using infused Nano-HBOCs infusion effectively attenuates stroke injury.

Nano-HBOCs can be used in multifunctional composites in combination with other functional materials boost their therapeutic efficacy. Hosaka [142] et al. synthesized covalent core-shell structural protein clusters called Hb-HSA based on purified bHb and HSA. Next, platinum nanoparticles with SOD and CAT-mimetic enzyme activities were combined with HSA on the outer layer of protein clusters to form Hb-HSA<sub>3</sub> (PtNP) with antioxidant capacity. Based on the pathological microenvironment characteristic of stroke, Liu [81] et al. introduced

PDA to noncovalently engage Hb and methoxatin (M) to form PMH, whose surface-coated ROS-sensitive linker (thioacetal, termed TK linker) connects to a specific fibrin-binding peptide (CREKA, termed C-peptide), and they developed a bioinspired nanoerythrocyte (C-peptide-PRG-TK linker @PDA-Methoxatin Hb, referred to as CPTK@PMH). The half-life of free fluorescence labeled Hb was measured using a microplate reader, and the pharmacokinetic results showed that CPTK@PMH could significantly prolong Hb half-life from  $12.045 \pm 1.251$  h to about 35 h. In a murine MCAO model, CPTK@PMH was shown to target the thrombus and ischemic site, release  $O_2$  in response to hypoxic signals, regulate the metabolic microenvironment, inhibit the oxidative stress injury, and reduce the cerebral infarct size during reperfusion, which ultimately protecting neurons from acute injury and improving neurological function after cerebral ischemia (Fig. 3).

#### Inhibiting an inflammatory reaction

An inflammatory response is a key factor in the development and outcome of an ischemic stroke [143]. A series of inflammatory cascades are triggered after an ischemic stroke. Among these, microglial activation in brain tissue and neutrophil infiltration from peripheral blood into brain tissue are the hallmarks underlying microglial transformation from the M1 to M2 type. In particular, these processes inhibit neutrophil infiltration, which is a potential direction for treating cerebral ischemic injury [144]. Yin et al. [145] modified MG1 and RVG29 peptide chains based on erythrocyte-derived nanovesicles to prepare engineered nanoerythrocytes (NEMR) to promote microglial reprogramming. In an MCAO model, NEMR targeted the cerebral infarct site to attenuate tissue damage by delivering/-releasing oxygen, while inducing the overexpression of heme oxygenase-1 and CO production through the Hb metabolism. This promotes microglial transformation into the M2 type, ultimately improving therapeutic outcomes in ischemic stroke. Furthermore, the modifications of MG1 and RVG29 peptides effectively prevented NEMR uptake immune cells in the bloodstream (including mononuclear macrophages), and improved the immune escape ability of NEMR. In addition, Shimbo et al. [146, 147] found that LEH inhibited neutrophil infiltration, matrix metalloproteinase-9 (MMP-9) [148] expression, and oxygen-free radical production, reduced cerebral infarct size and edema volume, and improved neurological indices in a rat MCAO model.

Although few studies have reported the applications of Nano-HBOCs in ischemic stroke, current research confirms that Nano-HBOCs can serve as an effective therapeutic option by targeting drug delivery and easily

passing through the damaged blood–brain barrier, et al. However, stroke is a complex disease, and the unique advantages of Nano-HBOCs include their ability to relieve the ischemic and hypoxic state of brain tissues, co-load drugs, and ultimately achieve comprehensive treatment by combining thrombolysis, anti-inflammation and neuron protection. This approach also promotes rehabilitation, providing a diversified platform for treating of ischemic stroke.

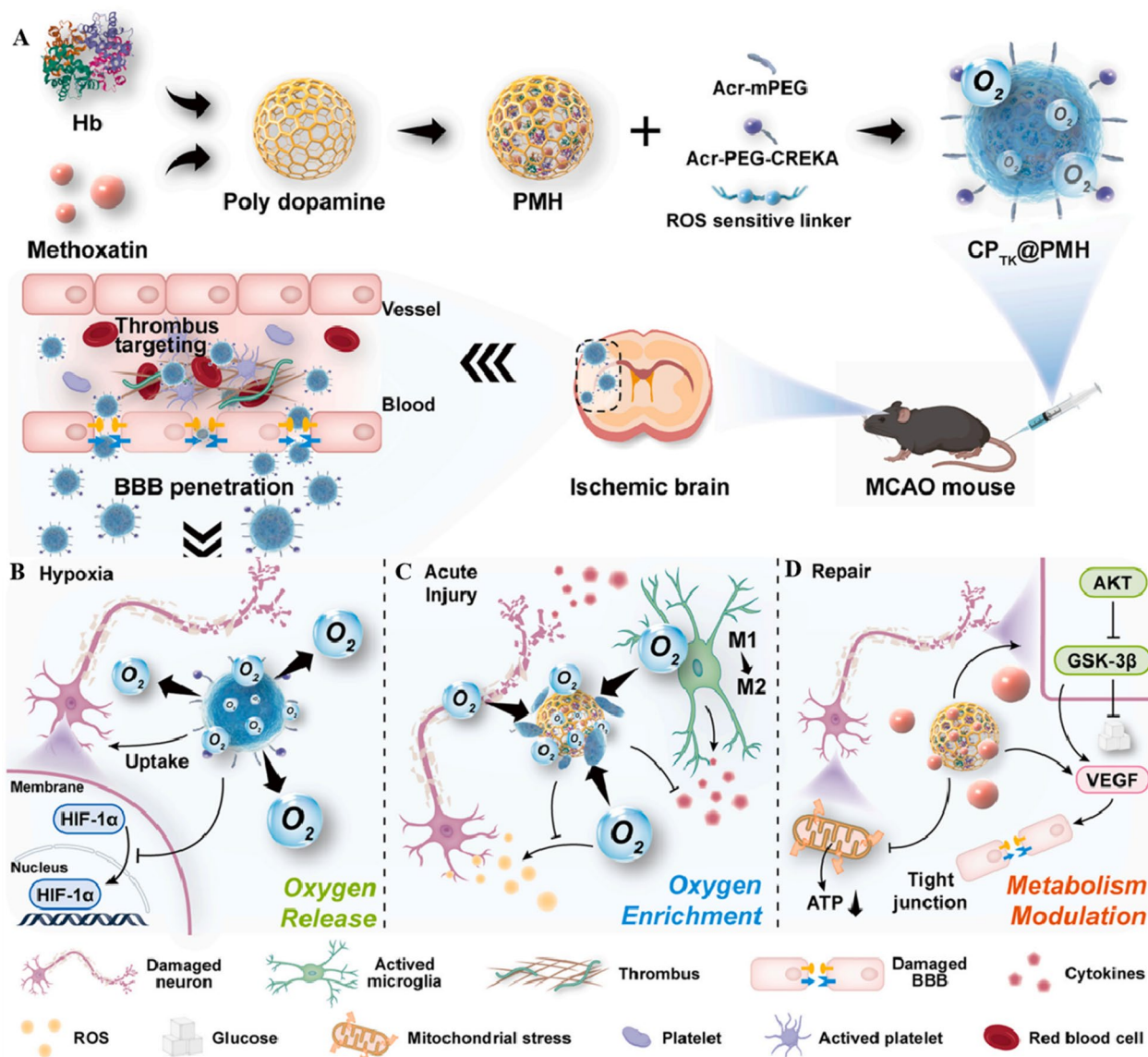
#### Nano-HBOCs for cancer therapy

Hypoxia is a common feature of most malignant tumors. Cancer cells proliferate in a disorderly manner and consume large amounts of oxygen [149, 150], which results in a tumor microenvironment (TME) [151–153] that is characterized by hypoxia accompanied by weak acids and high levels of  $H_2O_2$ , and it directly contributes to cancer cell metastasis, invasion, and angiogenesis [154, 155]. Hypoxic TME decreases the antitumor efficacy of chemotherapy, radiation therapy (RT), photodynamic therapy (PDT), and sonodynamic therapy (SDT) [156–160]. As oxygen carriers, Nano-HBOCs showed excellent stability and oxygen supply capacity, improved  $O_2$  delivery efficiency in hypoxic regions [161], alleviated the hypoxic state of tumors, and enhanced antitumor efficacy.

#### Increasing drug sensitivity

The increased expression of hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) in TME facilitates the adaptation of tumor cells to hypoxia [162, 163]. This is attributed to the development of chemical resistance that promotes the expression of p-glycoprotein (P-gp), which leads to chemotherapy failure [164, 165]. Therefore, improving the hypoxic environment of tumors is important to enhance the efficacy of chemotherapy. Kawaguchi et al. [166] reported that liposomes encapsulating hemoglobin (h-LEH) with high  $O_2$  affinity improved the oxygenation capacity of tumor cells in a mouse rectal cancer (Colon26) model. h-LEH combined with chemotherapeutic drugs by inhibiting HIF-1 $\alpha$  activity, which prevented angiogenesis and inhibited tumor growth and metastasis. Human organic cation transporter 2 (OCT2) abnormalities confer renal cell carcinoma (RCC) increased resistance to chemotherapeutic agents such as oxaliplatin (OXA) and decitabine (DAC) under hypoxic conditions [167, 168]. To tackle this issue, Chen [169] et al. developed a simple and efficient oxygen nanocarrier based on Hb (H-NPs) combined with DAC. The H-NPs increased the oxygen content of the RCC tissue, alleviated hypoxia-induced loss of DAC activity, and enhanced the sensitivity of RCC cells to the combined treatment with DAC and OXA.





**Fig. 3** **A** Illustration of the CPTK@PMH nanoerythrocyte formation and metabolic microenvironment modulation in ischemic brain: nanoerythrocyte accumulation in ischemic core via microthrombus binding and uptake by neurovascular unit after blood brain barrier penetration; **B** hypoxia-responsive oxygen release to relieve necroptosis; **C** oxygen balance regulation to alleviate acute reperfusion injury via oxygen enrichment, ROS scavenging and microglia polarization; **D** repair promotion achieved by metabolic microenvironment modulation via energy and glucose metabolism activation and blood brain barrier protection. **A–D** Reproduced with permission [83]. Copyright 2019, Nano Today.

**Targeted drug releasing**

A hypoxic environment causes the overexpression of glucose transporter isoform 1 (GLUT1) [170] on the surface of cancer cells, and nanocarriers modified with glucose can identify cancer cells via GLUT1. Thus, Bu et al. [171] designed Nano-HBOC with both oxygen-carrying capacity and cancer cell-specific recognition. The synthesis of the triblock copolymer poly [2-(methacrylamido) glucopyranose]-b-poly (methacrylic acid)-b-poly (butyl methacrylate) (PMAG-b-PMAA-b-PBMA) micelles indicated that Hb was bound to the

micelles via a condensation reaction to form Hb-conjugated micelles (HCM). PMAG-linked GLUT1 specifically recognizes cancer cells, and HCM can bind O<sub>2</sub>. Meanwhile, the uptake of DOX-loaded HCM by tumor cells was increased in a model of human cervical cancer cells (HeLa), and Nano-HBOCs were confirmed to target cancer cell recognition and reduce tumor drug resistance, ultimately improving therapeutic efficacy. Tian et al. [83] designed and constructed camouflaged biomimetic nanocomposites (DHCNPnPs) based on PLGA encapsulated with Hb and DOX. They modified



the surface of the cancer cell membrane to realize homologous recognition. The DHCNPNPs exhibited strong self-recognition ability, affinity for homologous cancer cells in a model of human breast cancer cells (MCF-7 cells), and minimal toxicity to normal tissues. DHCNPNPs achieved  $O_2$ -independent conditions and high target selectivity in the TME by inhibiting HIF-1 $\alpha$ , down-regulating the expression of multidrug resistance gene 1 (MDR1) and P-gp, and increasing chemosensitivity, which improved the overall therapeutic efficacy (Fig. 4). Yang et al. [172] designed a multifunctional liposome, DOX-Hb-lipo (DHL), which was simultaneously loaded with human hemoglobin (hHb) and DOX. They showed that DHL could precisely release DOX and deliver  $O_2$  to specific hypoxic sites, thereby alleviating hypoxia-induced chemoresistance. DHL inhibited tumor growth in mouse breast cancer models (4T1) and mouse colon cancer models (CT-26), achieving in situ  $O_2$  release and relieving the hypoxic state of the TME, thereby providing an effective alternative for cancer treatment. Wang et al. [84] engineered an endogenous Nano-HBOCs called V(Hb)@DOX, which self-assembled resulting DOX-loaded biomimetic into hollow nanovesicles. In the acidic intertumoral environment, V(Hb)@DOX targeted M2-type macrophages and released DOX, thereby effectively suppressing of tumor growth and metastasis. V(Hb)@DOX prolonged the survival of mice bearing 4T1 orthotopic breast tumors and achieved long-lasting specific immunity in a murine colon cancer CT26 model.

#### Promoting ROS generation

ROS are derived from  $O_2$  and are formed through oxidative reactions or electron excitation, they mainly consist of hydroxyl radicals ( $\bullet OH$ ), superoxide anion ( $\bullet O_2^-$ ) and singlet oxygen ( $^1O_2$ ) [173]. When the levels of ROS exceed a certain threshold, they can disrupt the cellular structure and thus be used to inhibit tumor growth [174]. Nano-HBOCs can promote ROS production in three ways. Firstly, Nano-HBOCs can provide sufficient  $O_2$  as a feedstock for ROS production [175], while improving the TME. Furthermore, in the presence of ferrous ions (derived from Hb), high levels of  $H_2O_2$  in the TME undergo a Fenton and Haber-Weiss reaction, which continuously produce highly toxic  $\bullet OH$  [176]. Additionally, Hb can mimic horseradish peroxidase (HRP), which also produce  $\bullet OH$  from  $H_2O_2$  [177].

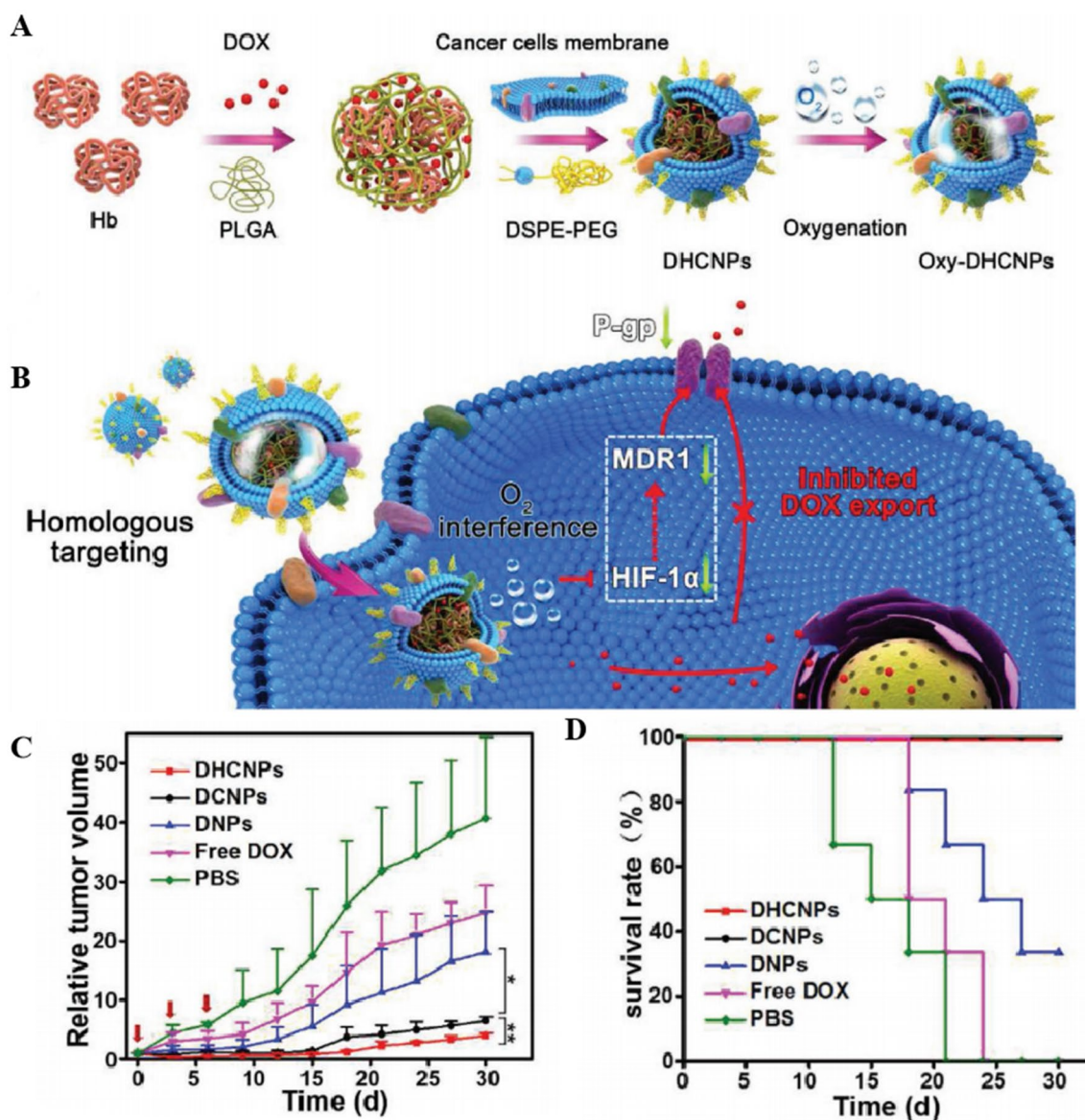
#### Application of Nano-HBOCs in RT

RT, a common method for cancer treatment, damages DNA using X-rays and generates ROS to inhibit tumor growth; its therapeutic effect is directly related to the  $O_2$

content in the TME [178, 179]. Sang et al. [180] developed an oxygen-enriched X-ray nanoprocessor (Hb@Hf-Ce6 NPs) based on metal-phenolic coordination to modulate the oxygen balance in the TME and reverse immunosuppression for cancer eradication and metastasis inhibition. In the nanoprocessor, the radiosensitizer hafnium (Hf) coordinated with chlorin e6 (Ce6) nanocomplexes and encapsulated Hb to deliver  $O_2$ . Under X-ray irradiation, Hf emitted radioluminescence to activate Ce6 for ROS generation, and Hb@Hf-Ce6 NPs released  $O_2$  at the tumor site. This promoted T cell-mediated systemic immunity to exert long-range antitumor effects on tumors and enhance RT efficacy, thereby exhibiting superior antitumor efficacy in a bilateral tumor model and a lung metastatic model of 4T1 breast cancer. (Fig. 5). Xia et al. [85] combined a Hb oxygen carrier with gold nanoparticles (Au) radiosensitizer to prepare Au-Hb NPs. Inspired by the fact that P-selectin on the surface of platelets (PLT) can specifically bind to CD44 on the surface of cancer cells [181], platelet-coated oxygen-carrying nanoparticles (Au-Hb@PLT NPs) were synthesized by coating Au-Hb NPs with PLT membrane. Au-Hb@PLT NPs could deliver sufficient  $O_2$  deep into the tumor tissue, relieve hypoxia, and improve the effect of RT. Au acts as a sensitizer, which enhances the sensitivity of tumor cells to X-rays and their therapeutic effects, showing good tumor-therapeutic efficiency and a few side effects in the HeLa cell model. Of note, a clear boundary between the normal parenchyma and dead cancer cells was confirmed using the hematoxylin and eosin staining (H&E), suggesting that Au-Hb@PLT + RT (2 Gy) killed tumor cells while causing relatively less damage to healthy tissue.

#### Application of Nano-HBOCs in PDT

Photodynamic therapy (PDT), a clinically recognized and minimally invasive tumor treatment, has become an alternative to chemotherapy [182, 183]. PDT includes a photosensitizer (PS), a light source, and sufficient  $O_2$ . Under PS catalysis,  $O_2$  is activated to ROS, such as  $\bullet OH$  and  $^1O_2$ , which in turn induce tumor vascular injury, thereby leading to tumor cell death [184–186]. However, the hypoxic state of the TME impairs PDT efficacy [187], which can be effectively reversed by increasing the  $O_2$  content of the tumor [188, 189]. Wang et al. [190] developed a new PDT therapy based on the concept of  $O_2$  “self-compensation ability,” which was synthesized by chemically conjugating Hb with polymeric micelles formed by triblock copolymers of poly (ethylene glycol)-block-poly (acrylic acid)-block-polystyrene (PEG-b-PAA-b-PS). They used Hb as a PS zinc phthalocyanine (ZnPc) carrier, which endowed the Hb-conjugated ZnPc PS carrier with good oxygen-binding capacity, antioxidant activity, and the ability to produce abundant  $^1O_2$ , thereby

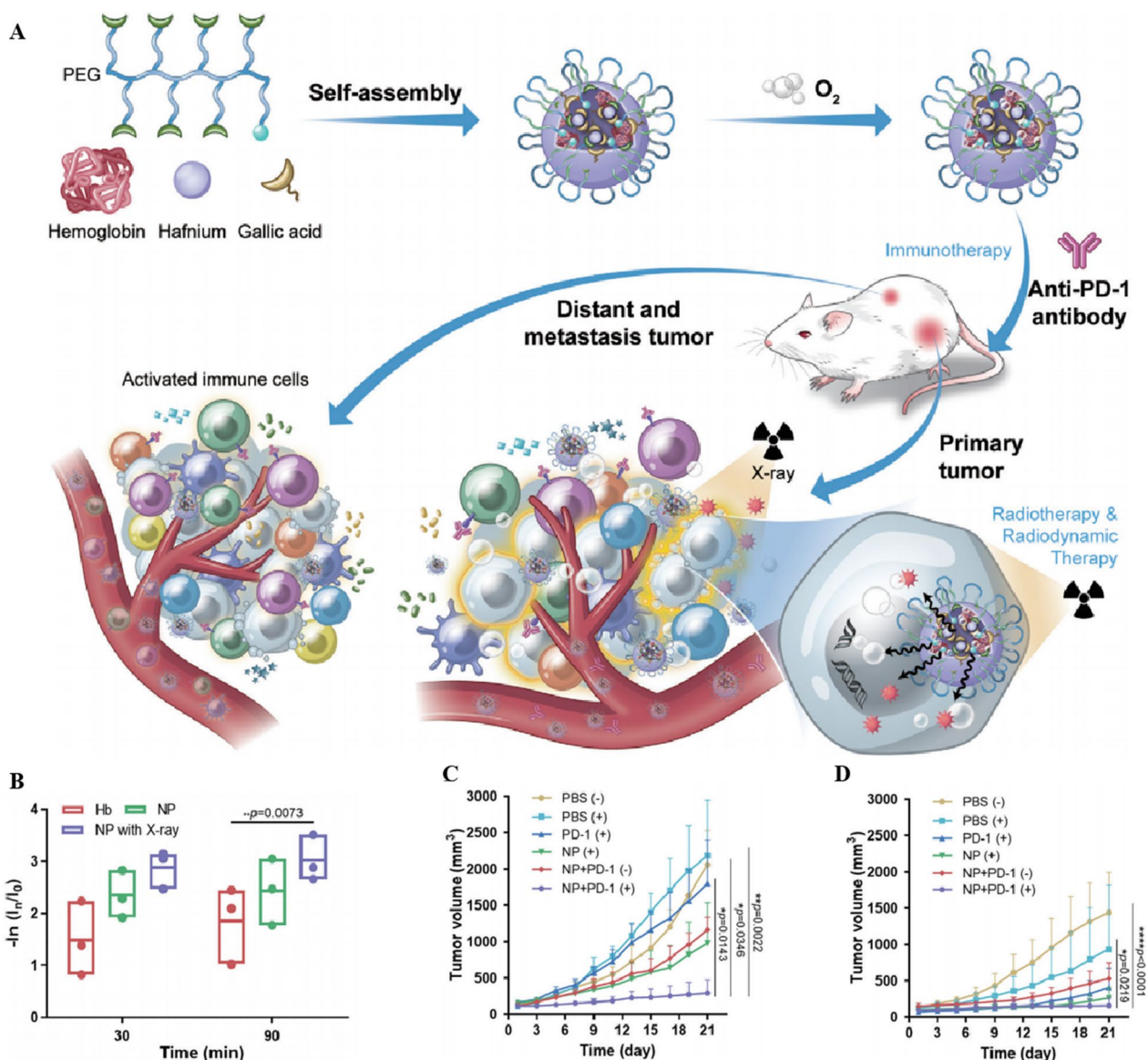


**Fig. 4** The design and function of DOX/Hb loaded PLGA-cancer cell membrane nanoparticles (DHCNPs) for homologous targeting and O<sub>2</sub> interference. **A** Synthesis of oxy-DHCNPs. DHCNPs were prepared by extrusion with preformed DOX/Hb-PLGA NPs, DSPE-PEG, and MCF-7 cancer cell membrane, and then were oxygenated to obtain oxy-DHCNPs. **B** Cellular functions of DHCNPs, including homologous targeting, downregulation of predictive markers (HIF-1 $\alpha$ , MDR1, and P-gp), and inhibited DOX export. **(C)**: MCF-7 tumor growth curves of different treated groups (scale bar 50  $\mu$ m). **(D)**: Survival rates of tumor-bearing mice in various groups. **A–D** Reproduced with permission [85]. Copyright 2017, Advanced Functional Materials.

exhibiting robust photocytotoxicity in an in vitro HeLa cell assay.

To address the low O<sub>2</sub> levels in tumor and enhance PDT efficacy, Tang et al. [76] used ferritin, a protein-based nanocage, to tether a large amount of PS (such as ZnF16Pc) to the surface of RBCs. They utilized Hb as a PS carrier to provide O<sub>2</sub>, loaded ZnF16Pc at ferritin molar ratio (1:40) to form P-FRT, and simultaneously produced PFRT-RBCS nanoparticles by coupling

RBCs. PFRT-RBCS prolonged the circulation half-life of Ps, which were labelled with IRDye800 that exhibited a high fluorescence in the blood ( $7.6 \pm 2.1\%ID\ g^{-1}$  at 1 h), and the signal remained strong for 24 h ( $3.2 \pm 0.4\%ID\ g^{-1}$  at 24 h). P-FRT-RBCs achieved continuous oxygen supply under hypoxia in the TME, thereby enhancing the ability of PDT to kill cancer cells in a U87MG tumor-bearing mouse model (human brain astroblastoma cells). Luo et al. [191] developed a biomimetic lipid-polymer



**Fig. 5** **A** Schematic illustration showed the Hb@Hf-Ce6 nanoparticles-mediated X-ray induced radiotherapy-radiodynamic therapy-immunotherapy for the eradication of both primary and distant tumors. Hb was encapsulated in the Hf-phenolic coordination nanopatform for oxygen delivery through self-assembly. **B** The oxygen release behavior of hemoglobin (Hb) and Hb@Hf-Ce6 NPs (NP) with/without X-ray irradiation was evaluated by  $Ru(bpy)_3Cl_2$  probe. Hb and NP were oxygenated previously. **C** Tumor volume growth curves for primary tumors. **D** Tumor volume growth curves for distant tumors. **A–D** Reproduced with permission [184]. Copyright 2021, Advanced Science.

nanoparticle loaded with PS (indocyanine green, ICG) and oxygen carrier (Hb). Nanoscale artificial RBCs (I-ARCs) were prepared by encapsulating Hb/ICG complexes in a PEG-modified lipid layer. In an MCF-7 human breast cancer model, the I-ARC provided sufficient  $O_2$ , and ICG in the I-ARC efficiently converted  $O_2$  to ROS under near-infrared (NIR) laser irradiation, thereby triggering sustained damage to tumor cells. Furthermore, I-ARC serves as an ideal fluorescence imaging probe to dynamically monitor the biodistribution of nanoparticles

and oxygen levels. The dynamic PDT process can be self-monitored through ICG, while serving as a diagnostic platform for self-monitoring therapy integration. Xu et al. [71] prepared a novel hemoglobin-polymer conjugate (HbTcMs) with high biocompatibility and stability as a carrier for both  $O_2$  and PS to provide additional  $O_2$  to enhance PDT efficacy. Compared with free Hb, HbTcMs not only retained the Hb oxygen-binding capacity but also exhibited higher stability against oxidation and trypsin digestion. In a 4T1 cell model, HbTcMs produced



numerous oxygen-derived  $^1\text{O}_2$  molecules and exerted enhanced phototoxicity by utilizing the oxygen supply ability of Hb. Cao et al. [192] constructed the BP@RB-Hb nanoparticles by assembling Hb into nanocomplexes with novel two-photon species: bis(pyrene) (BP) and traditional PS (RB). Hb acts as an  $\text{O}_2$  donor to provide additional  $\text{O}_2$  resistance to the TME at the tumor site through NP targeting. Simultaneously, the resonance energy transfer effect (FRET) inside the nanoparticles indirectly stimulated PS under irradiation with a two-photon laser, thereby improving the treatment depth. In a tumor-bearing mouse model, the material showed excellent oxygen-carrying performance and penetration depth, greatly enhancing PDT efficiency. In tumor-bearing mouse models, BP@RB-Hb nanoparticles exhibited excellent penetration depth and oxygen-carrying properties, which greatly improved PDT treatment efficiency. Chen et al. [193] proposed a protein hybridization method for combining HSA and Hb to develop a bioinspired hybrid protein oxygen nanocarrier (C@HPOC) loaded with photosensitizer e6 (Ce6), which overcomes tumor hypoxia via co-delivery of tumor-targeted  $\text{O}_2$  and PS and improves PDT efficacy by generating a large amount of  $^1\text{O}_2$ . C@HPOC stimulated robust systemic antitumor immunity and elicited potent antitumor metastatic and abscopal effects in a murine triple-negative breast cancer (4T1-mTNBC) model [194] (Fig. 6). Tian et al. [195] connected Hb to Ce6 to load sorafenib (SRF, an iron death promoter) and built a nanoplatfrom SRF@Hb-Ce6. Hb not only provides  $\text{O}_2$  and increases PDT efficacy, but it also synergizes with SPF, significantly increases the generation of lipid peroxides, downregulates GPX4 expression, and increases tumor cell iron death [196]. This platform shows promise as a nanoplatfrom for combination cancer therapy.

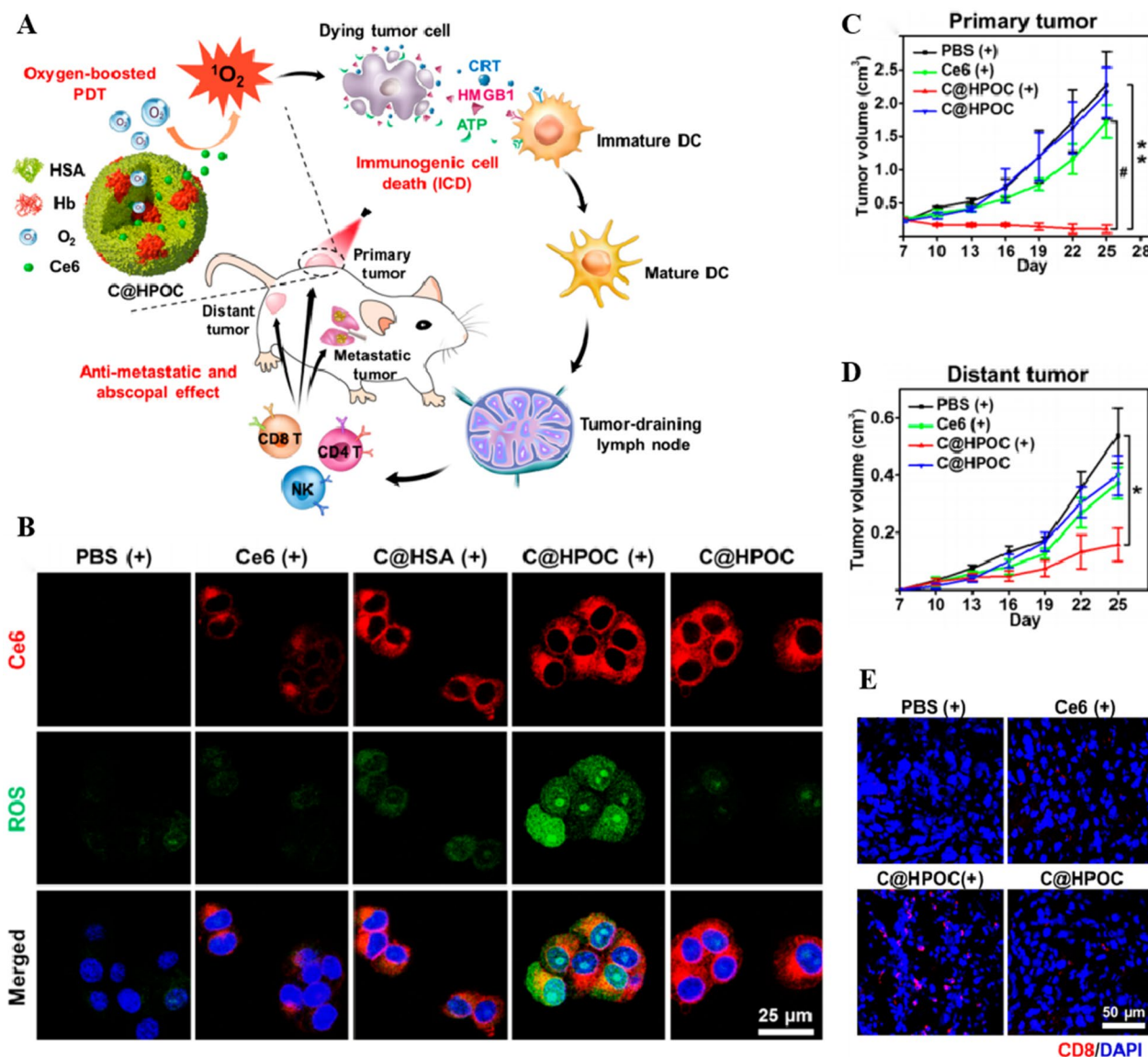
#### **Application of Nano-HBOCs in SDT**

SDT is an emerging cancer therapy that triggers ROS generation from abundant  $^1\text{O}_2$  using ultrasound (US), which leads to oxidative damage in tumor cells and enhanced therapeutic efficacy [197]. Its advantages include superior tissue penetration ability, minimal damage to surrounding healthy tissues, and lower skin sensitivity. The process of ROS generation using US irradiation of sonosensitizers during SDT requires the participation of  $\text{O}_2$ , which is similarly limited by hypoxia in the TME [198]. In this context, Yin et al. [199] designed a novel intensified oxygen supply sonosensitizer system (MnPcS@HPO) by reconstituting bHb and HSA through disulfide bonds and loading the sonosensitizer Mn-phthalocyanine (MnPcS). This system combines the oxygen-carrying capacity of Hb and the tumor-targeting properties of HSA. MnPcS@HPO was enriched in the hypoxic area of the tumor,

effectively alleviating the hypoxic state of the tumor. Simultaneously, MnPcS produces a large amount of  $^1\text{O}_2$  under US irradiation, effectively inhibiting tumor growth. Yuan et al. [72] constructed  $\text{O}_2$ @Hb@ZIF-8 (OHZ) nanoparticles formed from ZIF-8 encapsulated Hb, which not only provided abundant  $\text{O}_2$  for US-induced ROS generation, but also showed excellent biocompatibility, as evidenced by cell viability evaluation and imagine (H&E) of major organs. ZIF-8 was used as a drug carrier for improving the Hb packaging rate, achieving pH-responsive Hb/ $\text{O}_2$  release, generating a large amount of  $\text{O}_2$  in the acidic TME, and relieving tumor hypoxia, thereby providing a source of  $\text{O}_2$  for US-triggered ROS generation. In 4T1 tumor-bearing mice, OHZ nanoparticles could not only inhibit the growth of subcutaneous tumors but also have an excellent inhibitory effect on the growth of deep-seated tumors, which helped achieve the SDT treatment of tumors at different depths. Pan et al. [200] synthesized ZIF-90 in water under mild conditions to encapsulate bHb and prepare an oxygen carrier, ZIF-90@Hb. The dynamics of oxygen consumption showed that the initial current of ZIF-90@Hb ranged from 9.15  $\mu\text{A}$  to 4.18  $\mu\text{A}$ , while that of free Hb was 2.78  $\mu\text{A}$  and dropped to 0.55  $\mu\text{A}$  within 100 s. This observation indicated that ZIF-90@Hb demonstrated better oxygen carrying/releasing abilities than free Hb. Furthermore, Cun et al. [75] similarly combined bHb and Au nanoparticles to synthesize ultrasmall protein metal clusters (MNCs) to develop a novel HBOC. They combined the complexes of Hb@AuNCs incorporated into the reported MOFs, which increased their antioxidant capacity to inhibit the formation of methemoglobin (the fluorescence intensity of hydrogen peroxide decreased sharply by 70%, with a remarkable 39% decrease in methemoglobin). Overall, these studies suggest that the utilization of the ZIF metal framework in oxygen carriers [74] provides a multifunctional platform for the development of blood substitutes in the near future. Liang et al. [201] encapsulated PtIV, a platinum (Pt) prodrug, with Hb to prepare a multimodal nanoparticle for tumor-targeted US radiation-triggered cancer therapy, harnessing the good solubility of Hb as a sonosensitizer. These nanoparticles can treat large volumes of deeply located tumors and provide new ideas for cancer treatment.

Hypoxia is a hallmark of TME in solid cancers [202]. It not only promotes tumor growth, but also reduces the effectiveness of treatment. Nano-HBOCs have been reported to have the ability to enhance synergistic therapy with stable  $\text{O}_2$  supply and can be loaded with drugs targeting tumor cells. Numerous studies have investigated the use of Nano-HBOCs in tumor therapy, and significant therapeutic effects have been observed in vitro and in vivo. By combining Nano-HBOCs with drugs





**Fig. 6** **A** Schematic depiction of oxygen-augmented immunogenic PDT with C@HPOC for eliciting the anti-metastatic and abscopal effect. Human serum albumin (HSA) was hybridized with oxygen carrying hemoglobin (Hb) via intermolecular disulfide bonds to form a hybrid protein oxygen nanocarrier with Ce6 loaded (C@HPOC). Under laser irradiation, oxygen self-supplied nanoparticles (C@HPOC) elevated the generation of cytotoxic  $^1O_2$  and moreover triggered immunogenic cell death (ICD). C@HPOC-mediated PDT not only destroyed the primary tumors but also inhibited the distant tumors and lung metastasis by systemic anti-tumor immune responses. **B** Confocal images of cellular uptake and ROS generation in 4T1 tumor cells. **C** Growth curves of primary tumor on mice after various treatments. **D** Growth curves of distant tumor on mice in different treated groups. **E** Immunofluorescence staining detection of CD8 T cells (red) in tumor tissues. **A–E** Reproduced with permission [197]. Copyright 2018, American Chemical Society Nanomaterials.

and enhancing their abilities, we can potentially make a breakthrough in the clinical treatment of tumor.

**Nano-HBOCs for wound healing**

O<sub>2</sub> plays a key role in wound healing [203] by promoting cell proliferation, accelerating angiogenesis, reducing infection, and increasing collagen synthesis [204, 205]. The oxygenation level in the wound microenvironment

is a key rate-limiting factor for wound healing [206, 207]. Long-term hypoxic conditions lead to impaired neovascularization and limited wound healing [208], whereas a hypoxic environment promotes wound healing [209]. Therefore, in situ oxygen production, oxygen carriers, and various strategies to enhance oxygen supply have been used to relieve the hypoxic state of wounds,

promote angiogenesis, and enhance collagen remodeling, ultimately accelerating wound healing [210–213].

### Reducing oxidative stress

Diabetes mellitus (DM) is a chronic disease that affects millions of humans. Further, in more than 25% of patients with DM, factors such as uncontrollable bacterial infections [214], persistent inflammation [215], and hypoxia [216] can cause foot ulcers, which, in severe cases, can lead to amputation [217, 218]. According to previous studies, the long-term exposure of diabetic patients to a high glucose environment can functionally impair the of HIF-1 $\alpha$ / VEGF axis, resulting in reduced abilities to reverse and regulate the ischemic hypoxic state of the wound tissue [207, 219] to promote wound healing. Fukui [220] et al. suggested that LEH has the advantages of small particle size and high oxygen affinity, effectively perfusing obstructed blood vessels and improving the oxygen supply to the wound. Studies based on a diabetic mouse (dB/ dB) skin defect model have shown that h-LEH significantly increased the oxygen supply level of the wound microenvironment and inhibited the inflammatory cascade [221] while effectively improving the microcirculation and accelerating diabetic wound healing. MXene is a two-dimensional material with ultrathin layer topology and good biocompatibility [222]. Li et al. [79] used a system of H<sub>2</sub>O<sub>2</sub>/HbO<sub>2</sub> to catalyze the oxidative coupling of dopamine-grafted hyaluronic acid and PDA with encapsulated Ti<sub>3</sub>C<sub>2</sub>MXene nanosheets to form injectable hydrogels (Ti<sub>3</sub>C<sub>2</sub>@PDA or MXene@PDA NSs). HbO<sub>2</sub> exert HRP-like enzyme activity for catalyzing gel formation, whereas MXene scavenge excess free radicals, reduce oxidative damage, and maintain oxidative homeostasis while exerting a bacteriostatic effect [223]. Further, MXene and PDA have excellent photothermal conversion features that can generate heat to elevate temperature under 808 nm NIR irradiation, thereby achieving a Hb-reversible oxygen release in the wound microenvironment. In the dB/dB wound model, the hydrogel oxygen carrier continuously relieved the hypoxic state of the wound tissue, reduced oxidative stress, regulated macrophage polarization, promoted angiogenesis, and accelerated the healing of diabetic wounds.

### Promoting tissue repair

O<sub>2</sub> promotes tissue formation and repair, thereby accelerating wound healing [224, 225]. Currently, the development of miniature, universal, and customizable O<sub>2</sub> carries is progressing rapidly [226, 227] and holds great potential for applications in the field of wound healing. Liu et al. [228] combined molybdenum disulfide quantum dot-blended gelatin methacryloyl (GelMa) to produce inverse opal microparticles as microcarriers for

preparing NIR-triggered porous controllable oxygen carriers by coupling amide bonds with hHb for tissue repair. Based on a typical abdominal wall defect rat model, this oxygen carrier was found to improve the oxygen supply and play a role in supporting cell growth, thereby stimulating extracellular matrix secretion, promoting collagen and angiogenesis, accelerating granulation tissue formation, and promoting the repair of abdominal wall defects. The characteristics of oxygen carriers in tissue repair render them useful for wound healing and tissue engineering (Fig. 7).

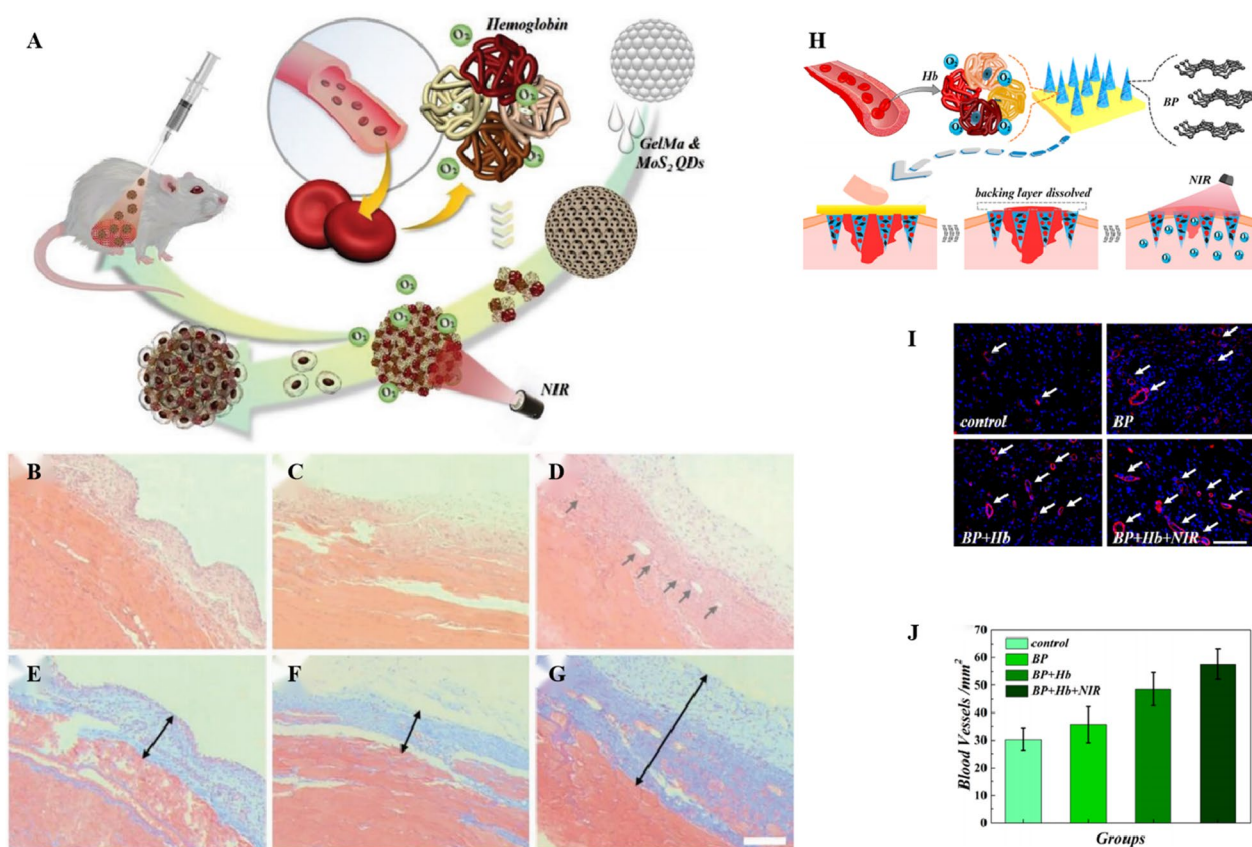
Zhang et al. [86] constructed responsive microneedles (MNs) simultaneously loaded with black phosphorus (BP) and hHb to overcome the limitation that most oxygen carriers can only contact the surface area of the wound without reaching the inner tissue. These responsive MNs could penetrate the epidermal layer to the interior of wound tissue in a painless, noninvasive, and sterile manner [229, 230], thereby achieving controllable delivery of O<sub>2</sub> and promoting wound healing. Further, a NIR controllable O<sub>2</sub> releasing carrier was fabricated using GelMA as a synthetic material for responsive MNS, which helped improve the mechanical strength and penetration ability of MNs loaded with quantum dots of BP and Hb. In a full-thickness cutaneous wound type I diabetes rat model, the treatment group exhibited the smallest wound area, narrowest wound bed, thickest regenerated epithelial tissue, and densest neovascular state, indicating excellent healing capacity compared to the control groups. These results suggest that this responsive MN oxygen carrier proposed by this research group has an ideal therapeutic effect on wound healing.

Currently, there are only a limited number of studies on the application of Nano-HBOCs to wound healing, and the main forms of drug delivery primarily involve hydrogels and microneedles. By expanding the options for administration, such as through spraying, we believe that Nano-HBOCs will be able to better meet the clinical requirements of wound healing and become an effective therapeutic method.

### Nano-HBOCs for other biomedical applications

Nano-HBOCs have also been used in other disease therapies, as discussed below.

Port-wine staining (PWS) is a non-neoplastic malformation caused by telangiectasia at birth [231] that mostly occurs in the head and neck region and severely affects patient penetrance and self-confidence [232, 233]. PWS grows modularly, tends to bleed [234], and forms fibrous scars on the skin surface, which become more pronounced after puberty [235]. Currently, PDT is a primary treatment strategy for PWS [236]. HbVs are smaller (250 nm) than RBCs and 1/40 the diameter of



**Fig. 7** **A** The schematic illustration of the light-responsive MoS<sub>2</sub> QDs integrated inverse opal microcarriers for controllable oxygen delivery and tissue repair. The H&E staining of **B–D** repaired samples after implantation for 2 weeks. Vessels in the samples are indicated with grey arrowheads. **B** Control, **C** Experimental I, **D** Experimental II. The Masson staining of **E–G** repaired samples after implantation for 2 weeks. Granulation tissue thickness in the samples are indicated with black arrowheads. **E** Control, **F** Experimental I, **G** Experimental II. **B–G** The scale bar is 200  $\mu\text{m}$ . **A–G** Reproduced with permission [233]. [88]. Copyright 2020, American Chemical Society Nanomaterials. Copyright 2019, small. **H** Schematic illustrations of wound healing using NIR responsive separable MNs which encapsulate BP QDs and oxygen-carrying Hb. **I** Corresponding double immunofluorescent staining of CD31 and  $\alpha$ -SMA on day 9. The arrows indicate the vascular ducts. The scale bars are 100  $\mu\text{m}$ . **J** Corresponding quantitative analysis of the blood vessel density on day 9. The scale bars are 50  $\mu\text{m}$ . **H–J** Reproduced with permission

RBCs; further, they are inclined to flow in the marginal zone of microvessels in the blood [237] and increase the hemoglobin concentration in microvessels, enabling a more efficient conversion of light energy into heat energy. Rikihisa et al. [238] applied a laser as a treatment for PS in PWS (at a wavelength of 595 nm) using the oxygen-carrying capacity of HbVs (or LEHs). An oxygen-rich environment enhanced the PDT effect, generated a large amount of ROS, damaged the vessel wall, treated PWS, and achieved a better therapeutic effect in an animal model with chicken wattle as PWS.

Pre-eclampsia (PE) can lead to pre-term birth in pregnant women and is a leading cause of maternal and fetal death [239–241]. Studies have shown that inadequate trophoblast invasion in the placenta leads to the narrowing of the spiral arteries of the placenta, thereby causing placental ischemia/hypoxia and limiting gas and nutrient exchange between the mother and fetus, which ultimately

causes intrauterine growth restriction, maternal hypertension, and organ damage [242–246]. HbV, which is a type of Nano-HBOC with a nanoscale size and no blood group antigens, can pass through narrow placental spiral arteries and capillaries to provide an efficient O<sub>2</sub> supply to the placenta [247, 248]. Ohta et al. [249] intravenously injected HbV into a rat preeclampsia model. The results showed that HbV alleviated fetal hypoxia by supplying oxygen while improving fetal growth restriction in a murine model of preeclampsia pregnancy. This study suggests that HbV is an effective treatment modality for fetal hypoxia caused by placental dysfunction during pregnancy.

*Porphyromonas gingivalis*, a gram-negative anaerobic bacterium, is one of the main pathogens causing oral infections (including gingivitis, periodontitis, and oral ulcers) [250]. Traditional broad-spectrum antibiotic therapies often lack selectivity and can lead to oral



flora dysbiosis [251]. Previous studies have shown that *P. gingivalis* can bind to and acquire porphyrin molecules from Hb; therefore, Hb can be used as a specific drug delivery vehicle to treat *P. gingivalis* infection [252]. Based on this idea, Bai et al. [253] complexed rat oxyhemoglobin (oxyHb) with an anionic amphiphilic PS (IR820) to produce a stable nano-photosensitizer oxyHb@IR820. oxyHb@IR820 targets *P. gingivalis*, mediating the synergistic antibacterial effects of photothermal therapy and PDT [254–256] and providing a new therapeutic strategy against a *P. gingivalis* infection. In an oral infection model of *P. gingivalis* in rats, oxyHb@IR820 supplied exogenous O<sub>2</sub> and addressed the limitation caused by the poor effectiveness of PDT on anaerobic bacterial infection in hypoxic environments. Meanwhile, the specific uptake of *P. gingivalis* oxyHb@IR820, which improved the photodynamic and photothermal conversion efficiencies, significantly inhibited bacterial growth and reduced the oral ulcer area.

Further, Nano-HBOCs showed great application prospects in other tissue engineering scenarios. For example, Paciello et al. [257] developed a novel smart cyclic oxygen-releasing biomaterial called G-HbOD, which demonstrated superior biocompatibility and oxygen supply capability for maintaining cell viability under hypoxia in 3D cell culture tests.

Vasosuppression [258] and oxidative stress [259] resulting from hypoxia are major obstacles to bioartificial islet (BAP) transplantation [260]. To address this issue in human islet allotransplantation, Mouré et al. [261], encapsulated marine worms (HEMOXCell) Hb [262], via alginate and silicone calcium peroxide, creating a compound that produced, carried, and released O<sub>2</sub>, increasing the O<sub>2</sub> concentration in BPA. This strategy effectively addressed the lack of O<sub>2</sub> within the graft in a closed environment, and promoting neovascularization. This novel technology provides innovative solutions for tissue and bioengineering O<sub>2</sub> supply. Enzyme biofuel cells (EBFC) are widely used as implanted biomedical devices in cardiac pacemakers [263], electrical sensors [264], and biosensors for physiological parameters [265]. However, the current catalysts for EBFC have very low catalytic performance and poor adhesion on the electrode surface. Chen et al. [266] anchored Hb on exogenous Ca<sup>2+</sup> and PO<sub>3-4</sub> to generate hydroxyapatite (Ca<sub>10</sub>(PO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub>, HAP) nanodots and reported a novel EBFC constructed from nanoengineered RBCs (NERBCs). Given its strong oxygen adsorption capacity, NERBCs exhibits excellent catalytic reduction capacity, biocompatibility, high selectivity, and lifespan when used as an EBFC-negative catalyst, thereby pointing to a new direction for fabricating bio-nanobatteries.

## Conclusions and perspectives

O<sub>2</sub> plays a vital role in life activities, and Hb is a natural carrier of oxygen in the body. The timely transfusion of RBCs can effectively restore blood volume and maintain tissue oxygen supply in both clinical and military treatment scenarios. HBOCs have become the main focus for artificial oxygen carriers due to their advantages of being closer to natural Hb and providing an effective method to solve the inherent problems of natural blood transfusion. The rapid development of nanotechnology and nanomaterials has opened new possibilities for HBOCs.

Nano-HBOCs are a new type of hemoglobin-based oxygen carriers developed using nanotechnology combined with nanomaterials. This combination, has helped improve the application scenarios of Hb and has shown certain advantages in enhancing the stability of the molecular structure of Hb, reducing vasoactivity, prolonging half-life, and improving biocompatibility. Furthermore, Nano-HBOCs have broad application prospects (Table 2), and have attracted increasing attention from the medical, material, chemical, biological, and engineering research attention. Despite the rapid development of Nano-HBOCs in the field of biomedical research, there are still several limitations associated Nano-HBOCs, which are listed below: Nano-HBOCs require complicated preparation processes and are associated with prohibitive costs. The development of simple, safe, and cost-effective preparation methods will be the focus of future studies.

- 1) Constructing Nano-HBOCs with characteristics and functions similar to those of natural RBCs is difficult; more in-depth systematic research and technological progress are required to achieve a complete biomimetic replacement of natural RBCs.
- 2) Clinical transformation and applications are significantly challenging. Currently, the vast majority of Nano-HBOCs are in the laboratory research stage, and relevant clinical research is yet to be conducted. Therefore, it is necessary to conduct strict and systematic safety and perform effective evaluations for diverse application scenarios to meet the regulatory requirements.
- 3) Non-functional methemoglobin poses a significant challenge in the development of Nano-HBOCs. However, existing studies have not sufficiently addressed the control of Hb oxidation and lacked molecular characterization. In our review, we emphasize the need to pay attention to methemoglobin production. Furthermore, we propose various approaches to address this issue, including low-temperature operation, reducing the reaction time, adding antioxidants, and employing carbon-bonding techniques



**Table 2** The key points of Nano-HBOCs, including approaches to fabrication, targeted fields and references

Key Points	Categories	Approaches	Targeted Fields	References
Liposome-encapsulated	TRM-645	PEG modify liposomes which mimicking the RBCs membrane	Hemorrhagic shock	[69]
	Erythromer (EM)	A novel shuttle, which adjust P <sub>50</sub> in response to pH changes		[107]
	HbPs	Hb encapsulated mPEG-PLGA through double emulsion technique		[70]
	LHE/HbV	Hb encapsulated by different type of vesicles	Hemorrhagic shock Ischemic stroke Cancer Wound healing PWS PE	[104, 137, 166, 220, 249, 267]
PDA surface-coated	Hb-PDA	Using a template-based co-precipitation technique	Hemorrhagic shock	[77]
	PDA-LtEc	A photocatalytic method		[73]
	CPTK@PMH	Engaging Hb and M to form PMH, connecting to a specific fibrin-binding peptide	Ischemic stroke	[81]
	MXene@PDA	Hyaluronic acid and PDA catalyzed by H <sub>2</sub> O <sub>2</sub> /HbO <sub>2</sub> to encapsulated Ti <sub>3</sub> C <sub>2</sub> MXene	Wound healing	[79]
ZIF-8	ZIF-8@Hb	MOFs	Ischemic stroke	[74]
	Hb@AuNCs	Synthesizing ultrasmall protein metal clusters	Cancer	[75]
	O <sub>2</sub> @Hb@ZIF-8	MOFs		[72]
Core-shell structural	HemoAct	Conjugating one Hb molecule with three HSA	Ischemic stroke	[140]
	Hb-HSA <sub>3</sub> (PtNP)	Combining platinum nanoparticles with HSA		[142]
	C@HPOC	Protein hybridization	Cancer	[193]
Others	H-NPs	Combining Hb with DAC	RCC	[169]
	V(Hb)@DOX	Coupling PCL self assembles to form hollow Nanovesicles	Cancer	[84]
	HCM	PMAG linking GLUT1		[82]
	DHCNPNS	Hb and DOX encapsulated in PLGA		[83]
	Au-Hb@PLT	Synthesized by coating Au-Hb NPs with PLT membrane		[85]
	P-FRT-RBC	Ferritin, a protein-based nanocage		[76]
	BP@RB-Hb	Assembling Hb into nanocomplexes with BP and PS		[192]
	OxyHb@IR820	oxyHb complexed with an anionic amphiphilic PS	P. gingivalis	[253]
	MN	Loading with black phosphorus	Wound healing	[86]
	G-HbOD	Conjugating human Hb to the surface of gelatin microspheres	3D cell culture	[257]
NERBC	Anchoring Hb on exogenous Ca <sup>2+</sup> and PO <sub>4</sub> <sup>3-</sup>	Bio-nanobatteries	[266]	

etc. in the further, we hope to develop simpler and more efficient methods for controlling and detecting methemoglobin to explore more simple and efficient methods for the control and detection of methemoglobin.

- 4) Biosafety is crucial in development of Nano-HBOCs for use in the treatment of various diseases. This is particularly important in scenarios involving high doses and multiple infusions, where in vivo metabolism, transportation, toxicity evaluation, and clearance of by-products of Nano-HBOCs are major concerns. However, there is a lack of standardized evaluation methods for hemoglobin biocompatibility, especially in tests such as antigenicity and complementation experiments. Therefore, a comprehensive and systematic standardized evaluation of Nano-HBOCs biocompatibility is essential for complete replacement of natural erythrocytes.

We firmly believe that further research on Nano-HBOCs holds immense potential for novel advancements and significant breakthroughs in multiple fields. This promising area of study not only exhibits great prospects for practical applications but also demonstrates the potential to revolutionize clinical practical and significantly enhance human health.

#### Abbreviations

AA	Arachidonic acid
BAP	Bioartificial islet transplantation
BP	Black phosphorus
bHb	Bovine hemoglobin
DHCNPNS	Camouflaged biomimetic nanocomposites
CAT	Catalase
Ec	Cell-free hemoglobin
DAC	Decitabine
DHL	DOX-Hb-lipo
DM	Diabetes mellitus
EBFC	Enzyme biofuel cells
EM	Erythromer

FDA	Food and drug administration
G-HbOD	Gelatin hemoglobin depot
GelMa	Gelatin methacryloyl
GM	Gelatin microspheres
GLUT1	Glucose transporter isoform 1
HCM	Hb-conjugated micelles
Hb	Hemoglobin
HbV	Hemoglobin vesicles
HBOCs	Hemoglobin-based oxygen carriers
HbPs	Hemoglobin-loaded nanoparticles
HbTcMs	Hemoglobin-polymer conjugate
HS	Hemorrhagic shock
HDAS	Hexadecyl carbamoyl methyl hexadecanoate
V(Hb)	Hollow nanovesicles
HeLa	Human cervical cancer cells
hHb	Human hemoglobin
HSA	Human serum albumin
HIF-1 $\alpha$	Hypoxia inducible factor-1 $\alpha$
HRP	Horseradish peroxidase
ICG	Indocyanine green
LEH	Liposome-encapsulated human hemoglobin
h-LEH	Liposomes encapsulating hemoglobin
RCC	Renal cell carcinoma
Lt	<i>Lumbricus terrestris</i>
MMP-9	Matrix metalloproteinase-9
MPA	Mean arterial pressure
MOFs	Metal-organic frameworks
MNs	Microneedles
MCAO	Middle cerebral artery occlusion
MnPcs	Mn-phthalocyanine
CT-26	Mouse colon cancer models
MDR1	Multidrug resistance gene 1
NERBCS	Nanoengineered red blood cells
NEMR	Nanoerythrocytes
Nano-HBOCs	Nanomaterial-related HBOCs
I-ARCs	Nanoscale artificial blood cells
NIR	Near-infrared
OHZ	O <sub>2</sub> @Hb@ZIF-8
OXA	Oxaliplatin
oxyHb	Oxyhemoglobin
PFCs	Perfluorocarbons
P-gp	P-glycoprotein
PDT	Photodynamic therapy
PDT	Photodynamic therapy
PS	Photosensitizer
PLT	Platelets
PDA	Polydopamine
PWS	Port-wine staining
PE	Pre-eclampsia
RT	Radiation therapy
ROS	Reactive oxygen species
RBCs	Red blood cells
FRET	Resonance energy transfer effect
SDT	Sonodynamic therapy
SOD	Superoxide dismutase
tMCAO	Transient MCAO
TME	Tumor microenvironment
TAM	Tumor-associated macrophage
ZnPc	Zinc phthalocyanine

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#### Author contributions

K.Z. and L.J.W. analyzed the data and drafted the manuscript. Y.X., X.Y.Z., G.X.Y., Y.Z.C. and Q.W. contributed to the data collection and interpretation. L.Z., H.Z., and G.C. designed this work and critically commented on the manuscript. All authors contributed to the development of the manuscript and approved the final draft.

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#### Declarations

#### Ethics approval and consent to participate

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#### Competing interests

The authors declare that they have no competing interests.

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#### References

- Motealleh A, Kehr NS. Injectable oxygen-generating nanocomposite hydrogels with prolonged oxygen delivery for enhanced cell proliferation under hypoxic and normoxic conditions. *J Mater Chem B*. 2020;8(19):4195–201.
- Ashammakhi N, et al. Advances in controlled oxygen generating biomaterials for tissue engineering and regenerative therapy. *Biomacromol*. 2020;21(1):56–72.
- Biro GP. Oxygen and ATP: the Energy Economy of the Cell. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 21–32.
- Werner M, et al. Femoral blood gas analysis, another tool to assess hemorrhage severity following trauma: an exploratory prospective study. *Scand J Trauma Resusc Emerg Med*. 2023;31(1):31.
- Dyer WB, et al. Recovery of organ-specific tissue oxygen delivery at restrictive transfusion thresholds after fluid treatment in ovine haemorrhagic shock. *Intensive Care Med Exp*. 2022;10(1):12.
- Chen WR, Chou CC, Wang CC. Phthalides serve as potent modulators to boost fetal hemoglobin induction therapy for  $\beta$ -hemoglobinopathies. *Blood Adv*. 2019;3(9):1493–8.
- Hönemann JN, et al. Hypoxia and cardiac function in patients with prior myocardial infarction. *Circ Res*. 2023;132(9):1165–7.
- Kabra R, et al. Hidden mystery behind unilateral cerebellar infarction in hanging: a case report. *Cureus*. 2022;14(11):e31115.
- Köhler D, et al. Hypoxic, anemic and cardiac hypoxemia: When does tissue hypoxia begin? *Dtsch Med Wochenschr*. 2023;148(8):475–82.
- Godoy DA, et al. Avoiding brain hypoxia in severe traumatic brain injury in settings with limited resources—a pathophysiological guide. *J Crit Care*. 2023;75:154260.
- Haddon A, et al. Cardiorespiratory responses to voluntary hyperventilation during normobaric hypoxia. *Aerosp Med Hum Perform*. 2023;94(2):59–65.
- Wang L, et al. Shen Shuai II recipe inhibits hypoxia-induced glycolysis by preserving mitochondrial dynamics to attenuate kidney fibrosis. *J Ethnopharmacol*. 2023;308:116271.
- Biro GP. Erythrocyte transfusion: brief history and current practice. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 3–19.
- Lee P, Chandel NS, Simon MC. Cellular adaptation to hypoxia through hypoxia inducible factors and beyond. *Nat Rev Mol Cell Biol*. 2020;21(5):268–83.
- Chen Z, et al. Hypoxic microenvironment in cancer: molecular mechanisms and therapeutic interventions. *Signal Transduct Target Ther*. 2023;8(1):70.
- Li B, et al. PEG-conjugated bovine haemoglobin enhances efficiency of chemotherapeutic agent doxorubicin with alleviating DOX-induced

- splenocardiac toxicity in the breast cancer. *Artif Cells Nanomed Biotechnol.* 2023;51(1):120–30.
17. Luo Z, et al. Hypoxia signaling in human health and diseases: implications and prospects for therapeutics. *Signal Transduct Target Ther.* 2022;7(1):218.
  18. Cherian VT. Physiological functions of blood. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 33–43.
  19. Cohn EJ. Blood: a brief survey of its chemical components and of their natural functions and clinical uses. *Blood.* 2015;126(24):2531.
  20. Mohanto N, Park YJ, Jee JP. Current perspectives of artificial oxygen carriers as red blood cell substitutes: a review of old to cutting-edge technologies using in vitro and in vivo assessments. *J Pharm Investig.* 2023;53(1):153–90.
  21. Ahmed MH, Ghatge MS, Safo MK. Hemoglobin: structure, function and allostery. In: Hoeger U, Harris JR, editors. *Vertebrate and invertebrate respiratory proteins, lipoproteins and other body fluid proteins*. Cham: Springer International Publishing; 2020. p. 345–82.
  22. Shu P, et al. Cefmetazole sodium as an allosteric effector that regulates the oxygen supply efficiency of adult hemoglobin. *J Biomol Struct Dyn.* 2023. <https://doi.org/10.1080/07391102.2023.2245043>.
  23. Yu S. Hemoglobin: physiology and hemoglobinopathy. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 45–51.
  24. Liu WL, et al. Aggressive man-made red blood cells for hypoxia-resistant photodynamic therapy. *Adv Mater.* 2018;30(35):e1802006.
  25. Zujalovic B, Hafner S. Damage control resuscitation: identification and treatment of life-threatening hemorrhage. *Anesth Analg.* 2020;130(5):e141–2.
  26. Carreau A, et al. Why is the partial oxygen pressure of human tissues a crucial parameter? Small molecules and hypoxia. *J Cell Mol Med.* 2011;15(6):1239–53.
  27. Haque N, et al. Hypoxic culture conditions as a solution for mesenchymal stem cell based regenerative therapy. *Sci World J.* 2013;2013:632972.
  28. Khan F, Singh K, Friedman MT. Artificial blood: the history and current perspectives of blood substitutes. *Discoveries (Craiova).* 2020;8(1):e104.
  29. Bialas C, Moser C, Sims CA. Artificial oxygen carriers and red blood cell substitutes: A historic overview and recent developments toward military and clinical relevance. *J Trauma Acute Care Surg.* 2019;87(15):s48–s58.
  30. Stanworth SJ, et al. Effects of the COVID-19 pandemic on supply and use of blood for transfusion. *Lancet Haematol.* 2020;7(10):e756–64.
  31. Coll-Satue C, et al. Stepping stones to the future of haemoglobin-based blood products: clinical, preclinical and innovative examples. *Biomater Sci.* 2021;9(4):1135–52.
  32. Dumont LJ, et al. Overnight, room temperature hold of whole blood followed by 42-day storage of red blood cells in additive solution-7. *Transfusion.* 2015;55(3):485–90.
  33. Brand A. Immunological complications of blood transfusions. *Presse Med.* 2016;45(7–8 Pt 2):e313–24.
  34. Xia S, et al. Addition of sodium pyruvate to stored red blood cells attenuates liver injury in a murine transfusion model. *Mediators Inflamm.* 2016;2016:3549207.
  35. Charbe NB, et al. A new era in oxygen therapeutics? From perfluorocarbon systems to haemoglobin-based oxygen carriers. *Blood Rev.* 2022;54:100927.
  36. Rangayasami A, et al. Influence of nanotechnology to combat against COVID-19 for global health emergency: a review. *Sensors International.* 2021;2:100079.
  37. Berg L, et al. Obstetric outcomes in Jehovah's Witnesses: case series over nine years in a London teaching hospital. *Arch Gynecol Obstet.* 2023. <https://doi.org/10.1007/s00404-023-06940-x>.
  38. Naicker YD, Ahmed N, Davids R. Damage control surgery of the critical Jehovah's Witness patient—a narrative review. *S Afr J Surg.* 2023;61(1):39–44.
  39. Liu H, et al. Classifications of blood substitutes. In: Liu H, Kaye AD, Jahr JS, editors, et al. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 119–29.
  40. Jahr JS, et al. Blood substitutes and oxygen therapeutics: a review. *Anesth Analg.* 2021;132(1):119–29.
  41. Wu L, et al. Perfluorocarbons-based (19)F magnetic resonance imaging in biomedicine. *Int J Nanomed.* 2020;15:7377–95.
  42. Castro CI, Briceno JC. Perfluorocarbon-based oxygen carriers: review of products and trials. *Artif Organs.* 2010;34(8):622–34.
  43. Park KM, Gerecht S. Hypoxia-inducible hydrogels. *Nat Commun.* 2014;5:4075.
  44. Spahn DR, Kocian R. Artificial O2 carriers: status in 2005. *Curr Pharm Des.* 2005;11(31):4099–114.
  45. Chen L, Yang Z, Liu H. Hemoglobin-based oxygen carriers: where are we now in 2023? *Med (Kaunas).* 2023. <https://doi.org/10.3390/medina59020396>.
  46. Mer M, et al. Hemoglobin glutamer-250 (bovine) in South Africa: consensus usage guidelines from clinician experts who have treated patients. *Transfusion.* 2016;56(10):2631–6.
  47. Cao M, et al. New applications of HBOC-201: a 25-year review of the literature. *Front Med (Lausanne).* 2021;8:794561.
  48. Romito BT, et al. The effect of SANGUINATE<sup>®</sup> (PEGylated carboxy-hemoglobin bovine) on cardiopulmonary bypass functionality using a bovine whole blood model of normovolemic hemodilution. *Perfusion.* 2020;35(1):19–25.
  49. Marrazzo F, et al. Inhaled nitric oxide prevents systemic and pulmonary vasoconstriction due to hemoglobin-based oxygen carrier infusion: A case report. *J Crit Care.* 2019;51:213–6.
  50. Wang Y, et al. Structural, functional and physicochemical properties of dextran-bovine hemoglobin conjugate as a hemoglobin-based oxygen carrier. *Process Biochem.* 2017;60:67–73.
  51. Huo S, et al. Ferrous hemoglobin and hemoglobin-based oxygen carriers acting as a peroxidase can inhibit oxidative damage to endothelial cells caused by hydrogen peroxide. *Artif Organs.* 2021;45(10):1229–39.
  52. Li T, Jing X, Huang Y. Polymer/hemoglobin assemblies: biodegradable oxygen carriers for artificial red blood cells. *Macromol Biosci.* 2011;11(7):865–75.
  53. Moradi S, Jahanian-Najafabadi A, Roudkenar MH. Artificial blood substitutes: first steps on the long route to clinical utility. *Clin Med Insights Blood Disord.* 2016;9:33–41.
  54. Sen Gupta A. Hemoglobin-based oxygen carriers: current state-of-the-art and novel molecules. *Shock.* 2019;52(1S):70–83.
  55. Wang Y, et al. A PEGylated bovine hemoglobin as a potent hemoglobin-based oxygen carrier. *Biotechnol Prog.* 2017;33(1):252–60.
  56. Bian Y, Chang TM. A novel nanobiotherapeutic poly-[hemoglobin-superoxide dismutase-catalase-carbonic anhydrase] with no cardiac toxicity for the resuscitation of a rat model with 90 minutes of sustained severe hemorrhagic shock with loss of 2/3 blood volume. *Artif Cells Nanomed Biotechnol.* 2015;43(1):1–9.
  57. Chang TM. Blood replacement with nanobiotechnologically engineered hemoglobin and hemoglobin nanocapsules. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2010;2(4):418–30.
  58. Bian Y, Chang TMS. Nanobiotechnological basis of an oxygen carrier with enhanced carbonic anhydrase for CO(2) transport and enhanced catalase and superoxide dismutase for antioxidant function. *Front Bioeng Biotechnol.* 2023;11:1188399.
  59. Mitchell MJ, et al. Engineering precision nanoparticles for drug delivery. *Nat Rev Drug Discov.* 2021;20(2):101–24.
  60. Bäuml H, Xiong Y, Georgieva R. Nanotechnology-based oxygen and drug carriers. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 169–73.
  61. Modery-Pawłowski CL, et al. Synthetic approaches to RBC mimicry and oxygen carrier systems. *Biomacromol.* 2013;14(4):939–48.
  62. Liu X, et al. Low-fouling electrospun hemoglobin nanoparticles with antioxidant protection as promising oxygen carriers. *Macromol Biosci.* 2020;20(2):e1900293.
  63. Nadimifar M, et al. Synthesis of bioactive hemoglobin-based oxygen carrier nanoparticles via metal-phenolic complexation. *Biomater Adv.* 2024;156:213698.
  64. Chang TMS. A brief history of the development of nanobiotechnology-based blood substitutes. In: Liu H, Kaye AD, Jahr JS, editors. *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 99–115.

65. Matsuhiro T, Sakai H. Artificial oxygen carriers, from nanometer- to micrometer-sized particles, made of hemoglobin composites substituting for red blood cells. *Particuology*. 2022;64:43–55.
66. Jia Y, Duan L, Li J. Hemoglobin-based nanoarchitectonic assemblies as oxygen carriers. *Adv Mater*. 2016;28(6):1312–8.
67. Tao Z, Ghoroghchian PP. Microparticle, nanoparticle, and stem cell-based oxygen carriers as advanced blood substitutes. *Trends Biotechnol*. 2014;32(9):466–73.
68. Jansman MMT, Hosta-Rigau L. Recent and prominent examples of nano- and microarchitectures as hemoglobin-based oxygen carriers. *Adv Colloid Interface Sci*. 2018;260:65–84.
69. Kaneda S, et al. Liposome-encapsulated hemoglobin, TRM-645: current status of the development and important issues for clinical application. *Artif Organs*. 2009;33(2):146–52.
70. Lu M, et al. Preparation, characterization and in vivo investigation of blood-compatible hemoglobin-loaded nanoparticles as oxygen carriers. *Colloid Surf B Biointerfaces*. 2016;139:171–9.
71. Xu X, et al. A photosensitizer loaded hemoglobin-polymer conjugate as a nanocarrier for enhanced photodynamic therapy. *J Mater Chem B*. 2018;6(12):1825–33.
72. Yuan M, et al. A robust oxygen-carrying hemoglobin-based natural sonosensitizer for sonodynamic cancer therapy. *Nano Lett*. 2021;21(14):6042–50.
73. Pozy E, Savla C, Palmer AF. Photocatalytic synthesis of a polydopamine-coated acellular mega-hemoglobin as a potential oxygen therapeutic with antioxidant properties. *Biomacromol*. 2023;24(5):2022–9.
74. Peng S, et al. Metal-organic framework encapsulating hemoglobin as a high-stable and long-circulating oxygen carriers to treat hemorrhagic shock. *ACS Appl Mater Interfaces*. 2019;11(39):35604–12.
75. Cun X, et al. Hemoglobin-stabilized gold nanoclusters displaying oxygen transport ability, self-antioxidation, auto-fluorescence properties and long-term storage potential. *RSC Adv*. 2023;13(23):15540–53.
76. Tang W, et al. Red blood cell-facilitated photodynamic therapy for cancer treatment. *Adv Funct Mater*. 2016;26(11):1757–68.
77. Wang Q, et al. Bioinspired polydopamine-coated hemoglobin as potential oxygen carrier with antioxidant properties. *Biomacromol*. 2017;18(4):1333–41.
78. Hu J, et al. Polydopamine-based surface modification of hemoglobin particles for stability enhancement of oxygen carriers. *J Colloid Interface Sci*. 2020;571:326–36.
79. Li Y, et al. Artificial nonenzymatic antioxidant mxene nanosheet-anchored injectable hydrogel as a mild photothermal-controlled oxygen release platform for diabetic wound healing. *ACS Nano*. 2022;16(5):7486–502.
80. Pan D, et al. Erythromer (EM), a nanoscale bio-synthetic artificial red cell: proof of concept and in vivo efficacy results. *Blood*. 2016;128(22):1027–1027.
81. Liu P, et al. Bioinspired nanoerythrocytes for metabolic microenvironment remodeling and long-term prognosis promoting of acute ischemic stroke. *Nano Today*. 2023;49:101806.
82. Bu H, et al. Synthesis of a hemoglobin-conjugated triblock copolymer for oxygen carrying and specific recognition of cancer cells. *RSC Adv*. 2017;7(76):48166–75.
83. Tian H, et al. Cancer cell membrane-biomimetic oxygen nanocarrier for breaking hypoxia-induced chemoresistance. *Adv Funct Mater*. 2017;27(38):1703197.
84. Wang Y, et al. Engineering endogenous tumor-associated macrophage-targeted biomimetic nano-RBC to reprogram tumor immunosuppressive microenvironment for enhanced chemo-immunotherapy. *Adv Mater*. 2021;33(39):e2103497.
85. Xia D, et al. Au-hemoglobin loaded platelet alleviating tumor hypoxia and enhancing the radiotherapy effect with low-dose X-ray. *ACS Nano*. 2020;14(11):15654–68.
86. Zhang X, et al. Black phosphorus-loaded separable microneedles as responsive oxygen delivery carriers for wound healing. *ACS Nano*. 2020;14(5):5901–8.
87. Munoz C, et al. Resuscitation after hemorrhagic shock in the microcirculation: targeting optimal oxygen delivery in the design of artificial blood substitutes. *Front Med (Lausanne)*. 2020;7:585638.
88. Chen G, et al. Effects of synthetic colloids on oxidative stress and inflammatory response in hemorrhagic shock: comparison of hydroxyethyl starch 130/0.4, hydroxyethyl starch 200/0.5, and succinylated gelatin. *Crit Care*. 2013;17(4):R141.
89. Bragin DE, et al. Addition of drag-reducing polymers to colloid resuscitation fluid enhances cerebral microcirculation and tissue oxygenation after traumatic brain injury complicated by hemorrhagic shock. *Adv Exp Med Biol*. 2021;1269:283–8.
90. Dyer WB, et al. An ovine model of hemorrhagic shock and resuscitation, to assess recovery of tissue oxygen delivery and oxygen debt, and inform patient blood management. *Shock*. 2021;56(6):1080–91.
91. Hallisey SD, Greenwood JC. Beyond mean arterial pressure and lactate: perfusion end points for managing the shocked patient. *Emerg Med Clin North Am*. 2019;37(3):395–408.
92. Yamamoto M, et al. Fluid resuscitation of hemorrhagic shock with hemoglobin vesicles in Beagle dogs: pilot study. *Artif Cells Blood Substit Immobil Biotechnol*. 2012;40(1–2):179–95.
93. Yuki Y, et al. Efficacy of resuscitative infusion with hemoglobin vesicles in rabbits with massive obstetric hemorrhage. *Am J Obstet Gynecol*. 2021;224(4):398.e1–398.e11.
94. Takase B, et al. Liposome-encapsulated hemoglobin (HbV) transfusion rescues rats undergoing progressive lethal 85% hemorrhage as a result of an anti-arrhythmic effect on the myocardium. *Artif Organs*. 2021;45(11):1391–404.
95. Sakai H. Overview of potential clinical applications of hemoglobin vesicles (HbV) as artificial red cells, evidenced by preclinical studies of the academic research consortium. *J Funct Biomater*. 2017. <https://doi.org/10.3390/jfb8010010>.
96. Fujihara M, et al. Primary and secondary immune responses to key-hole limpet hemocyanin in rats after infusion of hemoglobin vesicle, an artificial oxygen carrier. *Artif Organs*. 2014;38(3):234–8.
97. Sakai H, et al. One-year observation of Wistar rats after intravenous infusion of hemoglobin-vesicles (artificial oxygen carriers). *Artif Cells Blood Substit Immobil Biotechnol*. 2007;35(1):81–91.
98. Sakai H, et al. Metabolism of hemoglobin-vesicles (artificial oxygen carriers) and their influence on organ functions in a rat model. *Biomaterials*. 2004;25(18):4317–25.
99. Sakai H, et al. Hemoglobin-vesicles as oxygen carriers: influence on phagocytic activity and histopathological changes in reticuloendothelial system. *Am J Pathol*. 2001;159(3):1079–88.
100. Taguchi K, et al. Hepatically-metabolized and -excreted artificial oxygen carrier, hemoglobin vesicles, can be safely used under conditions of hepatic impairment. *Toxicol Appl Pharmacol*. 2010;248(3):234–41.
101. Taguchi K, et al. Biological responsiveness and metabolic performance of liposome-encapsulated hemoglobin (Hemoglobin-Vesicles) in apolipoprotein E-deficient mice after massive intravenous injection. *Biol Pharm Bull*. 2015;38(10):1606–16.
102. Shono S, et al. Intraosseous transfusion with liposome-encapsulated hemoglobin improves mouse survival after hypohemoglobinemic shock without scavenging nitric oxide. *Shock*. 2011;35(1):45–52.
103. Eriksson M, et al. The effect of hemorrhagic shock and intraosseous adrenaline injection on the delivery of a subsequently administered drug - an experimental study. *Scand J Trauma Resusc Emerg Med*. 2019;27(1):29.
104. Yadav VR, et al. Nanovesicular liposome-encapsulated hemoglobin (LEH) prevents multi-organ injuries in a rat model of hemorrhagic shock. *Eur J Pharm Sci*. 2016;93:97–106.
105. Nag OK, et al. Post-modification of preformed liposomes with novel non-phospholipid poly(ethylene glycol)-conjugated hexadecylcarbamoylmethyl hexadecanoic acid for enhanced circulation persistence in vivo. *Int J Pharm*. 2013;446(1–2):119–29.
106. Agashe H, et al. Improved formulation of liposome-encapsulated hemoglobin with an anionic non-phospholipid. *Colloids Surf B Biointerfaces*. 2010;75(2):573–83.
107. Mittal N, et al. Erythromer (EM), a Nanoscale Bio-Synthetic Artificial Red Cell. In: Liu H, Kaye AD, Jahr JS, editors., et al., *Blood substitutes and oxygen biotherapeutics*. Cham: Springer International Publishing; 2022. p. 253–65.
108. Okamoto W, et al. Hemoglobin-albumin clusters as an artificial O<sub>2</sub> carrier: Physicochemical properties and resuscitation from hemorrhagic shock in rats. *J Biomed Mater Res B Appl Biomater*. 2022;110(8):1827–38.



109. Liu Y, Ai K, Lu L. Polydopamine and its derivative materials: synthesis and promising applications in energy, environmental, and biomedical fields. *Chem Rev*. 2014;114(9):5057–115.
110. Zhao Y, et al. Hollow PDA-Au nanoparticles-enabled signal amplification for sensitive nonenzymatic colorimetric immunodetection of carbohydrate antigen 125. *Biosens Bioelectron*. 2015;71:200–6.
111. Baidukova O, et al. Antioxidative protection of haemoglobin micro-particles (HbMPs) by PolyDopamine. *Artif Cells Nanomed Biotechnol*. 2018;46(sup3):S693-s701.
112. Jansman MMT, et al. Hemoglobin-based oxygen carriers incorporating nanozymes for the depletion of reactive oxygen species. *ACS Appl Mater Interfaces*. 2020;12(45):50275–86.
113. Liu X, et al. Metal-organic framework-based oxygen carriers with anti-oxidant activity resulting from the incorporation of gold nanozymes. *Biomater Sci*. 2023;11(7):2551–65.
114. Lian X, et al. Enzyme-MOF (metal-organic framework) composites. *Chem Soc Rev*. 2017;46(11):3386–401.
115. Chen Y, et al. Acid-resistant mesoporous metal-organic framework toward oral insulin delivery: protein encapsulation, protection, and release. *J Am Chem Soc*. 2018;140(17):5678–81.
116. Wang S, et al. DNA-functionalized metal-organic framework nanoparticles for intracellular delivery of proteins. *J Am Chem Soc*. 2019;141(6):2215–9.
117. Liang K, et al. Biomimetic mineralization of metal-organic frameworks as protective coatings for biomacromolecules. *Nat Commun*. 2015;6:7240.
118. Chen G, et al. A convenient and versatile amino-acid-boosted biomimetic strategy for the nondestructive encapsulation of biomacromolecules within metal-organic frameworks. *Angew Chem Int Ed Engl*. 2019;58(5):1463–7.
119. Poddar A, et al. Encapsulation, visualization and expression of genes with biomimetically mineralized zeolitic imidazolate framework-8 (ZIF-8). *Small*. 2019;15(36):e1902268.
120. Jansman MMT, et al. Hemoglobin-based oxygen carriers camouflaged with membranes extracted from red blood cells: optimization and assessment of functionality. *Biomater Adv*. 2022;134:112691.
121. Sun H, et al. Real-time model-based cerebral perfusion calculation for ischemic stroke. *Comput Methods Programs Biomed*. 2024;243: 107916.
122. Demyanenko S, Dzreyan V, Sharifulina S. Histone deacetylases and their isoform-specific inhibitors in ischemic stroke. *Biomedicines*. 2021;9(10):1445.
123. Hurd MD, et al. Current status of ischemic stroke treatment: from thrombolysis to potential regenerative medicine. *Regenerat Ther*. 2021;18:408–17.
124. Wu DM, et al. Immune pathway activation in neurons triggers neural damage after stroke. *Cell Rep*. 2023;42(11):113368.
125. Jun-Long H, et al. Necroptosis signaling pathways in stroke: from mechanisms to therapies. *Curr Neuropharmacol*. 2018;16(9):1327–39.
126. Lin W, Powers WJ. Oxygen metabolism in acute ischemic stroke. *J Cereb Blood Flow Metab*. 2018;38(9):1481–99.
127. Tirapelli CR. Oxidative stress and vascular disease. *Curr Hypertens Rev*. 2020;16(3):162.
128. Zuluaga Tamayo M, et al. Astaxanthin complexes to attenuate muscle damage after in vivo femoral ischemia-reperfusion. *Mar Drugs*. 2019. <https://doi.org/10.3390/md17060354>.
129. Deuchar GA, et al. Preclinical validation of the therapeutic potential of glasgow oxygen level dependent (GOLD) technology: a theranostic for acute stroke. *Transl Stroke Res*. 2019;10(5):583–95.
130. Klaus JA, et al. Early treatment of transient focal cerebral ischemia with bovine PEGylated carboxy hemoglobin transfusion. *Artificial Cell Blood Substitutes Biotechnol*. 2010;38(5):223–9.
131. Kakehata J, et al. Therapeutic potentials of an artificial oxygen-carrier, liposome-encapsulated hemoglobin, for ischemia/reperfusion-induced cerebral dysfunction in rats. *J Pharmacol Sci*. 2010;114(2):189–97.
132. Kaneda S, et al. Efficacy of liposome-encapsulated hemoglobin in a rat model of cerebral ischemia. *Artif Organs*. 2014;38(8):650–5.
133. Cipolla MJ, et al. Pharmacologically increasing collateral perfusion during acute stroke using a carboxyhemoglobin gas transfer agent (Sanguinate™) in spontaneously hypertensive rats. *J Cereb Blood Flow Metab*. 2018;38(5):755–66.
134. Powanda DD, Chang TMS. Cross-linked polyhemoglobin-superoxide dismutase-catalase supplies oxygen without causing blood brain barrier disruption or brain edema in a rat model of transient global brain ischemia-reperfusion. *Artificial Cell Blood Substitutes Biotechnol*. 2002;30(1):23–37.
135. Xie Z, et al. The protective effect of polymerized porcine hemoglobin (pPolyHb) on transient focal cerebral ischemia/reperfusion injury. *Artif Cells Nanomed Biotechnol*. 2015;43(3):180–5.
136. Caswell JE, et al. A novel hemoglobin-based blood substitute protects against myocardial reperfusion injury. *Am J Physiol-Heart Circulatory Physiol*. 2005;288(4):H1796–801.
137. Komatsu H, et al. Effect of hemoglobin vesicle, a cellular-type artificial oxygen carrier, on middle cerebral artery occlusion- and arachidonic acid-induced stroke models in rats. *Neurosci Lett*. 2007;421(2):121–5.
138. Fukumoto D, et al. Liposome-encapsulated hemoglobin reduces the size of cerebral infarction in rats: effect of oxygen affinity. *Artif Organs*. 2009;33(2):159–63.
139. Kawaguchi AT, et al. Effect of oxygen affinity of liposome-encapsulated hemoglobin on cerebral ischemia and reperfusion as detected by positron emission tomography in nonhuman primates. *Artif Organs*. 2017;41(4):336–45.
140. Tomita D, et al. Covalent core-shell architecture of hemoglobin and human serum albumin as an artificial O<sub>2</sub> carrier. *Biomacromol*. 2013;14(6):1816–25.
141. Gekka M, et al. Novel hemoglobin-based oxygen carrier bound with albumin shows neuroprotection with possible antioxidant effects. *Stroke*. 2018;49(8):1960–8.
142. Hosaka H, et al. Hemoglobin-albumin cluster incorporating a Pt nanoparticle: artificial O<sub>2</sub> carrier with antioxidant activities. *PLoS ONE*. 2014;9(10):e110541.
143. Wang S, et al. Pan-immune-inflammatory value predicts the 3 month outcome in acute ischemic stroke patients after intravenous thrombolysis. *Curr Neurovasc Res*. 2023. <https://doi.org/10.2174/0115672026276427231024045957>.
144. Haupt M, et al. Preconditioning concepts for the therapeutic use of extracellular vesicles against stroke. *Stem Cells Transl Med*. 2023. <https://doi.org/10.1093/stcltm/szad055>.
145. Yin N, et al. Engineered nanoerythrocytes alleviate central nervous system inflammation by regulating the polarization of inflammatory microglia. *Adv Mater*. 2022;34(27):e2201322.
146. Shimbo D, et al. Post-ischemic intra-arterial infusion of liposome-encapsulated hemoglobin can reduce ischemia reperfusion injury. *Brain Res*. 2014;1554:59–66.
147. Shimbo D, et al. Superior microvascular perfusion of infused liposome-encapsulated hemoglobin prior to reductions in infarctions after transient focal cerebral ischemia. *J Stroke Cerebrovasc Dis*. 2017;26(12):2994–3003.
148. Choudhari OK, et al. Matrix metalloproteinase-9 gene polymorphism and its methylation in stroke patients. *Malays J Med Sci*. 2021;28(6):32–41.
149. Viillard C, Larrivée B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis*. 2017;20(4):409–26.
150. Liao C, et al. Tumor hypoxia: from basic knowledge to therapeutic implications. *Semin Cancer Biol*. 2023;88:172–86.
151. Dai Y, et al. Nanoparticle design strategies for enhanced anticancer therapy by exploiting the tumour microenvironment. *Chem Soc Rev*. 2017;46(12):3830–52.
152. Kwon S, et al. Nanomedicines for reactive oxygen species mediated approach: an emerging paradigm for cancer treatment. *Acc Chem Res*. 2019;52(7):1771–82.
153. Shi R, Tang YQ, Miao H. Metabolism in tumor microenvironment: Implications for cancer immunotherapy. *MedComm*. 2020;1(1):47–68.
154. Semenza GL. Oxygen sensing, hypoxia-inducible factors, and disease pathophysiology. *Annu Rev Pathol*. 2014;9:47–71.
155. Gilkes DM, Semenza GL, Wirtz D. Hypoxia and the extracellular matrix: drivers of tumour metastasis. *Nat Rev Cancer*. 2014;14(6):430–9.
156. Yang S, et al. Boosting the anti-tumor performance of disulfiram against glioblastoma by using ultrasmall nanoparticles and HIF-1 $\alpha$  inhibitor. *Compos B Eng*. 2022;243:110117.
157. Liu Y, et al. Modulation of hypoxia and redox in the solid tumor microenvironment with a catalytic nanoplatform to enhance

- combinational chemodynamic/sonodynamic therapy. *Biomater Sci.* 2023;11(5):1739–53.
158. Huang J, et al. Oxygen-carrying nanoplatform to reprogram tumor immunosuppressive microenvironment and enhance photothermal-immunotherapy. *Mater Today Bio.* 2023;19:100555.
159. Yuan CS, et al. Hypoxia-modulatory nanomaterials to relieve tumor hypoxic microenvironment and enhance immunotherapy: where do we stand? *Acta Biomater.* 2021;125:1–28.
160. Telarovic I, Wenger RH, Pruschy M. Interfering with tumor hypoxia for radiotherapy optimization. *J Exp Clin Cancer Res.* 2021;40(1):197.
161. Zhang J, et al. Nanotechnological strategies to increase the oxygen content of the tumor. *Front Pharmacol.* 2023;14:1140362.
162. Vaupel P, Multhoff G. Fatal alliance of hypoxia-/HIF-1 $\alpha$ -Driven microenvironmental traits promoting cancer progression. *Adv Exp Med Biol.* 2020;1232:169–76.
163. Mortezaee K, Majidpoor J. The impact of hypoxia on immune state in cancer. *Life Sci.* 2021;286:120057.
164. Doublier S, et al. HIF-1 activation induces doxorubicin resistance in MCF7 3-D spheroids via P-glycoprotein expression: a potential model of the chemo-resistance of invasive micropapillary carcinoma of the breast. *BMC Cancer.* 2012;12:4.
165. Samanta D, et al. Hypoxia-inducible factors are required for chemotherapy resistance of breast cancer stem cells. *Proc Natl Acad Sci U S A.* 2014;111(50):E5429–38.
166. Kawaguchi F, et al. Liposome-encapsulated hemoglobin improves tumor oxygenation as detected by near-infrared spectroscopy in colon carcinoma in mice. *Artif Organs.* 2017;41(4):327–35.
167. Liu Y, et al. Epigenetic activation of the drug transporter OCT2 sensitizes renal cell carcinoma to oxaliplatin. *Sci Trans Med.* 2016;8(348):348ra97–348ra97.
168. Zheng X, et al. Response to Comment on “epigenetic activation of the drug transporter OCT2 sensitizes renal cell carcinoma to oxaliplatin.” *Sci Trans Med.* 2017;9(391):eaam6298.
169. Chen L, et al. The failure of DAC to induce OCT2 expression and its remission by hemoglobin-based nanocarriers under hypoxia in renal cell carcinoma. *Theranostics.* 2020;10(8):3562–78.
170. Guo Y, et al. Cell microenvironment-controlled antitumor drug releasing-nanomicelles for GLUT1-targeting hepatocellular carcinoma therapy. *ACS Appl Mater Interfaces.* 2015;7(9):5444–53.
171. Synthesis of a hemoglobin-conjugated triblock copolymer for oxygen carrying and specific recognition of cancer cells. *RSC Adv.* 2017;7(76):48166–48175.
172. Yang, J, et al., Hypoxic tumor therapy by hemoglobin-mediated drug delivery and reversal of hypoxia-induced chemoresistance. *Biomaterials*, 2018. 182: p. 145–156.
173. Sies H, Jones DP. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat Rev Mol Cell Biol.* 2020;21(7):363–83.
174. Jia S, et al. Promoting reactive oxygen species generation: a key strategy in nanosensitizer-mediated radiotherapy. *Nanomedicine (Lond).* 2021;16(9):759–78.
175. Huang L, et al. Photodynamic therapy for hypoxic tumors: advances and perspectives. *Coord Chem Rev.* 2021;438:213888.
176. Li M, et al. Near-infrared light-initiated molecular superoxide radical generator: rejuvenating photodynamic therapy against hypoxic tumors. *J Am Chem Soc.* 2018;140(44):14851–9.
177. Wang S, et al. Hemoglobins from scapharca subcrenata (Bivalvia: Arcidae) likely play a bactericidal role through their peroxidase activity. *Comp Biochem Physiol B: Biochem Mol Biol.* 2021;253:110545.
178. Wang R, et al. PEGylated hollow gold nanoparticles for combined X-ray radiation and photothermal therapy in vitro and enhanced CT imaging in vivo. *Nanomed Nanotechnol Biol Med.* 2019;16:195–205.
179. Goel S, Ni D, Cai W. Harnessing the power of nanotechnology for enhanced radiation therapy. *ACS Nano.* 2017;11(6):5233–7.
180. Sang W, et al. Oxygen-enriched metal-phenolic X-ray nanoprocessor for cancer radio-radiodynamic therapy in combination with checkpoint blockade immunotherapy. *Adv Sci.* 2021;8(4):2003338.
181. Hu CM, et al. Nanoparticle biointerfacial by platelet membrane cloaking. *Nature.* 2015;526(7571):118–21.
182. Yu M, et al. Development of “smart” drug delivery systems for chemo/PDT synergistic treatment. *J Mater Chem B.* 2023;11(7):1416–33.
183. Mossakowska BJ, et al. Possible mechanisms of resistance development to photodynamic therapy (PDT) in vulvar cancer cells. 2022. <https://doi.org/10.3390/ijms23214689>.
184. Dang J, et al. Manipulating tumor hypoxia toward enhanced photodynamic therapy (PDT). *Biomater Sci.* 2017;5(8):1500–11.
185. Zhou Z, et al. Reactive oxygen species generating systems meeting challenges of photodynamic cancer therapy. *Chem Soc Rev.* 2016;45(23):6597–626.
186. Feng L, et al. Multifunctional UCNPs@MnSiO<sub>3</sub>@g-C(3)N(4) nanoplatform: improved ROS generation and reduced glutathione levels for highly efficient photodynamic therapy. *Biomater Sci.* 2017;5(12):2456–67.
187. Chae YC, et al. Mitochondrial akt regulation of hypoxic tumor reprogramming. *Cancer Cell.* 2016;30(2):257–72.
188. Liang T, et al. Adapting and remodeling: orchestrating tumor microenvironment normalization with photodynamic therapy by size transformable nanoframeworks. *Angew Chem Int Ed Engl.* 2021;60(20):11464–73.
189. Shi X, et al. Hemoglobin-mediated biomimetic synthesis of paramagnetic O(2)-evolving theranostic nanoprobe for MR imaging-guided enhanced photodynamic therapy of tumor. *Theranostics.* 2020;10(25):11607–21.
190. Wang S, et al. Synthesis of hemoglobin conjugated polymeric micelle: a ZnPc carrier with oxygen self-compensating ability for photodynamic therapy. *Biomacromol.* 2015;16(9):2693–700.
191. Luo Z, et al. Self-monitoring artificial red cells with sufficient oxygen supply for enhanced photodynamic therapy. *Sci Rep.* 2016;6:23393.
192. Cao H, et al. An assembled nanocomplex for improving both therapeutic efficiency and treatment depth in photodynamic therapy. *Angew Chem Int Ed Engl.* 2018;57(26):7759–63.
193. Chen Z, et al. Bioinspired hybrid protein oxygen nanocarrier amplified photodynamic therapy for eliciting anti-tumor immunity and abscopal effect. *ACS Nano.* 2018;12(8):8633–45.
194. Steeg PS. Tumor metastasis: mechanistic insights and clinical challenges. *Nat Med.* 2006;12(8):895–904.
195. Xu T, et al. Enhanced ferroptosis by oxygen-boosted phototherapy based on a 2-in-1 nanoplatform of ferrous hemoglobin for tumor synergistic therapy. *ACS Nano.* 2020;14(3):3414–25.
196. Wang H, et al. Discovery of ML210-Based glutathione peroxidase 4 (GPX4) degrader inducing ferroptosis of human cancer cells. *Eur J Med Chem.* 2023;254:115343.
197. Hu H, et al. Emerging nanomedicine-enabled/enhanced nanodynamic therapies beyond traditional photodynamics. *Adv Mater.* 2021;33(12):e2005062.
198. Jiang Q, et al. A hydrogen peroxide economizer for on-demand oxygen production-assisted robust sonodynamic immunotherapy. *Theranostics.* 2022;12(1):59–75.
199. Yin T, et al. Hypoxia-alleviated sonodynamic therapy based on a hybrid protein oxygen carrier to enhance tumor inhibition. *Biomater Sci.* 2021;10(1):294–305.
200. Pan ZQ, et al. Construction and evaluation of zeolitic imidazolate framework-encapsulated hemoglobin microparticles as oxygen carriers. *ACS Appl Bio Mater.* 2023;6(4):1471–8.
201. Liang G, et al. Reduction of platinum(IV) prodrug hemoglobin nanoparticles with deeply penetrating ultrasound radiation for tumor-targeted therapeutically enhanced anticancer therapy. *Angew Chem Int Ed.* 2023;62(22):e202301074.
202. Mortezaee K, Majidpoor J. The impact of hypoxia on extracellular vesicle secretome profile of cancer. *Med Oncol.* 2023;40(5):128.
203. Wijekoon A, Fountas-Davis N, Leipzig ND. Fluorinated methacrylamide chitosan hydrogel systems as adaptable oxygen carriers for wound healing. *Acta Biomater.* 2013;9(3):5653–64.
204. Younis I. Role of oxygen in wound healing. *J Wound Care.* 2020;29(Sup5b):S4–S10.
205. Gupta S, et al. Dynamic role of oxygen in wound healing: a microbial, immunological, and biochemical perspective. *Arch Razi Inst.* 2022;77(2):513–23.
206. Sen CK. Wound healing essentials: let there be oxygen. *Wound Repair Regen.* 2009;17(1):1–18.
207. Zhu Y, et al. Roxadustat promotes angiogenesis through HIF-1 $\alpha$ /VEGF/VEGFR2 signaling and accelerates cutaneous wound healing in diabetic rats. *Wound Repair Regen.* 2019;27(4):324–34.

208. Thangarajah H, et al. HIF-1 $\alpha$  dysfunction in diabetes. *Cell Cycle*. 2010;9(1):75–9.
209. Fosen KM, Thom SR. Hyperbaric oxygen, vasculogenic stem cells, and wound healing. *Antioxid Redox Signal*. 2014;21(11):1634–47.
210. Wang X, et al. In situ 3D bioprinting living photosynthetic scaffolds for autotrophic wound healing. *Research*. 2022. <https://doi.org/10.34133/2022/9794745>.
211. Kang JI, Park KM. Oxygen-supplying syringe to create hyperoxia-inducible hydrogels for in situ tissue regeneration. *Biomaterials*. 2023;293:121943.
212. Zhao E, et al. Separable microneedles with photosynthesis-driven oxygen manufactory for diabetic wound healing. *ACS Appl Mater Interfaces*. 2023;15(6):7725–34.
213. Chen H, et al. Dissolved oxygen from microalgae-gel patch promotes chronic wound healing in diabetes. *Sci Adv*. 2020;6(20):eaba4311.
214. Wang T, et al. Combined antioxidant-antibiotic treatment for effectively healing infected diabetic wounds based on polymer vesicles. *ACS Nano*. 2021;15(5):9027–38.
215. Hauck S, et al. Collagen/hyaluronan based hydrogels releasing sulfated hyaluronan improve dermal wound healing in diabetic mice via reducing inflammatory macrophage activity. *Bioact Mater*. 2021;6(12):4342–59.
216. Wang S, et al. Nanoenzyme-reinforced injectable hydrogel for healing diabetic wounds infected with multidrug resistant bacteria. *Nano Lett*. 2020;20(7):5149–58.
217. Jia Z, et al. Bioinspired conductive silk microfiber integrated bioelectronic for diagnosis and wound healing in diabetes. *Adv Func Mater*. 2021;31(19):2010461.
218. Chen Y, et al. Research advances in smart responsive-hydrogel dressings with potential clinical diabetic wound healing properties. *Mil Med Res*. 2023;10(1):37.
219. Thangarajah H, et al. The molecular basis for impaired hypoxia-induced VEGF expression in diabetic tissues. *Proc Natl Acad Sci U S A*. 2009;106(32):13505–10.
220. Fukui T, et al. Liposome-encapsulated hemoglobin accelerates skin wound healing in diabetic dB/dB mice. *Artif Organs*. 2017;41(4):319–26.
221. Rius J, et al. NF- $\kappa$ B links innate immunity to the hypoxic response through transcriptional regulation of HIF-1 $\alpha$ . *Nature*. 2008;453(7196):807–11.
222. Jin L, et al. An NIR photothermal-responsive hybrid hydrogel for enhanced wound healing. *Bioact Mater*. 2022;16:162–72.
223. Peng Y, et al. Construction of heparin-based hydrogel incorporated with Cu<sub>5</sub>S<sub>4</sub> ultrasmall nanozymes for wound healing and inflammation inhibition. *Bioact Mater*. 2021;6(10):3109–24.
224. Waris TS, et al. Chitosan-sodium percarbonate-based hydrogels with sustained oxygen release potential stimulated angiogenesis and accelerated wound healing. *J Biomed Mater Res B Appl Biomater*. 2023. <https://doi.org/10.1002/jbm.b.35344>.
225. Yang Z, et al. Oxygen-generating hydrogels as oxygenation therapy for accelerated chronic wound healing. *Adv Healthc Mater*. 2023. <https://doi.org/10.1002/adhm.202302391>.
226. Shiekh PA, Singh A, Kumar A. Oxygen-releasing antioxidant cryogel scaffolds with sustained oxygen delivery for tissue engineering applications. *ACS Appl Mater Interfaces*. 2018;10(22):18458–69.
227. Frykberg R, et al. Use of Topical Oxygen Therapy in Wound Healing. *J Wound Care*. 2023;32(Sup8b):S1–S32.
228. Liu Y, et al. Responsive porous microcarriers with controllable oxygen delivery for wound healing. *Small*. 2019;15(21):e1901254.
229. Zhang X, et al. Encoded microneedle arrays for detection of skin interstitial fluid biomarkers. *Adv Mater*. 2019;31(37):e1902825.
230. Li W, et al. Rapidly separable microneedle patch for the sustained release of a contraceptive. *Nat Biomed Eng*. 2019;3(3):220–9.
231. Priya S, et al. Concurrent occurrence of lobular capillary haemangioma and port-wine stain: a case report and literature review. *Cureus*. 2023;15(5):e38642.
232. Demirel Ögüt N. Acquired plantar port-wine stain (Fegeler syndrome) following prolonged standing. *J Cosmet Dermatol*. 2022;21(10):5234–5.
233. Jiang F, et al. Influence of port-wine stains on quality of life of children and their parents. *Acta Derm Venereol*. 2021;101(8):adv00516.
234. Artzi O, et al. Treatment of port wine stain with Tixel-induced rapamycin delivery following pulsed dye laser application. *Dermatol Ther*. 2020;33(1):e13172.
235. Sabeti S, et al. Consensus statement for the management and treatment of port-wine birthmarks in sturge-weber syndrome. *JAMA Dermatol*. 2021;157(1):98–104.
236. Yang C, et al. Mapping port wine stain in vivo by optical coherence tomography angiography and multi-metric characterization. *Opt Express*. 2023;31(9):13613–26.
237. Sakai H, et al. O<sub>2</sub> release from Hb vesicles evaluated using an artificial, narrow O<sub>2</sub>-permeable tube: comparison with RBCs and acellular Hbs. *Am J Physiol-Heart Circulatory Physiol*. 2003;285(6):H2543–51.
238. Rikihisa N, et al. Artificial red blood cells as potential photosensitizers in dye laser treatment against port-wine stains. *J Funct Biomater*. 2017;8(2):14.
239. Ranjbar A, et al. Machine learning models for predicting pre-eclampsia: a systematic review protocol. *BMJ Open*. 2023;13(9):e074705.
240. Davidson KW, et al. Screening for gestational diabetes: us preventive services task force recommendation statement. *JAMA*. 2021;326(6):531–8.
241. Henderson JT, et al. Preeclampsia screening: evidence report and systematic review for the US preventive services task force. *JAMA*. 2017;317(16):1668–83.
242. Burton GJ, et al. Rheological and physiological consequences of conversion of the maternal spiral arteries for uteroplacental blood flow during human pregnancy. *Placenta*. 2009;30(6):473–82.
243. Fruci S, et al. Pravastatin for severe preeclampsia with growth restriction: Placental findings and infant follow-up. *Eur J Obstet Gynecol Reprod Biol*. 2023;283:37–42.
244. Myatt L, Webster RP. Vascular biology of preeclampsia. *J Thromb Haemost*. 2009;7(3):375–84.
245. Powe CE, Levine RJ, Karumanchi SA. Preeclampsia, a disease of the maternal endothelium: the role of antiangiogenic factors and implications for later cardiovascular disease. *Circulation*. 2011;123(24):2856–69.
246. Cotechini T, et al. Inflammation in rat pregnancy inhibits spiral artery remodeling leading to fetal growth restriction and features of preeclampsia. *J Exp Med*. 2014;211(1):165–79.
247. Kawaguchi AT, et al. Liposome-encapsulated hemoglobin reduces the size of cerebral infarction in the rat. *Stroke*. 2007;38(5):1626–32.
248. Chang TMS. Therapeutic applications of polymeric artificial cells. *Nat Rev Drug Discovery*. 2005;4(3):221–35.
249. Ohta H, et al. Potential new non-invasive therapy using artificial oxygen carriers for pre-eclampsia. *J Funct Biomater*. 2017. <https://doi.org/10.3390/jfb8030032>.
250. Mysak J, et al. Porphyromonas gingivalis: major periodontopathic pathogen overview. *J Immunol Res*. 2014;2014:476068.
251. Yang J, et al. Porphyromonas gingivalis oral infection promote T helper 17/Treg imbalance in the development of atherosclerosis. *J Dent Sci*. 2017;12(1):60–9.
252. Paramaesvaran M, et al. Porphyrin-Mediated Cell Surface Heme Capture from Hemoglobin by *Porphyromonas gingivalis*. *J Bacteriol*. 2003;185(8):2528–37.
253. Bai L, et al. Oxyhemoglobin-based nanophotosensitizer for specific and synergistic photothermal and photodynamic therapies against porphyromonas gingivalis oral infection. *ACS Biomater Sci Eng*. 2023;9(1):485–97.
254. Mao C, et al. Local photothermal/photodynamic synergistic therapy by disrupting bacterial membrane to accelerate reactive oxygen species permeation and protein leakage. *ACS Appl Mater Interfaces*. 2019;11(19):17902–14.
255. Zhang H, et al. Dual-mode antibacterial conjugated polymer nanoparticles for photothermal and photodynamic therapy. *Macromol Biosci*. 2020;20(2):e1900301.
256. Hu X, et al. Synergistic antibacterial strategy based on photodynamic therapy: Progress and perspectives. *Chem Eng J*. 2022;450:138129.
257. Paciello A, et al. Hemoglobin-conjugated gelatin microsphere as a smart oxygen releasing biomaterial. *Adv Healthc Mater*. 2016;5(20):2655–66.
258. Jones GL, et al. Time course and quantification of pancreatic islet revascularization following intraportal transplantation. *Cell Transplant*. 2007;16(5):505–16.

259. Rodriguez-Brotons A, et al. Impact of pancreatic rat islet density on cell survival during hypoxia. *J Diabetes Res*. 2016;2016:3615286.
260. Okere B, et al. Cell therapies for pancreatic beta-cell replenishment. *Ital J Pediatr*. 2016;42(1):62.
261. Mouré A, et al. Extracellular hemoglobin combined with an O<sub>2</sub>-generating material overcomes O<sub>2</sub> limitation in the bioartificial pancreas. *Biotechnol Bioeng*. 2019;116(5):1176–89.
262. Le Pape F, et al. Advancement in recombinant protein production using a marine oxygen carrier to enhance oxygen transfer in a CHO-S cell line. *Artif Cells Nanomed Biotechnol*. 2015;43(3):186–95.
263. Sue C-Y, Tsai N-C. Human powered MEMS-based energy harvest devices. *Appl Energy*. 2012;93:390–403.
264. Zebda A, et al. Mediatorless high-power glucose biofuel cells based on compressed carbon nanotube-enzyme electrodes. *Nat Commun*. 2011;2:370.
265. Weltin A, et al. Polymer-based, flexible glutamate and lactate microsenors for in vivo applications. *Biosens Bioelectron*. 2014;61:192–9.
266. Chen H, et al. In Situ Engineering of Intracellular Hemoglobin for Implantable High-Performance Biofuel Cells. *Angew Chem Int Ed Engl*. 2019;58(20):6663–8.
267. Rikihisa N, et al. Artificial red blood cells as potential photosensitizers in dye laser treatment against port-wine stains. *J Funct Biomater*. 2017. <https://doi.org/10.3390/jfb8020014>.

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