## **REVIEW**

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# Recent developments in two-dimensional molybdenum disulfde-based multimodal cancer theranostics

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## **Abstract**

Recent advancements in cancer research have led to the generation of innovative nanomaterials for improved diagnostic and therapeutic strategies. Despite the proven potential of two-dimensional (2D) molybdenum disulfde  $(MoS<sub>2</sub>)$  as a versatile platform in biomedical applications, few review articles have focused on MoS<sub>2</sub>-based platforms for cancer theranostics. This review aims to fll this gap by providing a comprehensive overview of the latest developments in 2D MoS<sub>2</sub> cancer theranostics and emerging strategies in this field. This review highlights the potential applications of  $2D$  MoS<sub>2</sub> in single-model imaging and therapy, including fluorescence imaging, photoacoustic imaging, photothermal therapy, and catalytic therapy. This review further classifes the potential of 2D MoS<sub>2</sub> in multimodal imaging for diagnostic and synergistic theranostic platforms. In particular, this review underscores the progress of 2D MoS<sub>2</sub> as an integrated drug delivery system, covering a broad spectrum of therapeutic strategies from chemotherapy and gene therapy to immunotherapy and photodynamic therapy. Finally, this review discusses the current challenges and future perspectives in meeting the diverse demands of advanced cancer diagnostic and theranostic applications.

**Keywords** Two-dimensional materials, Molybdenum disulfde, Nanomedicine, Bioimaging, Cancer therapy

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#### **Introduction**

In the post-COVID era, the urgency for further advancements in cancer theranostics has become increasingly intense [\[1](#page-25-0), [2\]](#page-25-1). While conventional therapeutic and diagnostic modalities have benefts, they also have inherent limitations. These limitations include unavoidable adverse reactions due to the nonspecifcity of chemotherapy agents, the inability to eliminate micrometastases resulting from the localized nature of surgery and radiation therapy, and the inadequate imaging resolution provided by commonly used contrast agents [\[3–](#page-25-2)[5\]](#page-25-3). Consequently, these limitations underscore the urgent need for the development of more efficacious theranostic strategies. Notably, emerging nanotechnologies, particularly two-dimensional (2D) materials, have been positioned at the forefront of research in accelerating cancer theranostics [[6,](#page-25-4) [7](#page-25-5)]. Among the various 2D materials recently explored, 2D transition metal dichalcogenides (TMDs) stand out for their promising advantages. These advantages include (1) their small size, which allows for the enhanced permeation and retention (EPR) effect  $[8, 9]$  $[8, 9]$  $[8, 9]$  $[8, 9]$  $[8, 9]$ ; (2) their potential for ensuring biocompatibility, physiological stability and safety  $[10, 11]$  $[10, 11]$  $[10, 11]$  $[10, 11]$ ; and  $(3)$  their high specific surface area, which facilitates the easy loading or delivery of functional molecules, imaging or therapeutic agents, thus combining both imaging and therapeutic functionalities [[12–](#page-25-10)[14](#page-25-11)].

Among all known TMDs, 2D molybdenum disulfide  $(MoS<sub>2</sub>)$  has garnered considerable attention in the biomedical feld because of its exceptional inherent properties  $[15, 16]$  $[15, 16]$  $[15, 16]$  $[15, 16]$ . These properties include

photoluminescence (PL) resulting from an indirect bandgap and distinct fuorescence properties; remarkable photothermal properties, such as strong absorption in the near-infrared (NIR) region; an outstanding photothermal conversion rate; enzyme-like catalytic activity; and a potent X-ray attenuation ability attributed to the high atomic number of Mo [[17–](#page-25-14)[20\]](#page-26-0). Consequently,  $2D$  MoS<sub>2</sub> has been utilized in applications such as optical imaging, computerized tomography (CT), photothermal therapy (PTT), and catalytic therapy. Like other 2D nanomaterials, 2D  $MoS<sub>2</sub>$  has an ultrahigh surface-tovolume ratio, thereby acting as a versatile nanoplatform to load various agents for tumor treatment [\[21](#page-26-1)]. However, challenges such as limited biocompatibility and a propensity to aggregate in physiological environments have restricted the widespread application of conventional 2D  $MoS_2$  [[22,](#page-26-2) [23](#page-26-3)]. The growing demand for multifunctionality in biomedical felds has necessitated the optimization of 2D  $MoS<sub>2</sub>$ . Surface modification strategies, involving the introduction of functional molecules such as biocompatible polyethylene glycol (PEG) and targeted folic acid (FA), have been employed to confer increased physiological stability, enhanced biocompatibility, and improved targeting capabilities to 2D  $MoS_2$  [\[24](#page-26-4), [25\]](#page-26-5). Furthermore, the incorporation of other nanoparticles can provide additional functionality and expand the range of applications for 2D  $MoS<sub>2</sub>$ . For instance, the attachment of superparamagnetic nanoparticles can endow 2D  $MoS<sub>2</sub>$  with magnetic properties, broadening its application in magnetic resonance imaging (MRI) [\[26\]](#page-26-6).

While still in its nascent stage, the study of  $2D$  MoS<sub>2</sub> has made notable strides in biomedicine in recent years. Given that a comprehensive review of 2D  $MoS<sub>2</sub>$ for synergistic applications in imaging, therapy, and drug delivery remains elusive, this review aims to fll that gap by summarizing relevant advancements. In this context, we summarize the synthetic strategies utilized and elucidate the properties retained following surface modifcation. Furthermore, we emphasize the potential biomedical applications of these properties, with a particular focus on bioimaging and tumor therapy. Future prospects and challenges are also discussed, aiming to chart a course toward the development of more efficacious 2D  $MoS<sub>2</sub>$ -based cancer theranostic strategies.

#### **Evaluation of 2D MoS<sub>2</sub> performance in nanomedicine**

Unlike early 2D materials such as graphene, which is hampered by high toxicity, black phosphorus that is limited by instability, and hexagonal boron nitride that is limited by its insulating nature and lack of strong NIR absorption, 2D MoS<sub>2</sub> displays minimal toxicity and robust chemical stability [\[27](#page-26-7)[–29\]](#page-26-8). Its outstanding optical properties, catalytic functionality, and high specifc surface area, coupled with the advantages of a small size for drug delivery, show signifcant potential in cancer theranostics [[30\]](#page-26-9). Additionally, 2D  $MoS<sub>2</sub>$  is amenable to scalable, high-quality synthesis, making it a promising candidate for transformation [\[31](#page-26-10)]. However, the practical application of 2D  $MoS<sub>2</sub>$  in tumor diagnosis and treatment faces considerable challenges. The optimization of synthesis methods is crucial for producing biomedical-grade 2D  $MoS<sub>2</sub>$  to increase its clinical scalability. Improvements in therapeutic efficacy and safety are also necessary. These issues can be partially addressed through surface modifcations, which can impart greater biocompatibility, physiological stability, and targeting ability to 2D  $MoS<sub>2</sub>$  [[32\]](#page-26-11). We evaluate  $2D$  MoS<sub>2</sub> from the perspectives of synthesis, inherent characteristics, and tolerance properties, aiming to align its biomedical specifcations with the requirements of cancer theranostics.

#### **Scalability**

Different synthetic strategies produce 2D  $MoS<sub>2</sub>$ nanomaterials with distinct properties [\[33\]](#page-26-12). Currently, various synthesis strategies have been investigated with two main approaches: top-down strategies, including mechanical exfoliation, chemical exfoliation and liquidphase exfoliation (LPE), and bottom-up strategies, including chemical vapor deposition (CVD) and hydrothermal or solvothermal methods [[34–](#page-26-13)[37](#page-26-14)]. Topdown strategies involve peeling or etching large-sized  $MoS<sub>2</sub>$  into mono- or few-layered nanosheets. In contrast, bottom-up strategies refer to the process of assembling small building blocks (atomic or molecular building blocks) into relatively large nanoscale  $MoS<sub>2</sub>$  [[38\]](#page-26-15). In the following sections, we briefy describe these classic synthetic methods while considering their feasibility and limitations from a biomedical perspective.

Physical exfoliation, or mechanical exfoliation, is a typical top-down method used to synthesize  $2D$  MoS<sub>2</sub>. In this process, a tape and a substrate are usually required to peel and hold thin flms, respectively. Due to the weak van der Waals force between layers,  $2D$  MoS<sub>2</sub> is easily exfoliated from a bulk crystal, adhered to the tape, transferred to and retained on the appropriate substrate [[39\]](#page-26-16). The simplicity of the operation and the cleanness of the obtained 2D  $MoS<sub>2</sub>$  inspired physical exfoliation as a promising synthetic strategy, but it has not been popularized because the uncontrollability of the peel size or thickness and the low production yield limit its use in biomedical applications [[33,](#page-26-12) [40](#page-26-17)].

Chemical exfoliation, achieved through the intercalation of ions or molecules, is another method used

to produce 2D  $MoS<sub>2</sub>$  [\[35\]](#page-26-18). Among various candidates, alkali metals, particularly lithium (Li), are prominent because of their high reduction potential, reactivity, and mobility within the  $MoS<sub>2</sub>$  layers [\[41](#page-26-19)]. Morrison et al. submerged  $MoS<sub>2</sub>$  powder in n-butyllithium to prepare Li ion (Li<sup>+</sup>)-intercalated MoS<sub>2</sub>. Following the transfer of the intercalated material into water, the Li in the gaps reacts vigorously with the water to evolve hydrogen, resulting in the adjacent layers separating and dispersing in the water  $[42]$  $[42]$ . Although high throughput can be achieved by successive intercalation, the conventional Li intercalation method is time-consuming and requires water-free conditions, which limits its universality [\[38](#page-26-15), [43\]](#page-26-21). Fortunately, some methods, such as microwave/ ultrasonication-assisted intercalation, have accelerated the reaction process and improved efficiency  $[44]$  $[44]$ . In addition, the toxicity of residual  $Li<sup>+</sup>$  is a serious concern, especially in biomedical applications  $[38]$ . Therefore, several rational non-Li intercalants, such as potassium ions and ammonia/ammonium ions, have been explored for more secure applications [\[45](#page-26-23), [46](#page-26-24)]. Importantly, the use of some natural polymers and biomacromolecules, such as chitosan (CS) and bovine serum albumin (BSA), as intercalants could improve the dispersity and biocompatibility of 2D  $MoS<sub>2</sub>$  in biological systems [\[47](#page-26-25), [48\]](#page-26-26).

LPE proceeds in three stages: bulk  $MoS<sub>2</sub>$  slab dispersion in a specifc solvent, ultrasonication, and centrifugation  $[49]$ . Due to the acoustic cavitation effects,  $MoS<sub>2</sub>$  slabs are successfully exfoliated into nanosheets upon ultrasonication. In addition, appropriate solvents can provide shear force and weaken the interactions between layers, which commits to the preparation of 2D  $MoS<sub>2</sub>$  dispersions in collaboration with ultrasonication [[50\]](#page-26-28). However, commonly used dispersants such as N-methylpyrrolidone present challenges in removal because of the strong adsorption mediated by the specifc surface energy of  $2D$  MoS<sub>2</sub> and the high boiling point of organic solvents [\[51,](#page-26-29) [52\]](#page-26-30). Not only could this strong adsorption lead to fake deposition, but the toxicity of these solvents limits the broad application of 2D  $MoS<sub>2</sub>$ , especially in the context of biomedicine [[50,](#page-26-28) [52,](#page-26-30) [53](#page-26-31)]. Strikingly, the use of easy-to-remove, low-toxicity or even nontoxic surfactants such as deoxyribonucleic acid (DNA)/ribonucleic acid (RNA) nucleotides has partly solved this problem, achieving the safe and efficient production of nanosheets, especially for biomedical purposes [\[54](#page-26-32)].

CVD is a typical bottom-up method for the lowcost and large-scale production of  $MoS<sub>2</sub>$  monolayers [[55\]](#page-26-33). In this process, the source materials, generally sulfur powder and Mo-based oxide/chloride, are frst sublimated such that the precursor reagents are in the gas state. After being transported through the inert carrier gas, the reactant vapor then difuses to the substrate and adsorbs onto the substrate surface. The adsorbed atoms (Mo and S) subsequently difuse along the substrate surface and react with each other to form films [\[56](#page-26-34)]. Unfortunately, the difficulty in transferring 2D  $MoS<sub>2</sub>$ into physiological solutions and its instability under physiological conditions limit its biomedical applications [[38,](#page-26-15) [57](#page-26-35)].

As one of the simplest synthesis methods, the hydrothermal/solvothermal method has received much attention. This process involves  $2D$  MoS<sub>2</sub> crystallization from an aqueous/organic solution after the hydrothermal reaction between source materials [\[58](#page-26-36)]. Generally, the reaction is conducted in a Teflon-lined stainless-steel autoclave at moderate temperature and high pressure with Mo and S precursor solutions as reactants, which are usually S-containing salt/organic and Mo-containing salt/oxide. As the reaction ends and the reaction system cools down, 2D  $MoS<sub>2</sub>$  crystallizes [\[33](#page-26-12), [41](#page-26-19)]. It is important to note that this preparation method may not be universally applicable, as specifc adjustments might be required depending on the reactants and desired properties of the fnal product. Due to its ability to grow high-quality, uniform, pure and even biocompatible  $2D$  MoS<sub>2</sub> materials, the hydrothermal/solvothermal method has been widely adopted and is the choice in the biomedical feld [\[38](#page-26-15), [41\]](#page-26-19). For instance, with ammonium tetrathiomolybdate as the precursor, Wang et al. prepared PEGylated 2D  $MoS<sub>2</sub>$  via a solvothermal reaction in a PEG-400 aqueous solution  $[24]$  $[24]$  $[24]$ . Notably, PEGylated nanosheets have high colloidal stability and biocompatibility, indicating their excellent application value in biomedicine.

Scalability is a crucial factor when considering practical clinical applications. The synthesis of 2D  $MoS<sub>2</sub>$ must be feasible on a large scale while maintaining consistency and quality. Chemical exfoliation ofers higher throughput, especially with advancements such as microwave/ultrasonication-assisted intercalation, which improve the efficiency and scalability  $[35]$  $[35]$  $[35]$ . LPE is also scalable, particularly when nontoxic surfactants are used to simplify solvent removal and reduce toxicity, making it viable for industrial applications [\[54](#page-26-32)]. CVD can produce large-area 2D  $MoS<sub>2</sub>$  at low cost and is suitable for industrial applications, although it requires modifcations for stability in physiological solutions [\[55](#page-26-33)]. The hydrothermal/solvothermal method is noted for its ability to produce high-quality, uniform 2D  $MoS<sub>2</sub>$  under moderate conditions, making it cost-efective and highly efficient for large-scale production  $[41]$  $[41]$ . Thus, CVD and hydrothermal/solvothermal methods are promising for scalable production for consistent and reproducible theranostic applications, while improved chemical exfoliation and LPE techniques also show potential.

#### **Toxicity and biosafety**

To date, toxicity remains a critical obstacle hindering the widespread application of 2D nanomaterials. Even though 2D  $MoS<sub>2</sub>$  is less hazardous than some other materials, such as graphene and its analogs, its toxicity cannot be overlooked [[59,](#page-26-37) [60\]](#page-26-38). To this end, toxicological studies have demonstrated that 2D  $MoS<sub>2</sub>$  has mild toxicity, which is primarily linked to increased oxidative stress and physical disruption. Oxidative stress is a major contributor to nanotoxicity, such as organelle damage and cell death [\[61](#page-26-39)]. A study of Kupfer cells revealed that the dissolution of nanosheets and the release of hexavalent Mo contribute to mitochondrial reactive oxygen species (ROS) generation, inducing caspase 3/7-mediated apoptosis. This effect is dose dependent, with statistically signifcant reductions in cell viability observed at concentrations greater than 25 μg/mL [[62\]](#page-27-0). Importantly, even chronic low-dose  $(1 \mu g/mL)$ exposure (7 days) can induce severe toxic efects. HaCaT keratinocytes exhibited a loss of cell membrane integrity, mitochondrial dysfunction, endoplasmic reticulum (ER) disorder, and nuclear damage after chronic exposure. These changes are attributed to the disruption of cell membrane integrity due to high cellular internalization and increased oxidative stress caused by faster electron transfer from  $MoS<sub>2</sub>$ , which seizes electrons from the mitochondrial membrane, disturbing normal electron transport. The function of cell organelles is subsequently compromised, resulting in a failure to maintain normal metabolic activity [\[63\]](#page-27-1). ROS generation induced by chemically exfoliated  $MoS<sub>2</sub>$  also mediates DNA cleavage, another key mechanism of nanotoxicity [\[64](#page-27-2)]. In addition to oxidative stress induction, 2D  $MoS<sub>2</sub>$  can directly cause physical damage to cells and organelles. For example, the internalization of 2D  $MoS<sub>2</sub>$  disrupts cell membrane integrity, as mentioned above [\[63](#page-27-1)]. The 2D  $MoS<sub>2</sub>$  can penetrate the mitochondrial lipid membrane through hydrophobic interactions, causing heterogeneous lipid packing and resulting in damage [[65\]](#page-27-3). In addition to intrinsic toxicity, the distribution state of 2D  $MoS<sub>2</sub>$  is also a significant factor influencing its toxicity. Wang et al. showed that aggregated  $MoS<sub>2</sub>$ induced strong proinfammatory and profbrogenic responses in vitro and acute lung infammation in mice, whereas exfoliated  $MoS<sub>2</sub>$  had little or no effect [\[22](#page-26-2)]. This finding underscores the importance of further modifcation and functionalization not only to increase the biocompatibility of 2D  $MoS<sub>2</sub>$  itself but also to reduce aggregation and increase physiological stability to improve safety.

Given that the inherent toxicity of 2D  $MoS<sub>2</sub>$  and damage, such as infammation caused by nanosheet aggregation, can highlight the concern of wide bioapplication and clinical translation, optimizing the biocompatibility and physiological stability of 2D  $MoS<sub>2</sub>$ remains essential  $[22, 62, 66]$  $[22, 62, 66]$  $[22, 62, 66]$  $[22, 62, 66]$  $[22, 62, 66]$  $[22, 62, 66]$  $[22, 62, 66]$ . In this respect, many polymers or biomimetic molecules, such as PEG, CS and lipids, have been extensively used to functionalize 2D  $MoS<sub>2</sub>$  for increased biosafety. PEG is considered the preferred polymer in drug delivery systems because of its structural fexibility, amphiphilicity, and especially well-established safety profle [[67\]](#page-27-5). Hao et al. prepared  $MoS<sub>2</sub>-PEG$  by anchoring lipoic acid-conjugated PEG (LA-PEG) to the surface defect sites of 2D  $MoS<sub>2</sub>$  via Mo-S bonding [\[68](#page-27-6)]. As expected, PEG provided 2D  $MoS<sub>2</sub>$  with enhanced biocompatibility. Severe toxicity in vitro and in vivo was not detected even at high concentrations, and further long-term toxicity within a reasonable dose range could be neglected in light of the almost complete clearance of  $MoS<sub>2</sub>-PEG$  within 30 d. Remarkably, PEGylation also greatly increased the physiological stability, with  $MoS<sub>2</sub>-PEG$  being fairly stable in diverse physiological solutions, including RPMI-1640 medium, phosphate-buffered saline (PBS), and fetal bovine serum. On the other hand, CS, a natural and abundant biopolymer, is usually used as a biocompatible agent [\[69](#page-27-7)]. Yin et al. introduced CS to the surface of 2D  $MoS<sub>2</sub>$  via physical interactions to synthesize  $MoS<sub>2</sub>-CS$ [[70\]](#page-27-8). MoS<sub>2</sub>-CS nanosheets are well dispersed in water and other physiological buffers and exhibit greater physiological stability and biocompatibility than  $MoS<sub>2</sub>$ . Lipids have emerged as excellent candidates for surface coatings because of their ability to improve the solubility of drugs, their biodegradability, and their biocompatibility [\[71](#page-27-9)]. Building on this foundation, Xie et al. developed  $MoS_{2}$ -lipid nanocomposites [[72\]](#page-27-10). Lipid modifcation signifcantly improved the stability and biocompatibility of 2D MoS<sub>2</sub>. Unlike untreated MoS<sub>2</sub>, which exhibited severe coagulation in water, PBS, and cell culture media over 48 h,  $MoS<sub>2</sub>-lipid$  maintained excellent dispersibility and stability in all three environments. Additionally, lipid modifcation increased the hydrophilicity of  $MoS<sub>2</sub>$ , resulting in a much lower BSA adsorption rate than that of unmodified  $MoS<sub>2</sub>$ , thus indicating the enhanced biocompatibility of  $MoS_{2}$ -lipid.

#### **Targeting**

As targeting is the functional basis of efficient and safe tumor diagnosis and treatment, achieving precise targeting should always be our goal [[73\]](#page-27-11). Due to the small size of ordinary 2D  $MoS<sub>2</sub>$ , targeting is achieved primarily through the EPR effect, also referred to as passive targeting [[74](#page-27-12)]. Furthermore, modifcation with

adaptive biological molecules such as PEG can enhance the passive targeting capability of  $2D$  MoS<sub>2</sub>. This expectation is reasonable since PEGylation helps reduce reticuloendothelial system uptake, thus improving accumulation in the tumor [[75\]](#page-27-13). However, relying solely on the EPR efect is inadequate in practice, whereas smart and precise targeting is sought. This demand has encouraged the development of active targeting and response targeting methods based on high-specifcity targeting ligands and sensitive stimulus-responsive elements, respectively [[76](#page-27-14), [77](#page-27-15)]. Currently, targeting ligands, including peptides such as arginine–glycine– aspartate (RGD) and small molecules such as FA and hyaluronic acid (HA), have been widely utilized to decorate 2D  $MoS<sub>2</sub>$  [\[78–](#page-27-16)[80\]](#page-27-17). The corresponding receptors are typically overexpressed on tumor cells instead of healthy cells, thereby presenting tumor specifcity and conferring tumor targeting to 2D  $MoS<sub>2</sub>$ . For example, RGD selectively binds to ανβ3 integrin, which is commonly overexpressed in tumor cells such as HeLa cells. Additionally, the FA receptor is upregulated in various types of epithelial cancers and 90% of ovarian carcinomas and is expressed at low levels in healthy tissues. HA can specifcally bind to cluster determinant 44 (CD44), which is overexpressed on various tumor cell surfaces. Guided by these ligands,  $2D$  MoS<sub>2</sub> attaches to tumor cells expressing the corresponding receptors for internalization, enabling improved targeting and bioavailability [\[81](#page-27-18), [82\]](#page-27-19). In another dynamic targeting strategy,  $2D \text{ MoS}_2$  is modified with responsive molecules or chemical bonds such as HA and disulfde bonds (S–S). These modifications respond to internal (pH, enzymes, redox, etc.) or external (light, heat, ultrasound, etc.) stimuli for effective targeting  $[80-83]$  $[80-83]$  $[80-83]$ . For example, hyaluronidase and glutathione (GSH), which are abundant in the tumor microenvironment (TME) but absent in normal tissues, can efficiently cleave HA and reduce S–S. This property endows 2D  $MoS<sub>2</sub>$  with responsive targeting and the precise release of loaded substances from  $MoS_2-HA$  or  $MoS_2-SS$  as carriers.

#### **Approaches for single‑mode imaging and therapy**

The early diagnosis of cancer and subsequent effective therapies are undeniably critical for a good prognosis [[84\]](#page-27-21). Various noninvasive bioimaging techniques, such as MRI, photoacoustic imaging (PAI), and fuorescence imaging (FI), can be employed for early diagnosis  $[85-87]$  $[85-87]$  $[85-87]$ . Through the development of tailored imaging agents, these powerful techniques have demonstrated efficacy in the identification and characterization of diseases in their nascent stages [[88](#page-27-24)]. Due to their unique optical and photothermal properties and additional or enhanced characteristics conferred by loading, 2D

 $MoS<sub>2</sub>$  and its compounds have displayed immense potential in these imaging modalities as imaging agents. In addition to imaging applications,  $2D$  MoS<sub>2</sub> is also a promising therapeutic agent for cancer treatment, especially as a photothermal agent (PTA), due to its excellent photothermal properties [[57\]](#page-26-35). Furthermore, 2D  $MoS<sub>2</sub>$  has been investigated as an outstanding catalyst for therapeutic interventions because of its high surface area, abundant defects, and sulfur vacancies [\[89,](#page-27-25) [90](#page-27-26)]. In this work, we present the advantages of 2D  $MoS<sub>2</sub>$ in approaches for single-mode imaging and therapy, including FI, PAI, PTT and catalytic therapy.

#### **2D MoS2 in fuorescence imaging**

FI has attracted considerable interest and has emerged as a rapidly evolving and promising imaging method in the biomedical context  $[91]$  $[91]$ . Its attractiveness lies in its high sensitivity, impressive resolution, quick feedback, and substantial safety due to its noninvasive nature and absence of ionizing radiation [[92,](#page-27-28) [93](#page-27-29)]. These advantageous characteristics have positioned FI as an indispensable tool in the exploration of various biological phenomena. Notably, the unique inherent PL or fluorescence characteristics of 2D  $MoS<sub>2</sub>$  make them an optimal choice for FI. Building on these fndings, Qi et al. utilized 2D  $MoS<sub>2</sub>$  as a fluorescent label for imaging HepG2 cells [\[94](#page-27-30)]. Their experiments revealed a crucial finding: numerous  $2D$  MoS<sub>2</sub> molecules with strong fuorescence were internalized into cells. When the cells were irradiated with broad-band excitation light, specifcally UV (300–400 nm), blue (400–500 nm), and green (500–600 nm) light, the fuorescently labeled cells appeared blue, green, and red, respectively, in the fluorescence images. The cells exhibited vibrant colors corresponding to the specifc wavelength of light used, which allowed easy diferentiation. Additionally, they showed clear morphologies in the bright feld image, highlighting the effectiveness of 2D  $MoS<sub>2</sub>$  as a tool for detailed cellular studies.

Theoretically, coupling these strongly fluorescent 2D  $MoS<sub>2</sub>$  with fluorescent dyes may result in a dual fluorescence effect. More importantly, the use of 2D  $MoS<sub>2</sub>$ as a carrier enhances the accumulation of fuorescent dyes through the EPR efect, thereby potentially generating greater efectiveness. Unfortunately, the fuorescence emission quenching or decreasing property of  $2D$  MoS<sub>2</sub> renders that phenomenon invalid [[95\]](#page-27-31). Wang et al. solved this problem to some extent through an "intermediary" strategy [[96](#page-27-32)]. In this frontier work,  $MoS<sub>2</sub>$ -PEG was embedded into mesoporous silica nanoparticles mesoporous silica nanoparticles (MSNs), playing the role of an "intermediary", and then the aggregation-induced emission fluorogen PhENH<sub>2</sub> was chemically grafted onto the MSNs, triggering

robust fuorescence emission (Fig. [1](#page-6-0)A, B). FA was also linked to the surface of the MSNs, leading to the synthesis of  $PhENH_{2}-MoS_{2}-FA$  MSNs. Interestingly, after an incubation with  $PhENH<sub>2</sub>-MoS<sub>2</sub>-FA$  MSNs, MDA-MB-231 cells, which highly express FA receptors, exhibited potent red fluorescence. This fluorescence intensity notably surpassed that of HepG2 cells, which lack FA receptors, emphasizing the targeting efficacy of these nanoparticles.

In addition to directly targeting tumor cells for imaging, 2D  $MoS<sub>2</sub>$  can also be used for molecular detection imaging, allowing researchers the ability to distinguish tumor cells from normal cells. Due to oxidative metabolism disorders, more hydrogen peroxide  $(H_2O_2)$  accumulates in tumor cells than in normal cells [[97](#page-27-33)]. These findings indicate that  $H_2O_2$  and  $H_2O_2$ -induced oxidative stress could serve as markers for tumor cells. Based on this information, Liu et al. layered 2D  $MoS<sub>2</sub>$  with ortho-phenylenediamine (OPD) to quantify the intracellular concentrations of  $H_2O_2$  and thus distinguished cells under pathological conditions [[98\]](#page-27-34). During this procedure, 2D  $MoS<sub>2</sub>$ , which possesses



<span id="page-6-0"></span>Fig. 1 A Schematic diagram of the preparation of PhENH<sub>2</sub>-MoS<sub>2</sub>-FA MSNs and their fluorescence effect. **B** Fluorescence spectra of PhENH<sub>2</sub>-MoS<sub>2</sub>-FA MSNs dispersed in water. C Schematic diagram of the sensing principle of AgNC@MoS<sub>2</sub> probe. **D** UV–Vis absorption spectra and synchronous scanning fluorescence spectra of AgNC@MoS<sub>2</sub> probe. Reproduced with permission [[96](#page-27-32)]. Copyright 2019, Elsevier. Reproduced with permission [[102\]](#page-27-35). Copyright 2020, Elsevier

high peroxidase (POD)-mimicking catalytic activity, decomposed  $H_2O_2$  into hydroxyl radicals (OH $\cdot$ ) to oxidize OPD. The resulting OPD oxide then emitted fluorescence with a maximum at  $\sim$  557 nm upon excitation at 450 nm. This light emission characteristic enabled a clear distinction between human foreskin fbroblasts and HeLa cells via intensity feedback (the fuorescent signal was low for the former but higher for the latter) [\[98,](#page-27-34) [99](#page-27-36)].

Although the fuorescence quenching property of 2D  $MoS<sub>2</sub>$  is regrettable on direct imaging, as mentioned above, encouragingly, this property is used in the "switching" strategy to provide a higher target-tobackground ratio for FI. Briefly, the use of 2D  $MoS<sub>2</sub>$  as a quencher maintains the fuorescence in an "of" state in circulation, while the signal turns to an "on" state with the degradation of nanoparticles mediated by some specifc molecules and the consequent release of the dye at the target site [[100](#page-27-37)]. Building on this strategy, Jia et al. grafted an adenosine triphosphate (ATP) aptamer (Apt) labeled photosensitizer, chlorine e6 (Ce6), onto 2D  $MoS<sub>2</sub>$ for intracellular ATP imaging. Upon internalization into cancer cells, these compounds accumulate in ATP-rich lysosomes. Here, the intracellular ATP binds to the Apt, resulting in the release of Ce6-Apt and consequently the restoration of Ce6 fuorescence [[101\]](#page-27-38). Furthermore, Xu et al. synthesized a  $AgNC@MoS$ , probe by assembling fuorescent DNA-silver nanoclusters (DNA-AgNCs) with the ATP Apt as a template and LA-PEG-FA on 2D  $M_0S_2$ (Fig. [1](#page-6-0)C)  $[102]$  $[102]$  $[102]$ . This method allows positive targeting imaging and a quantitative analysis of intracellular ATP concentrations (Fig. [1](#page-6-0)D). In practical terms, when  $AgNC@MoS<sub>2</sub>-PEG-FA$  is taken up by tumor cells overexpressing the FA receptor, the recognition of ATP by the probe triggers the release of DNA-AgNCs and the recovery of fluorescence. The released DNA-AgNCs can be measured via inductively coupled plasma-mass spectrometry. This step is crucial, as it provides accurate quantitative information and enables a quantitative analysis of intracellular ATP concentrations. In summary,  $2D$   $MoS<sub>2</sub>$  offers significant potential for direct cellular imaging and molecular monitoring. However, the application of 2D  $MoS<sub>2</sub>$  in clinical FI remains elusive primarily due to the need for enhancements in brightness, photostability, and safety, necessitating further efectiveness and biocompatibility modifcations.

#### **2D MoS2 in photoacoustic imaging**

As an emerging imaging modality, PAI has attracted much attention because of its excellent superiority with respect to higher resolution than traditional MRI, deeper penetration than optical imaging, and lack of radiation exposure [\[103,](#page-27-39) [104](#page-28-0)]. When exposed to pulsed light, instant thermoelastic expansion

occurs in the photoabsorption tissue and creates an ultrasound wave, which is subsequently detected by an ultrasonic transducer and converted into an image [[105\]](#page-28-1). Benefting from the pronounced light absorbance, high photothermal conversion rate, ability to produce ultrasonic waves and exceptional resolution improved by the generation of air microbubbles,  $2D$  MoS<sub>2</sub> is believed to be a promising contrast agent for PAI [[12](#page-25-10), [106](#page-28-2)]. In this context, Chen et al. prepared single-layer 2D  $MoS<sub>2</sub>$  nanosheets and explored their potential for augmenting photoacoustic  $(PA)$  signals  $[107]$  $[107]$ . The nanosheets were efficiently internalized into glioma cells. This internalization resulted in strong PA signals with an impressively sensitive detection limit of approximately 100 cells. Encouraging outcomes were also observed in subcutaneous and orthotopic tumor-bearing mice. The tumor tissue nearly 1.5 mm below the skull was clearly visualized. Nevertheless, relying solely on simple  $2D$  MoS<sub>2</sub> when encountering gliomas at even deeper sites is insufficient, necessitating the utilization of more advanced contrast agents. For this reason, Liu et al. introduced indocyanine green (ICG), a dye with a high NIR extinction coefficient, onto the surface of 2D  $MoS<sub>2</sub>$ , obtaining MoS<sub>2</sub>-ICG with higher sensitivity  $[108, 109]$  $[108, 109]$  $[108, 109]$  $[108, 109]$ . This adjustment provided clear benefits. The absorption peak (800 nm) of  $MoS<sub>2</sub>-ICG$  was significantly greater than that of  $2D$  MoS<sub>2</sub> (675 nm). This shift allowed PAI to be conducted at longer wavelengths, which could lower the background noise and increase the penetration depth. Remarkably, the PA signal observed upon 800 nm laser excitation of  $MoS<sub>2</sub>$ -ICG was approximately 16-fold greater than that of  $MoS_{2}$ . This enhancement exhibited a concentration-dependent profle and was coupled with outstanding photostability. In practice, this enhancement translated to an impressive imaging depth of up to 3.5 mm when  $MoS<sub>2</sub>-ICG$  was used for in vivo PAI of orthotopic gliomas in the brain. This depth was almost twice that achieved using  $2D$  MoS<sub>2</sub>, suggesting the significant potential of  $MoS<sub>2</sub>$ -ICG in PAI applications. In summary, the advancements obtained in PAI applications using 2D  $MoS<sub>2</sub>$  and  $MoS<sub>2</sub>$ -ICG highlight the critical role of contrast agents in enhancing the capabilities of PAI. While 2D  $MoS<sub>2</sub>$  shows remarkable efficacy in visualizing tumor tissues near the surface, its limitations at greater depths necessitate the exploration of modifed agents such as  $MoS<sub>2</sub>$ -ICG. This finding underscores the importance of continuous optimization to meet demands for higher-resolution imaging, particularly in deep tissue imaging.

#### **2D MoS2 in photothermal therapy**

PTT is a technique that utilizes the principle of converting absorbed light into heat, thereby inducing

thermal damage in the tumor [[110\]](#page-28-6). In this process, PTAs possess excellent photothermal properties and safety and are critical factors for achieving high therapeutic efficacy [[111\]](#page-28-7). Among various PTAs, 2D MoS<sub>2</sub> stands out because of its admirable optical absorption, photothermal conversion efficiency, and biocompatibility.

In a recent study, Chen et al. synthesized PEGylated  $MoS<sub>2</sub>$  nanosheets  $(MoS<sub>2</sub>-PPEG)$  with poly(acrylic acid) (PAA), resulting in the ability of the obtained nanosheets to degrade (Fig. [2](#page-8-0)A) [[112](#page-28-8)]. Notably, the  $MoS<sub>2</sub>-PPEG$  demonstrated exceptional stability and photothermal properties. For example, upon 300 s of NIR laser irradiation, the temperature of the  $MoS<sub>2</sub>-PPEG$ dispersion at a concentration of 200  $\mu$ g/mL increased from 25.8 to 51.4 °C, whereas the corresponding temperature change for water was only 1.3 °C. In subsequent in vivo investigations, the temperature of  $MoS<sub>2</sub>-PPEG$  reached 52.1 °C, whereas the temperature of PBS increased approximately 1 °C. Intriguingly, while the tumor volume of the mice treated with  $MoS<sub>2</sub>-PPEG$ or NIR laser irradiation alone continuously increased throughout the detection period, PTT with  $MoS<sub>2</sub>-PPEG$ dramatically suppressed tumor growth (Fig. [2](#page-8-0)B). Moreover, the body weights of the mice remained stable throughout the treatment period, suggesting a high tolerance to PTT (Fig. [2](#page-8-0)C).

Enhanced targeting modifcations have been engineered to ensure high specifcity in recognizing and efficiently targeting tumor cells. Nucleic acid Apts, which are single-stranded oligonucleotides, exhibit excellent



<span id="page-8-0"></span>Fig. 2 A Schematic diagram of MoS<sub>2</sub>-PPEG preparation and in vivo PTT. Relative tumor volume (B) and body weight (C) of mice under different treatments. **D** Schematic diagram of precise PTT with APT modified 2D MoS<sub>2</sub>. **E** Relative tumor volume of mice under different treatments. Reproduced with permission [[112](#page-28-8)]. Copyright 2017, American Chemical Society. Reproduced with permission [[114\]](#page-28-9). Copyright 2021, Elsevier

specificity and affinity for their target molecules [\[113\]](#page-28-10). In related work, Pang et al. modified 2D  $MoS<sub>2</sub>$  with aminonucleic Apt and BSA, creating  $MoS<sub>2</sub>-BSA-Apt$  noted for its precise targeting ability and favorable biocompatibility (Fig. [2D](#page-8-0)) [\[114](#page-28-9)]. When exposed to 808 nm NIR laser irradiation, these nanosheets demonstrated a potent ability to ablate tumors. Remarkably, following the injection of  $MoS<sub>2</sub>-BSA-Apt$  and subsequent 808 nm laser irradiation for 10 min, the breast tumors in the mice were entirely eliminated after 14 days of PTT. This fnding contrasted sharply with the outcomes observed in mice treated with 2D  $MoS<sub>2</sub>$  and laser irradiation alone, which failed to efectively inhibit the growth of tumors (Fig. [2](#page-8-0)E). Furthermore, Liu et al. prepared CD44-targeted 2D MoS<sub>2</sub>, referred to as the MoS<sub>2</sub>@ protein A@CD44 antibody (Ab) [[115](#page-28-11)]. This complex was achieved by coating anti-CD44 Ab on the 2D  $MoS<sub>2</sub>$ surface through the connection of a layer of Protein A mixed with bovine BSA. Upon cellular uptake, CD44 targeted  $MoS<sub>2</sub>$  led to the depletion of membrane CD44 in lysosomal degradative compartments, resulting in a decrease in RhoA activation (GTP-RhoA) and its downstream epithelial–mesenchymal transition (EMT)related transcription factors, thereby weakening tumor progression. This result was also expected, as CD44 is known to induce the EMT, a process during which tumor cells lose their original epithelial phenotype and acquire a mesenchymal phenotype characterized by increased stemness, chemoresistance, and invasiveness [\[116](#page-28-12)]. Moreover, PTT utilizing  $MoS<sub>2</sub>@CD44Ab$  induced an increase in the intracellular temperature, which disrupted the activation of these signals and further hampered the biological behavior of the tumor.

Enhanced photothermal therapeutic efficacy is anticipated with the combination of additional PTAs and 2D  $MoS<sub>2</sub>$ , with the potential of heterojunction architectures to enhance the efficient separation and migration of photoexcited charges [\[117\]](#page-28-13). Carbon dots (CDs) have been extensively investigated due to their advantages, such as their excellent safety profle and tunable photochemical properties [\[118\]](#page-28-14). Despite the lower photothermal efficiency of CDs, their heterojunctions combined with 2D  $MoS<sub>2</sub>$  have exhibited a markedly enhanced photothermal efect compared with that of single 2D  $MoS<sub>2</sub>$ . Expanding on this concept, Geng et al. reported 0D/2D/0D sandwich heterojunctions, such as  $NIR-CD/MoS<sub>2</sub>$ , with 0D-N-doped CDs and 2D  $MoS<sub>2</sub>$  nanosheets [[119\]](#page-28-15). The results revealed that the absorbance and photothermal conversion efficiency were remarkably greater than those of the pristine NIR-CDs and  $MoS<sub>2</sub>$ . This enhanced performance was also evident in in vivo studies, where the temperature of the tumors in the NIR-CD/MoS<sub>2</sub> group rapidly

increased upon exposure to 808 nm laser irradiation, resulting in complete inhibition of 4T1 tumor growth and no occurrence of pulmonary metastatic nodules at an ultralow power density. Conversely, the tumors in the saline, single NIR-CD/MoS<sub>2</sub>, and saline with laser irradiation groups rapidly increased in size within 18 days, and a considerable number of metastatic nodules was observed. Collectively, these examples exemplify the satisfactory efficacy of 2D  $MoS<sub>2</sub>$  and its nanocomposites in PTT. Each study contributes uniquely to the feld, either through advancements in biocompatibility modifcations, targeting strategies, or the development of new heterojunction architectures. Future research should continue to explore additional optimizations, further improving the therapeutic outcomes of PTT.

#### **2D MoS2 in catalytic therapy**

Nanocatalytic therapy has recently emerged as a promising alternative for tumor treatment, leveraging nanozymes to induce catalytic reactions in vivo that generate abundant ROS. These ROS have the potential to signifcantly impair and even annihilate malignant cells  $[120]$  $[120]$ . Among these nanozymes, 2D MoS<sub>2</sub>, which functions similarly to POD, ofers a stable and costefective platform for catalytic activity with outstanding and adjustable catalytic activity in vivo [\[121](#page-28-17), [122](#page-28-18)].

Recently, Wang et al. highlighted an innovative nanocatalyst,  $BTO/MoS<sub>2</sub>QCA$ , which responds to the acidic conditions of the TME and is activated by ultrasound (Fig.  $3A$  $3A$ ) [[123](#page-28-19)]. This nanocatalyst is constructed from few-layer  $MoS<sub>2</sub>$  nanosheets grown on the surface of piezoelectric tetragonal barium titanate (T-BTO) and is further modifed with pH-responsive cinnamaldehyde (CA). In the acidic TME, CA initiates the breakdown of endogenous molecules to release substantial quantities of  $H_2O_2$ . The generated  $H_2O_2$  is subsequently transformed into OH· through a series of reactions catalyzed by POD-like BTO/MoS<sub>2</sub> (Fig.  $3B$  $3B$ ). The effectiveness of this reaction is influenced by the ultrasound-induced microscopic pressure, which segregates the positive and negative charges generated by BTO, thereby increasing the interaction between  $MoS<sub>2</sub>$ and  $H_2O_2$  and leading to continuous OH $\cdot$  production (Fig.  $3C$ ). Furthermore, MoS<sub>2</sub> contributes to the depletion of GSH in cancer cells, intensifying oxidative stress. This stress reduces the levels of GSH-POD-4, disrupting the redox balance and inducing tumor cell ferroptosis (Fig.  $3D$ ). The in vivo studies underscore the superior antitumor efficacy of  $BTO/MoS<sub>2</sub>@CA$ under ultrasound. Compared with the control groups treated with PBS and  $CA +$ ultrasound, the BTO/MoS<sub>2</sub>@ CA+ultrasound group exhibited signifcant tumor growth inhibition, characterized by pronounced tumor



<span id="page-10-0"></span>Fig. 3 A Schematic diagram of BTO/MoS<sub>2</sub>@CA catalytic activity for tumor treatment. B Reaction scheme for H<sub>2</sub>O<sub>2</sub> decomposition and TMB oxidation. **C** Schematic illustration of the surface positive and negative charges distribution of BTO driven by ultrasound. **D** Schematic diagram of ferroptosis mediated by BTO/MoS<sub>2</sub>@CA. Reproduced with permission [\[123\]](#page-28-19). Copyright 2023, Wiley

necrosis, hemorrhage, and infltration of infammatory cells.

The high specific surface area and abundant surface active sites of 2D  $MoS<sub>2</sub>$  facilitate its role as a cocatalyst support for single-atom catalysis, increasing its potential in cancer catalytic therapy [[124\]](#page-28-20). In this context, Yang et al. developed a 2D composite nanocatalyst, termed  $MoS<sub>2</sub>@SA-Fe-PEG (MSFP)$ , by integrating atomically dispersed iron (Fe) onto a  $MoS<sub>2</sub>$  support and modifying the surface with amine-polyethylene glycol  $(NH_2-PEG)$ [ $125$ ]. This configuration harnesses the Fenton reaction, a well-known Fe-mediated redox process, to decompose  $H_2O_2$  into OH $\cdot$  at the atomic level [[126,](#page-28-22) [127\]](#page-28-23). The doped Fe atoms catalyze Fenton reactions, while the

abundant sulfur vacancy defects generated on both the fat surface and edge of the nanosheet after Fe doping increase the surface electron density and thereby favor electron capture by  $H_2O_2$  to promote OH· production. Importantly, the  $MoS<sub>2</sub>$  support serves as a cocatalyst, enhancing the reduction of  $Fe^{3+}$  to  $Fe^{2+}$  through its  $Mo^{4+}$  sites, thereby facilitating the Fenton process efficiently. Comparative studies underscore the superior catalytic anticancer capabilities of MSFP to traditional nanocatalysts. In contrast to the control group ( $Fe<sup>2+</sup>$  or  $MoS<sub>2</sub>-PEG treatment$ ), which exhibited significant tumor proliferation, the MSFP-treated group displayed distinct tumor suppression and prolonged lifetimes  $[125]$ . These studies illustrate signifcant advancements in the feld of

nanocatalytic therapy using 2D  $MoS<sub>2</sub>$ -based nanozymes. As an emerging feld, this topic warrants further in-depth research, including enhancing the catalytic efficiency, selectivity, specificity, and biocompatibility of these nanozymes. Future research should focus on the interplay between their intrinsic structures and external environments to optimize these systems and improve their safety and efficacy in clinical settings  $[128]$  $[128]$  $[128]$ .

#### **Synergistic strategies in imaging and therapy**

Despite recent achievements in cancer diagnosis and treatment, single imaging modalities and therapies often lack the required diagnostic precision and therapeutic efficacy, indicating a strong desire to realize multimodal imaging (MMI), combination therapy, and even highly integrated platforms [[129](#page-28-25)[–131](#page-28-26)]. In particular, due to its large surface area and fascinating photochemical properties,  $2D$  MoS<sub>2</sub> was developed as a promising platform for MMI and cancer treatment.

#### **2D MoS2 in multimodal imaging**

Importantly, limitations remain for each single imaging modality that must be improved, such as the restricted

tissue penetration of optical techniques, the radioactivity of CT, and the low sensitivity of MRI [[132](#page-28-27)]. Fortunately, MMI, which is founded on the aforementioned modalities, takes the best out of the worst, whereby the integration of unique strengths and the remedy for each other's shortcomings provide novel insights toward improving the imaging capability [[130\]](#page-28-28). In this regard, as a promising contrast agent,  $2D$  MoS<sub>2</sub> is adaptable to multiple imaging methods simultaneously, leading to more possibilities for the advancement of MMI.

Indeed, numerous MMI methods that combine two or more imaging techniques, such as FI, CT, MRI and PAI, have been extensively investigated in recent years [[130,](#page-28-28) [133](#page-28-29)]. Among these methods, the dual imaging technique based on  $2D$  MoS<sub>2</sub> has tremendous potential, considering the diverse combinations of intrinsic properties, such as excellent fuorescence properties and photothermal properties, and has emerged as a representative approach in MMI [[134](#page-28-30), [135](#page-28-31)]. For example, the dual-modality platform of  $MoS<sub>2</sub>-HA$  can serve as a fuorescer and PA contrast agent with both fuorescence and PAI capabilities (Fig. [4](#page-11-0)A) [\[136\]](#page-28-32). Subsequent in vivo bioimaging experiments revealed that the fuorescence



<span id="page-11-0"></span>Fig. 4 A Schematic diagram of theranostic HA-MoS<sub>2</sub> conjugates. B In vivo fluorescence imaging of HA-MoS<sub>2</sub> conjugates. C The PA amplitude enhancement of HA-MoS<sub>2</sub> conjugates compared to the control (PBS). **D** Schematic diagram of hydrogel loading MoS<sub>2</sub> nanosheets preparation and synergistic PTT/PDT for cancer. Tumor photographs (**E**), relative tumor volume (**F**) and body weight (**G**) of mice under diferent treatments. Reproduced with permission [[136](#page-28-32)]. Copyright 2018, Wiley. Reproduced with permission [[143](#page-28-33)]. Copyright 2022, Wiley

signal from  $MoS_{2}$ -HA was markedly observable in the primary tumor (Fig. [4](#page-11-0)B). In terms of the PA signals,  $MoS<sub>2</sub>-HA$  demonstrated an astounding 1860-fold increase on average compared with that of PBS, far exceeding the several 100-fold enhancements achieved by traditional PA agents (Fig. [4C](#page-11-0)) [\[136](#page-28-32), [137](#page-28-34)].

In addition, by leveraging its strong fuorescence quenching ability and broad absorption in the NIR region,  $2D$  MoS<sub>2</sub> has emerged as a versatile candidate for FI quenchers and PAI contrast agents, enabling multimodal molecular detection and imaging. Li et al. introduced an activatable nanoprobe  $(MoS<sub>2</sub>)@$ polydopamine (PDA)-PEG/peptide, MPPF) engineered for dual-mode near-infrared fuorescence (NIRF)/ ratiometric PAI to detect endogenous furin activity [[138\]](#page-28-35). The 2D  $MoS<sub>2</sub>$  undergoes a coating process with PDA and PEG, after which it is covalently functionalized with Cy7-labeled furin substrate peptides. The enzymatic cleavage of these peptides by furin triggers the release of Cy7 molecules from the MPPF nanoprobes, thereby restoring their fuorescence. Concurrently, this process disrupts the fuorescence resonance energy transfer from Cy7 to  $MoS<sub>2</sub>$  and facilitates the efficient removal of small Cy7 molecules from tumor tissues, resulting in a prompt decrease in the PA signal at 768 nm (PA768). In contrast, the PA signal at 900 nm (PA900), attributed to 2D  $MoS<sub>2</sub>$ , decreases gradually due to its substantial size and slow tumor clearance. This coordinated alteration in NIRF and ratiometric PA signals thus represents a novel method for the real-time visualization of endogenous furin activity. These examples underscore the critical role of  $2D$  MoS<sub>2</sub> as a MMI platform in advancing the precision of biomedical imaging and overcoming the limitations of traditional imaging modalities. These advancements are pivotal for the progression of personalized medicine, ensuring targeted and efective treatments.

#### **2D MoS2 in combination therapy**

The need for increased antitumor efficacy has led to a gradual shift from monotherapy to multimodal therapy. This trend further underscores the need to develop a complementary strategy aimed at achieving synergistic therapy with the desired efficacy  $[139]$ . In this context, we discuss a synergistic PTT/photodynamic therapy (PDT) strategy as an example. Importantly, drug delivery for combination therapy is not discussed here but will be addressed in the next section.

PDT is capable of generating ROS to kill cancer cells, especially in deep tissues  $[140]$  $[140]$ . This capacity significantly ofsets the limited efectiveness of PTT in these regions. Conversely, PTT has been proven to improve tumor oxygenation, a crucial advantage in countering the tumor hypoxia commonly observed after prolonged PDT [[141](#page-28-38),

[142](#page-28-39)]. Evidently, the integration of these therapies presents synergistic potential that exceeds the capabilities of each treatment. In a recent study, Qi et al. investigated the application of 2D MoS<sub>2</sub> in a hydrogel context  $[143]$ . This study developed an injectable polysaccharide hydrogel loaded with  $BSA-MoS<sub>2</sub>$  nanosheets for synergistic PTT/ PDT in breast cancer (Fig. [4D](#page-11-0)). The hydrogel enables the sustained presence of  $BSA-MoS<sub>2</sub>$  around tumor cells for prolonged therapeutic efficacy. Upon NIR irradiation,  $BSA-MoS<sub>2</sub>$  in the hydrogel demonstrates a remarkable ability to annihilate tumor cells via photothermal conversion. Additionally, under 808 nm laser irradiation, a large amount of ROS is generated within cells, augmenting the efficacy of the PDT modality in tumor management. Highlighting the synergistic impact of PTT/PDT, the weights of the tumors treated with the  $BSA-MoS<sub>2</sub>$  hydrogel and laser were significantly lower than those of the PBS group and even the BSA- $MoS<sub>2</sub>$ with laser group (Fig.  $4E-G$  $4E-G$ ). These findings underscore the significant potential and importance of 2D  $MoS<sub>2</sub>$ in multimodal treatment strategies. Such approaches could not only reduce the intensity of each treatment modality, thereby mitigating adverse reactions, but also significantly enhance the therapeutic efficacy compared with single-modality therapies. This finding supports a shift toward more personalized medical approaches.

#### **Synergistic strategies in cancer‑targeting drug delivery**

The high specific surface area of 2D  $MoS<sub>2</sub>$  makes it an excellent drug carrier. Through surface modification, the hydrophilicity, stability, and biocompatibility of  $2D$  MoS<sub>2</sub> can be improved, making it an ideal delivery platform that provides an efective solution for drug insolubility and the degradation of genes and immune reagents [[38](#page-26-15), [144](#page-28-40), [145](#page-28-41)]. Efective cancer treatment relies not only on the design of therapeutic agents but also on their ability to reach and penetrate tumor tissues efficiently. In this regard, 2D  $MoS<sub>2</sub>$  with enhanced stability and biocompatibility allows relatively long circulation times in the bloodstream, increasing the likelihood of reaching tumor sites. The small size of 2D  $MoS<sub>2</sub>$ enables it to exploit the EPR effect, allowing preferential accumulation in tumor tissues due to the characteristic leaky vasculature and poor lymphatic drainage of tumors [[146\]](#page-28-42). Within the TME, 2D  $MoS<sub>2</sub>$  can be efficiently taken up by tumor cells through endocytosis, followed by the subsequent release of drugs  $[147]$  $[147]$  $[147]$ . Moreover, 2D MoS<sub>2</sub> conjugated with targeting ligands can achieve increased tumor specificity and reduce off-target effects  $[148]$ . The transport oncophysics of 2D  $MoS<sub>2</sub>$  is still insufficiently described. Issues such as the fluid dynamics of  $2D$  MoS<sub>2</sub> and methods to overcome the extracellular matrix barrier



#### <span id="page-13-0"></span>**Table 1** Functionalized 2D MoS<sub>2</sub> for drug delivery and theranostic

Molybdenum disulfide (MoS<sub>2</sub>), polyethylene-glycol (PEG), arginine-glycine-aspartate (RGD), succinimidyl 3-[2-pyridyldithio] propionate (SPDP), doxorubicin (DOX), cytosine-phosphate-guanine (CpG), medium-sized MoS<sub>2</sub> (MM), low PEI0.8 k coverage (PL), erythrocyte (RBC), barium titanate (BT), polydopamine (PDA), folic acid (FA), gemcitabine (Gem), bovine serum albumin (BSA), polyethylenimine (PEI), lipoic acid (LA), curcumin (Cur), erlotinib (Er), hyaluronic acid (HA), 6-azidohexylguanidinerich α-helical polypeptide-decorated amino-modified MoS<sub>2</sub> (APM), programmed death-ligand 1 (PD-L1), PD-L1 siRNA (siPDL1), anti-PDL1 antibody (aPDL1), hyperbranched polyglycidyl (HPG), triphenylphosphonium-glibenclamide (TG), glucose-HPG functionalized MoS<sub>3</sub> (GPM), 5-(4-(diphenylamino) phenyl) furan-2-pyridine (TFPy), ionic liquid (IL), α-tocopheryl succinate (α-TOS), 1-methyl-tryptophan (1-MT), cisplatin (Pt), gambogic acid (GA), cadmium trioxide (Gd<sub>2</sub>O<sub>3</sub>), photothermal therapy (PTT), photodynamic therapy (PDT), photoacoustic imaging (PAI), piezo-catalytic therapy (PCT), computerized tomography (CT), magnetic resonance imaging (MRI)

require further investigation to improve drug delivery efficiency  $[149, 150]$  $[149, 150]$  $[149, 150]$  $[149, 150]$ . Undeniably, 2D MoS<sub>2</sub> has shown promising potential in cancer-targeting drug delivery. In the following paragraphs, we discuss the progress in drug delivery-based combination therapies (Table [1](#page-13-0)).

#### **Chemotherapy‑based combination therapy**

The atomically thin planar structure and extraordinary surface-area-to-volume ratio of 2D  $MoS<sub>2</sub>$  makes it an ideal drug carrier for drug delivery applications [\[38\]](#page-26-15). In addition, its combination with antitumor agents, such as doxorubicin (DOX), further endows 2D  $MoS<sub>2</sub>$  with the ability to treat diferent malignancies [[151\]](#page-29-4). In view of the photothermal efects and chemotherapeutic potential conferred by drug loading, the combination of these two modalities is anticipated to generate superior therapeutic outcomes.

In a pioneering work, Liu et al. documented the tremendous potential of 2D  $MoS<sub>2</sub>$  in drug delivery and cancer combination therapy [[79\]](#page-27-40). Specifcally, drug loading ratios of up to~239% for DOX were achieved by electrostatic attraction. This performance is a substantial advancement considering the typical range of 10–30% observed in conventional nanoparticle-based drug delivery systems. Following PEG modifcation, the resulting  $MoS<sub>2</sub>-PEG-DOX$  nanosheets exhibited enhanced biocompatibility, leading to a safer antitumor response. However, single injections of  $MoS<sub>2</sub>-PEG-DOX$ did not yield satisfactory therapeutic outcomes, as evidenced by slightly delayed tumor growth in the absence of NIR laser irradiation. In sharp contrast, the combination of chemotherapy and PTT with  $MoS<sub>2</sub>-PEG/$ DOX and NIR irradiation led to the dramatic inhibition of tumor growth [[19\]](#page-26-40).

Although  $MoS<sub>2</sub>-PEG$  has shown high biocompatibility and efficacy in chemo-photothermal therapy, the linear polymers represented by PEG could be subject to more rapid clearance from the blood circulation than their branched counterparts, potentially leading to inadequate treatment durability [[152\]](#page-29-5). Wang et al. developed branched hyperbranched polyglycidyl (HPG)-modifed  $MoS<sub>2</sub> (MoS<sub>2</sub>-HPG)$  by absorbing HPG onto the surface of  $MoS<sub>2</sub>$  to address this issue, and this molecule displayed excellent dispersion, stability, biocompatibility, and, most importantly, a prolonged in vivo circulation time. Additionally,  $MoS_{2}$ -HPG was utilized as a drug carrier

to deliver DOX, producing superior antitumor efects through chemo-photothermal therapy [[153\]](#page-29-11). Based on these findings,  $MoS<sub>2</sub>-HPG$  was further employed for the codelivery of DOX and chloroquine (CQ). Importantly,  $MoS_{2}$ -HPG exhibited a high degree of loading efficiency for DOX and CQ, with rates of 88.9% and 92.4%, respectively. Furthermore, the release of drugs from the nanosheets was signifcantly enhanced by laser irradiation, resulting in the efficient eradication of incubated multidrug-resistant HeLa cells [\[154](#page-29-12)]. Nonetheless, exogenous substances can ultimately be eliminated by the immune system, underscoring the need for endogenous materials such as erythrocyte (RBC) membranes to prolong the retention time in the blood and enhance therapeutic efficacy  $[155]$  $[155]$ . Li et al. designed an RBC membrane-camouflaged  $MoS<sub>2</sub>$ -based nanosystem for DOX delivery to address this issue [[156\]](#page-29-6). In this system, the RBC membrane contributes to the synergistic efect of chemotherapy and PTT by  $MoS<sub>2</sub>-DOX$ , enabling the nanoparticles to efectively enter tumor tissue with enhanced hydrophilicity, immune evasion capability and long circulation properties.

Tumor-specifc ligands such as FA, peptides and biotin were incorporated onto the surface of 2D  $MoS<sub>2</sub>$  to further improve the therapeutic efficacy and minimize unwanted efects. Murugan et al. constructed an FA-based targeting core–shell nanoparticle, MoS<sub>2</sub>@ barium titanate (BT)-PDA-FA (MBPF), with BT and PDA providing excellent biocompatibility  $[157]$ . This platform was then adapted to carry gemcitabine (Gem), amalgamating chemotherapy with PTT. Through FA receptor-mediated endocytosis, MBPF was internalized, and Gem was released into the cytoplasm. Upon exposure to the NIR laser, heat was generated, simultaneously promoting Gem release and increasing the therapeutic efect. In addition, compared with Gem-loaded MBPF (34.6%), MBPF+NIR (39.8%), and  $MoS<sub>2</sub>@BT$  (31.8%), breast cancer cells exposed to MBPF+Gem+NIR exhibited increased cytotoxicity (81.3%). In addition, Mo et al. chemically attached the ανβ3 integrin binding peptide RGD to  $MoS<sub>2</sub>$ -PEG [[78\]](#page-27-16). They further combined it with thiolated DOX (SH-DOX) via a disulfde linkage, leading to the creation of the  $\text{RGD}/\text{MoS}_2/\text{DOX}$  nanodrug system, termed  $MoS<sub>2</sub>-PEG-RGD-succinimidy$  3-[2-pyridyldithio] propionate (SPDP)-DOX (MPRS-DOX) (Fig. [5](#page-15-0)A). This innovative methodology endows  $MoS<sub>2</sub>$  with the ability to target ανβ3 integrin and respond to GSH, resulting in an increase in antitumor efficiency (Fig.  $5B$ , [C](#page-15-0)). In vitro experiments revealed that HeLa cells overexpressing ανβ3 integrin exhibited a viability rate of approximately 67% when treated with the MPRS-DOX nanodrug containing  $1.25 \mu$ g/mL DOX for 48 h. This percentage was notably less than the 98% viability observed in cells treated with MPRS alone. Moreover, when the dose of the MPRS-DOX nanodrug was increased to 10 μg/mL DOX, the viability of the HeLa cells decreased to 22.2%. Impressively, when ανβ3 integrin was combined with 808-nm NIR laser irradiation, the cytotoxic impact on ανβ3 integrin-overexpressing tumor cells intensifed, reducing HeLa cell viability to 45% (Fig. [5](#page-15-0)D).

Similarly, a  $MoS<sub>2</sub>$ -based biotin-functionalized nanoplatform was developed for targeted delivery. Due to the heightened demand for biotin for the accelerated growth of cancer cells, biotin receptors (BiRs) are overexpressed in cancer cells, making them important targets for cancer diagnosis and treatment [[158](#page-29-14), [159](#page-29-15)]. In light of this information, Liu et al. developed an innovative  $MoS<sub>2</sub>$ -based nanoplatform, DOX-Biotin-BSA-polyethyleneimine (PEI)-lipoic acid  $(LA)$ -MoS<sub>2</sub>-LA-PEG (DOX-BBPL-MoS<sub>2</sub>-LP), which was enhanced with LA-PEG and BSA to increase its dispersibility and colloidal stability and incorporated biotin to enable the specifc targeting of human cervical cancer cells [\[147](#page-29-0)]. The engineered nanoplatform generates considerable heat upon NIR light (808 nm) stimulation, not only ablating tumors but also facilitating the targeted release of DOX. This process enables potent synergy between PTT and chemotherapy. In addition, Chen et al. utilized a biotin-enhanced nanodrug delivery system to achieve the codelivery of curcumin (Cur) and erlotinib (Er) [\[160](#page-29-8)]. Cur is known to inhibit Er resistance by maintaining Ikappa-B expression and reducing phosphatidylinositol kinase levels in the epidermal growth factor receptor downstream signaling pathway  $[161, 162]$  $[161, 162]$  $[161, 162]$  $[161, 162]$ . This action facilitates the release of the apoptotic proteins caspase-3 and caspase-9, thereby promoting the apoptosis of tumor cells and enhancing the efectiveness of chemotherapy [[163\]](#page-29-18). Upon NIR irradiation,  $MoS<sub>2</sub>$ -PEG-Biotin efficiently converts the absorbed light into heat, enabling the photothermal ablation of cancer cells and consequently enhancing the antitumor efficacy. Research indicates that  $MoS<sub>2</sub>-PEG-Biotin-Cur/Er$  has a remarkable tumor growth inhibition rate of approximately 95.6% and signifcantly reduces the tumor volume under NIR irradiation, which is attributed mainly to the dynamic combination of enhanced synergistic chemotherapy and PTT  $[160]$  $[160]$ . Notably, 2D MoS<sub>2</sub> has been extensively investigated as a carrier for chemotherapeutic drug delivery. The innovative approaches discussed, including high drug loading capacities, enhancements in biocompatibility, and novel delivery strategies, underscore the potential of 2D  $MoS<sub>2</sub>$  to serve as an exemplary carrier. The integration of 2D  $MoS<sub>2</sub>$  with various functional modifcations demonstrates its versatility as a platform for enhancing the efficacy and safety of chemotherapy and PTT.



<span id="page-15-0"></span>Fig. 5 A Schematic diagram of MoS<sub>2</sub>/RGD/DOX nanodrug preparation for targeted delivery, intracellular GSH-triggered DOX release, and synergistic chemo/photothermal antitumor therapy. **B** Fluorescence microscope images showing the uptake of MPRS-DOX by Hela cells. **C** Fluorescence microscope images showing Hela cells apoptosis in diferent treatment groups. **D** Cell viability of Hela cells under diferent treatments. Reproduced with permission [[78\]](#page-27-16). Copyright 2022, Elsevier

#### **Gene therapy‑based combination therapy**

Small interfering RNA (siRNA)-induced gene therapy has been identifed as a promising and innovative approach for cancer treatment, as it can efectively inhibit tumor development by suppressing the expression of specifc genes [[164,](#page-29-19) [165\]](#page-29-20). Nonetheless, the naked form of siRNA is susceptible to degradation by both intracellular and extracellular nucleases, while its negatively charged nature hinders its cellular uptake  $[166]$  $[166]$  $[166]$ . The key to successful gene therapy relies heavily on the development of suitable siRNA carriers for efficient siRNA delivery into cells. In this context, the potential use of 2D  $MoS<sub>2</sub>$  in siRNA delivery has been explored, given its exceptional ability to deliver chemotherapeutic agents.

Polo-like kinase 1 (PLK1), a well-known oncogene, is a critical regulator of DNA replication [\[167](#page-29-22)]. Kou et al. synthesized  $MoS_{2}$ -PEG-PEI, wherein PEI provided a positive charge to bind to and deliver the negatively charged PLK1 siRNA [[168\]](#page-29-23). Following

 $MoS<sub>2</sub>$ -induced siRNA transfection, the efficacy of PLK1 silencing achieved with MoS<sub>2</sub>-PEG-PEI/siPLK1 was comparable to that achieved with Lipofectamine 2000 at an N/P ratio of 20. Notably, as the N/P ratio increased, the proportion of apoptotic cells treated with  $MoS<sub>2</sub>-PEG-PEI/siPLK1$  increased, indicating the efficacy of  $MoS<sub>2</sub>$  as a transfection carrier and the promise of gene therapy. In addition, the synergy of PTT and gene therapy is expected to yield more favorable outcomes. Kong et al. constructed a generation 5 (G5) poly(amidoamine) dendrimer-MoS<sub>2</sub> (G5-MoS<sub>2</sub>) platform for the combination B-cell lymphoma-2 (Bcl-2) gene silencing and PTT of tumors (Fig.  $6A$  $6A$ )  $[169]$  $[169]$ . The G5 dendrimers possess both a compact size and positive surface potential. These characteristics are pivotal, as they aid in the delivery of the Bcl-2 siRNA, enabling the downregulation of the Bcl-2 protein in cancer cells and inhibiting their growth. On the other hand, under photothermal conditions, the destruction of cancer cell



<span id="page-16-0"></span>Fig. 6 A Schematic diagram of the synthesis of G5-MoS<sub>2</sub> nanosheets and combinational gene silencing and PTT. B Schematic illustration of preparation of HAPM@siPDL1 and HAPM@siPDL1-mediated PD-L1 down-regulation and mPTT. Reproduced with permission [\[169\]](#page-29-24). Copyright 2017, American Chemical Society. Reproduced with permission [[173](#page-29-9)]. Copyright 2024, Elsevier

skeletons through hyperthermia-induced mechanisms can be achieved, resulting in synergistic cancer cell therapy  $[169, 170]$  $[169, 170]$  $[169, 170]$  $[169, 170]$ . The evidence of its effectiveness is clear: a marked decrease in cell viability was observed when cells were treated with  $G5-MoS<sub>2</sub>/siRNA$  polyplexes and laser irradiation, in contrast to cells treated with standalone siRNA with or without laser treatment. In support of these fndings, in vivo experiments revealed that while  $G5-MoS_2/siRNA$  treatment inhibited tumor growth to some extent, coupling  $G5-MoS<sub>2</sub>/siRNA$ treatment with laser irradiation not only considerably suppressed growth but also achieved complete tumor elimination. This finding underscores the strong synergy between PTT and gene silencing [\[169](#page-29-24)].

Instead of directly inhibiting or eliminating tumors, as with traditional oncogenes, innovative platforms are being developed to silence immune checkpointrelated genes to enhance antitumor immune responses. Programmed cell death protein 1/programmed deathligand 1 (PD-1/PD-L1) is a classic immune checkpoint that enables tumor cells to foster an immunosuppressive TME through the overexpression of PD-L1 [\[171,](#page-29-26) [172\]](#page-29-27). In light of these fndings, Ye et al. developed a nanoplatform

that carries PD-L1 siRNA (siPDL1), resulting in potent antitumor immune reactions [[173\]](#page-29-9). A 6-azidohexyl guanidine-rich α-helical polypeptide and HA were combined with 2D  $MoS<sub>2</sub>$  to form  $HAPM@siPDL1$ through complexation with siPDL1 to construct the nanoplatform (Fig. [6](#page-16-0)B). After targeted accumulation in tumors, HAPM@siPDL1 provoked efficient cytosolic delivery and PD-L1 silencing due to the potent membrane-penetrating capability of the polypeptides, resulting in attenuated immunosuppression. Importantly, the combination with mild PTT, characterized by tumor cell ablation in the low-temperature range  $(42-46 \degree C)$ , complements this efect by enhancing immunogenicity through tumor cell apoptosis and antigen release [\[174](#page-29-28)]. This synergistic action with siPD-L1 maximized the antitumor immune response, efectively increasing immunosuppression and reinstating immune surveillance. It has also displayed remarkable antitumor efficacy, with an approximately 95% tumor inhibition rate observed in mice treated with HAPM@siPDL1 under NIR irradiation within the 18-day observation period [[173\]](#page-29-9). From delivering siRNAs that target oncogenes such as PLK1 to engaging immune checkpoints such as PD-L1,

2D  $MoS<sub>2</sub>$  has versatile potential for both direct cancer treatment and immunomodulation. Building on this foundation, the integration of gene silencing with PTT represents a more efective approach to cancer therapy. Future research should not be confned to siRNAs alone; broader gene therapy strategies, including the use of messenger RNA and antisense oligonucleotides, should be explored to broaden the therapeutic potential [[175](#page-29-29)].

#### **Immunotherapy‑based combination therapy**

In recent years, immunotherapy has emerged as a powerful clinical strategy for cancer treatment. By employing the immune system to eradicate cancer cells, immunotherapy has demonstrated robust antitumor activity while mitigating metastasis and recurrence [\[176](#page-29-30)]. Various agents, such as cytosine–phosphate–guanine (CpG) and anti-PDL1 antibodies (aPDL1), are often used to augment the activation of the immune system against cancer cells [[177,](#page-29-31) [178](#page-29-32)]. Importantly, combination with delivery nanoplatforms could further increase cancer immunotherapy efficiency and reduce off-target adverse efects by increasing the accumulation within tumor tissues and the internalization of immunotherapeutic components [[179\]](#page-29-33).

Currently,  $2D$  MoS<sub>2</sub> is being investigated as a vehicle for immunoreagents. For example, Han et al. fabricated  $MoS<sub>2</sub>-PEG-CpG$  to deliver the immune adjuvant CpG for photothermal-enhanced immunotherapy  $[180]$  $[180]$ . The mammalian immune system recognizes CpG through Toll-like receptor 9. This interaction leads to the release of anticancer cytokines, activates helper T-cell 1-based cellular and humoral efector functions, and subsequently induces potent cytotoxic T lymphocyte (CTL) activity [\[177](#page-29-31), [181,](#page-29-35) [182](#page-29-36)]. However, these nucleic acids cannot easily cross the cell membrane due to their negative charges, and they are vulnerable to degradation by nucleases  $[183, 184]$  $[183, 184]$  $[183, 184]$ . Therefore, the delivery of CpG via  $MoS<sub>2</sub>$  represents a highly desirable solution to this challenge. Importantly, 2D  $MoS<sub>2</sub>$  loaded with CpG has been applied to target head and neck squamous cell carcinoma, a cancer type characterized by a highly immunosuppressive TME [\[185,](#page-29-39) [186](#page-29-40)]. Li et al. coated medium-sized and CpG-loaded 2D  $MoS<sub>2</sub>$  with low  $PEI_{0.8 \text{ k}}$  coverage to synthesize CpG@MM-PLs (Fig. [7](#page-17-0)A) [[145\]](#page-28-41). CpG@MM-PLs efectively promoted dendritic cell (DC) maturation and CTL function, thereby potentially reversing immunosuppression in this challenging context. In vivo, CpG@MM-PL treatment efectively



<span id="page-17-0"></span>**Fig. 7 A** Schematic diagram of CpG@MM-PL preparation and its antitumor therapy. **B** Schematic diagram and timeline of antitumor treatment with CpG@MM-PL. Tumor growth (**C**), survival curves (**D**) of mice with diferent treatments. **E** Schematic diagram and timeline of antitumor treatment with CpG@MM-PL combined with PD-1 antibody. Tumor growth (**F**), survival curves (**G**) of mice with diferent treatments. Reproduced with permission [[145](#page-28-41)]. Copyright 2023, Wiley

reduced tumor growth (Fig. [7](#page-17-0)B-D). Notably, when combined with immune checkpoint blockade, CpG@ MM-PLs exhibited an increased capacity to retard tumor growth, which was signifcantly greater than that of single CpG@MM-PL treatment (Fig. [7E](#page-17-0)-G).

In addition to mere combination therapies, the codelivery of immune enhancers and checkpoint inhibitors has been successfully implemented. Tang et al. pioneered a codelivery platform,  $MoS_{2}$ -aPDL1-V9302, designed for delivering both aPDL1 and V9302, aiming to amplify the anticancer immune response in triplenegative breast cancer cells  $[187]$ . The glutamine transporter inhibitor V9302 selectively targets alanine– serine–cysteine transporter 2, efectively obstructing glutamine absorption and thus reversing the nutritional deprivation exerted by cancer cells on immune cells [\[188](#page-29-41)]. Importantly, V9302 prompts the strategic repositioning of tumor-infltrating lymphocytes from the periphery of the tumor to its core  $[189]$ . This shift creates a conducive environment for immune checkpoint inhibitors to function, specifically by blocking additional PD-L1. The resulting synergy between V9302 and aPDL1 is evident in their ability to signifcantly increase lymphocyte proliferation, enhance lymphocyte functionality, and increase the efectiveness of anti-PDL1 therapy. In vivo experiments have shown that the administration of  $MoS<sub>2</sub>-aPDL1-V9302$  leads to elevated glutamine levels in the tumor interstitial fuid, which correlates with a substantial increase in activated CTLs, efectively inhibiting the growth of tumors [[187\]](#page-29-10). In conclusion, by facilitating the delivery of immune adjuvants such as CpG and aPDL1, 2D  $MoS<sub>2</sub>$  nanoplatforms address significant challenges such as immune suppression within the TME. The use of 2D  $MoS<sub>2</sub>$  not only improves the stability and efficacy of the immunoreagents but also synergistically enhances their therapeutic outcomes, particularly when combined with other treatments such as PTT and checkpoint inhibitors, demonstrating promising capabilities in advancing cancer immunotherapy.

#### **Photodynamic therapy‑based combination therapy**

2D  $MoS<sub>2</sub>$  has been leveraged not only for its role in catalyzing the generation of ROS for PDT but also as a carrier for agents to increase PDT efficiency. Xu et al. employed glucose-HPG-functionalized  $MoS_2$ (GPM) for the targeted delivery of the PDT dyes triphenylphosphonium (T) and glibenclamide (G) conjugated with Cy7.5 (Cy7.5-TG@GPM) (Fig. [8](#page-19-0)A) [[190\]](#page-30-0). This formulation achieves dual tumor- and subcellular-targeted PDT. Specifcally, Cy7.5-TG@GPM can target tumor cells expressing high levels of glucose transporter 1, ensuring signifcant tumor uptake. Upon exposure to NIR, Cy7.5-T and Cy7.5-G are released,

which target the mitochondria and ER, respectively. This release triggers the production of ROS that impair mitochondria and provoke ER stress, leading to the death of tumor cells. In detail, ROS-induced ER stress not only initiates the proapoptotic signaling cascade but also cooperatively promotes the release of cytochrome C from the mitochondria, causing tumor cell apoptosis. Moreover, such mitochondrial dysfunction disrupts ATP production, which in turn downregulates the expression of multidrug resistance (MDR)-related P-glycoprotein, thus contributing to overcoming MDR. The in vivo studies revealed remarkable success in inhibiting tumor growth and reversing MDR in multidrug-resistant HeLa cell tumors in nude mice, highlighting its potential for advanced therapeutic strategies (Fig. [8B](#page-19-0)).

In addition to the potent outcomes of 2D  $MoS<sub>2</sub>$ in PDT, further investigations have focused on the synergistic application of PDT and PTT to achieve better efficacy. In a pioneering work, Li et al. synthesized a novel nanocomposite,  $MoS<sub>2</sub>-TFPy$ , by intercalating aggregation-induced emission luminogens (AIEgens) mercapto-PEG-5-(4-(diphenylamino) phenyl) furan-2 pyridine (TFPy-SH) into the layers of  $MoS<sub>2</sub>$  nanosheets (Fig. [8](#page-19-0)C) [\[191](#page-30-1)]. This process not only expanded the interlayer spacing of the  $MoS<sub>2</sub>$  nanosheets but also induced a crystal phase transformation from 2H to 1 T. As a consequence of this structural transformation, compared with the pristine material,  $MoS_{2}-TFPy$ demonstrated an enhanced capability for photothermal conversion. The incorporation of positively charged AIEgens granted  $MoS<sub>2</sub>-TFPy$  the ability to accurately target mitochondria and increase the efficiency of ROS generation, establishing it as a highly efective photosensitizer in PDT applications. The therapeutic platform built on  $MoS<sub>2</sub>-TFPy$  integrated the enhanced PTT effect of  $MoS_2$ -TFPy with the PDT efficacy of TFPy-SH, culminating in a synergistic  $1+1>2$ ' effect on tumor therapy. Comparative in vitro and in vivo studies revealed that the antitumor efficacy of individual PTT  $(MoS<sub>2</sub>+NIR)$  or PDT  $(MoS<sub>2</sub>+white$  light) treatments and simple combinations of PTT and PDT  $(MoS<sub>2</sub>+TFPy-SH+white light+NIR)$  were significantly inferior to that of the combined  $MoS_2$ -TFPy+white light+NIR treatment, thereby underscoring the superior synergistic antitumor effect (Fig.  $8D$ , [E\)](#page-19-0). These studies highlight the significant potential of 2D  $MoS<sub>2</sub>$  in PDT and its combination with PTT for enhanced cancer treatment. By serving as a carrier for PDT agents and facilitating their targeted delivery to tumor cells, 2D  $MoS<sub>2</sub>$  not only improves the efficiency of ROS generation but also enhances subcellular targeting, contributing to the comprehensive eradication of cancer cells.



<span id="page-19-0"></span>**Fig. 8 A** Schematic illustration of mitochondria and ER-targeting synergistic PDT by Cy7.5-TG@GPM under NIR irradiation. **B** Relative tumor volume of mice under different treatments. C Schematic illustration for the fabrication of MoS<sub>2</sub>-TFPy and its synergistic anti-tumor PTT/PDT. Antitumor efcacy in vitro (**D**, relative cell viability) and in vivo (**E**, relative tumor volume) under diferent treatments. Reproduced with permission [\[190\]](#page-30-0). Copyright 2022, Elsevier. Reproduced with permission [[191](#page-30-1)]. Copyright 2024, Elsevier

Furthermore, the synergistic use of  $MoS<sub>2</sub>$  in combined PDT and PTT treatments represents a powerful strategy to overcome drug resistance and achieve superior therapeutic outcomes.

### **Synergistic strategies for precision theranostic applications**

The advancement of precision medicine has spurred significant interest in the development of an efficient theranostic platform that integrates imaging and therapeutic functionalities [[192,](#page-30-7) [193](#page-30-8)]. In this context, 2D  $MoS<sub>2</sub>$  offers a promising protocol for the simultaneous application of diagnostic bioimaging and combination therapy based on PTT or drug delivery (Table [1](#page-13-0)).

The exploration of dual-mode imaging-guided combination therapy has garnered signifcant attention in the field of cancer treatment. This emerging approach integrates diagnostics and therapeutics, aiming to increase the precision and efficacy of cancer therapy. In a recent study, Lu et al. engineered a versatile nanoplatform, designated ionic liquid (IL)- $MoS_{2}$ -PEGblock-poly(IL) (PEG-b-PIL)@DOX, for PA/thermal imaging-guided synergistic PTT/chemotherapy (Fig.  $9A$ ) [\[194](#page-30-2)]. This nanosheet was exfoliated with an antitumor IL and further tailored with PEG-b-PIL to improve its biodegradability, biocompatibility, and physiological stability. Its excellent photothermal properties endow it with exceptional capabilities in PAI, thermal imaging and PTT. In addition, its advantageous surface-area-to-volume ratio facilitates the efficient delivery of DOX for chemotherapy. The results revealed a pronounced PA signal at the tumor location at 12 and 24 h after intravenous administration, with the signal gradually increasing over time (Fig. [9B](#page-20-0)). Thermal imaging revealed a significant increase in temperature to 55.8 °C under 808 nm laser irradiation



<span id="page-20-0"></span>Fig. 9 A Schematic diagram of IL-MoS<sub>2</sub>-PEG-b-PIL preparation and its application in synergistic photothermal/chemotherapy and PAI for cancer. **B** Quantitative analysis of PA value. **C** Relative tumor volume of mice after different treatments. **D** Schematic diagram of 2D MoS<sub>2</sub>-based platform facilitating CT/PA/thermal imaging-guided photothermal-selective chemotherapy. CT (**E**) and PA (**F**) values of tumors in nude mice after MPT (I) and MPTF (II) intravenous injection. Thermal profles (**G**) of tumors in nude mice under NIR irradiation after I and II intravenous injection. **H** Relative tumor volume of mice under diferent treatments. Reproduced with permission [\[194](#page-30-2)]. Copyright 2023, Wiley. Reproduced with permission [\[196](#page-30-4)]. Copyright 2022, Elsevier

for 10 min, which was signifcantly greater than that observed with the saline injection (37.1 °C). Moreover, the group treated with IL-MoS<sub>2</sub>-PEG-b-PIL@DOX exhibited markedly superior tumor growth inhibition compared with the groups treated with IL-MoS<sub>2</sub>-PEG-b-PIL and free DOX, underscoring the enhanced efficacy of the combination of chemo-photothermal therapy (Fig. [9](#page-20-0)C). In a parallel study, Xia et al. exploited the unique properties of  $MoS<sub>2</sub>$  to facilitate a PA/ thermal imaging-guided combination of PTT and piezocatalytic therapy (PCT) [[195\]](#page-30-3). Benefting from the

photothermal effect,  $MoS<sub>2</sub>-PEG$  is employed not only as a potent contrast agent for PAI and thermal imaging but also as an efficacious PTA for PTT. Additionally,  $MoS<sub>2</sub>-PEG$  exhibits a remarkable piezotronic effect, converting mechanical vibration energy into electrical energy under the stimulation of ultrasound-mediated micropressure. This conversion triggers ROS generation for cancer PCT to further kill cancer cells. After an intravenous injection of  $MoS<sub>2</sub>-PEG$  nanosheets into 4T1 tumor-bearing mice, signifcant PA signals and an increase in temperature were observed under 1064 nm

laser irradiation within just 10 min. Notably, the tumors were almost completely inhibited in the experiments where the tumors were treated with  $MoS<sub>2</sub>-PEG$ nanosheets combined with NIR and ultrasound.

Based on the promising results achieved through dual imaging, MMI-guided combination therapy has been further explored to advance the acquisition of precision medicine. In a recent study, Li et al. covalently blended PEGylated α-tocopheryl succinate (α-TOS) and FA on 2D  $MoS<sub>2</sub>$  to construct a comprehensive treatment platform,  $MoS<sub>2</sub>-PEG-TOS-FA (MPTF)$  (Fig. [9D](#page-20-0)) [[196](#page-30-4)]. Based on excellent CT/PA/thermal imaging capacities conferred by high atomic numbers and photothermal effects, MPTF could be employed to locate ovarian tumors preoperatively using MMI (Fig.  $9E-G$ ). The inclusion of FA ensures targeted delivery, meaning that MPTF will predominantly accumulate within tumors, paving the way for more effective treatments  $[197]$  $[197]$ . α-TOS can induce tumor cell apoptosis with no toxicity to healthy tissues [\[198](#page-30-10)]. Building on these advantages, highly efficient PTT could be activated to completely ablate the entire solid tumor under safe NIR irradiation (Fig. [9](#page-20-0)H). Finally, locally infltrating and metastatic cancer cells are killed by α-TOS to prevent recurrence. Notably, the excellent efficacy and safety of synergistic therapy resulted in a 100% survival rate of tumor-bearing mice over 91 days [[196\]](#page-30-4). Furthermore, Hu et al. developed 1-methyl-tryptophan (1-MT) cisplatin (Pt)-PPDA@MoS<sub>2</sub> complexes, realizing CT/PA/ thermal imaging-guided chemo-photothermal immunotherapy [\[199\]](#page-30-5). NIR laser irradiation (PTT) combined with Pt (chemotherapy) can destroy tumor cells and efectively induce immunogenic cell death and DC maturation, signifcantly bolstering T-cell-mediated antitumor immune responses. This effect is further amplified by 1-MT, which blocks the immune checkpoint associated with indoleamine 2,3-dioxygenase-mediated tryptophan metabolism. This blockade interferes with tryptophan metabolism, hindering the development of tumor-regulatory T cells and fostering the activation of T-cell-driven immunotherapy [[199–](#page-30-5)[201](#page-30-11)]. Impressively, the use of 1-MT-Pt-PPDA@MoS<sub>2</sub> with laser therapy led to complete tumor eradication in just 8 days, underscoring the unparalleled potency of this trimodal therapeutic approach. In addition, the excellent photothermal properties and X-ray absorption ability of the nanoplatform enabled comprehensive CT/PA/thermal imaging of the tumors. As expected, the CT/PAI signals of the tumors tended to increase with increasing Mo concentrations. The thermal images were also obviously obtained under 808 nm laser irradiation for 300 s, with the temperature in the tumor region increasing to 61 °C, which was much higher than that of the injection of PBS (less than 40 °C) [[199\]](#page-30-5).

Based on their inherent properties, loading and delivering various imaging components and therapeutic agents will lead to more possibilities in cancer therapeutics and diagnostics. For example, Liu et al. designed functionalized nanosheets termed  $MoS<sub>2</sub>-HA-diethylenetriaminepentaacetic acid (DTPA)$ gadolinium  $(Gd)/$ gefitinib  $(Gef)$  (Fig. [10](#page-22-0)A) [[202](#page-30-12)]. These nanosheets were crafted by surface decoration with HA, conjugation with Gd ions via DTPA and the physical incorporation of Gef. The functionalized nanoplatform is capable of achieving the targeted codelivery of Gd-based contrast agents and the anticancer drug Gef, enabling both MRI and synergistic chemo-photothermal therapy. By incorporating HA as a targeting ligand, the nanosheets can efficiently direct the loaded Gd toward cancer cells that overexpress the HA receptor  $[203]$  $[203]$ . This targeted approach results in enhanced relaxivity, which is 3.3 times greater than that of commercial contrast agents such as DTPA-Gd, and high-resolution images of the tumor. In terms of therapy, the nanoplatform efectively converts the absorbed NIR light into heat, which not only induces the photothermal ablation of cancer cells but also spurs the release of Gef, laying the foundation for efective synergistic therapy. When measured against either chemotherapy or PTT alone, this combined approach is signifcantly more efective in inhibiting tumor growth in mice injected with lung cancer cells [\[202\]](#page-30-12). Similarly, Cai et al. revealed an integrated nanocomposite, gambogic acid  $(GA)/M_0S_2/BSA$ -cadmium trioxide  $(Gd_2O_3)$ -HA (GMH), where 2D  $MoS<sub>2</sub>$  is functionalized with BSA- $Gd<sub>2</sub>O<sub>3</sub>$  and further augmented with GA, serving as both a chemotherapeutic and a heat shock protein 90 (HSP90) inhibitor [\[204](#page-30-6), [205\]](#page-30-14). This composite is designed for MRI-guided combined low-temperature PTT (43–45 °C) and chemotherapy (Fig. [10](#page-22-0)B). BSA- $Gd<sub>2</sub>O<sub>3</sub>$ , an MRI contrast agent, provides benefits such as excellent biocompatibility, a straightforward synthesis process, and a high relaxation rate [\[206\]](#page-30-15). Impressively, the T1 relaxation rate (r1) of  $MoS<sub>2</sub>/BSA-Gd<sub>2</sub>O<sub>3</sub>$  was 3.5 times higher than that of conventional Gd-DTPA. GA is particularly critical, as it specifcally inhibits the overexpression of HSP90 when subjected to elevated temperatures, thereby reducing tumor thermoresistance and facilitating distinct types of apoptosis at relatively low temperatures [[205\]](#page-30-14). Consequently, the integration of GA with PTT enables the killing of tumors at low temperatures without harming adjacent normal organs and efectively provides deep-seated tumors with sufficient heat to minimize recurrence. Consistent with our theoretical predictions, PTT with  $MoS<sub>2</sub>/$ BSA-Gd<sub>2</sub>O<sub>3</sub>-HA resulted in only partial inhibition of tumor growth at a relatively low temperature (43 °C), whereas mice treated with GMH exhibited signifcant



<span id="page-22-0"></span>Fig. 10 A Schematic diagram of HA-grafted MoS<sub>2</sub> as a carrier for co-delivering Gef and Gd-based contrast agents for MRI guided chemo-photothermal therapy. **B** Schematic illustration of GA/MoS<sub>2</sub>/BSA-Gd<sub>2</sub>O<sub>3</sub>-HA for MRI-guided combined low-temperature PTT and chemotherapy. Reproduced with permission [[202](#page-30-12)]. Copyright 2019, Elsevier. Reproduced with permission [\[204](#page-30-6)]. Copyright 2021, Elsevier

tumor inhibition  $[204, 205]$  $[204, 205]$  $[204, 205]$ . The aforementioned studies illustrate the advanced utilization of  $MoS<sub>2</sub>$  nanoplatforms in dual-mode and MMI-guided combination therapies. These approaches represent significant progress in overcoming MDR, enhancing therapeutic efficacy, and facilitating comprehensive tumor imaging. Innovations in  $MoS<sub>2</sub>$ -based theranostics are paving the way for more precise and efective treatments, advancing personalized medicine in oncology.

#### **Challenges and future perspectives**

The exploration of 2D  $MoS<sub>2</sub>$  as a theranostic platform for cancer treatment exemplifes how the primary objectives of modern oncology align with those of nanomedicine. This alignment emphasizes the development of innovative methods by integrating various treatment modalities and diagnostic techniques. This review highlights the potential of 2D  $MoS<sub>2</sub>$  to revolutionize cancer treatment through the development of single-mode theranostic and synergistic platforms that combine

diagnostic and targeted therapeutic functionalities, aligning with the precision medicine paradigm. However, the path from the research bench to the patient bedside is laden with challenges that must be meticulously navigated.

One of the most pressing challenges is the ability to control synthesis process of  $2D$  MoS<sub>2</sub> nanocomposites to ensure the uniformity, reproducibility, and scalability necessary for clinical applications  $[57, 207]$  $[57, 207]$  $[57, 207]$ . The complexity of the materials requires precise control over the synthesis process, which is crucial for the clinical translation of  $MoS<sub>2</sub>$ -based theranostics. Moreover, given the unknown potential risks, evaluating the toxicity of nanomaterials is crucial. While the low toxicity of 2D  $MoS<sub>2</sub>$  has been preliminarily reported in cellular and murine models, the path to clinical translation requires extensive, systematic investigations [\[62](#page-27-0), [65,](#page-27-3) [68\]](#page-27-6). Further assessments at both the cellular and subcellular levels are essential to fully explore the implications of these therapies, including potential side efects on organelle structure and cellular signaling pathways [\[208\]](#page-30-17). In murine models, toxicity is often assessed by evaluating the accumulation in organs such as the liver and lungs, as well as weight loss. However, a comprehensive assessment of biotoxicity should encompass additional systems, including the cardiovascular, reproductive, and nervous systems, to provide a holistic understanding of the potential risks [[209](#page-30-18)]. Additionally, more extensive use of larger animal models and human-mimicking systems, such as organoids and microfluidic systems, is necessary to accurately represent patient biology and tumor heterogeneity [\[210](#page-30-19), [211\]](#page-30-20). In conclusion, a thorough understanding of the long-term toxicity and systemic efects of these nanomaterials is essential for establishing their clinical viability.

The pursuit of increased targeting capabilities is paramount to enhance therapeutic efficacy and minimize side effects. Research on drug delivery strategies utilizing 2D  $MoS<sub>2</sub>$  platforms is still in its infancy. After 2D  $MoS<sub>2</sub>$  is successfully delivered to tumors by leveraging overexpressed receptors on tumor cell membranes and the EPR efect, utilizing both endogenous and exogenous stimuli to achieve smart regulation within tumors is crucial. Future research should focus on innovative targeting mechanisms that are responsive to biological cues such as ROS, hypoxia, and external stimuli such as ultrasound or electric felds [\[212](#page-30-21)]. Furthermore, the range of drugs delivered using  $2D$  MoS<sub>2</sub> remains limited. Considering the rapid advancements in the application of 2D materials in drug delivery, future research on 2D  $MoS<sub>2</sub>$  should delve deeper into exploring its potential uses in new domains. This information is pertinent in the delivery of biologics, such as oncolytic viruses, where

novel delivery mechanisms may signifcantly increase therapeutic efficacy  $[213]$  $[213]$ . Additionally, the complexity and dynamics of cancer require a deeper understanding of the antitumor effects of 2D  $MoS<sub>2</sub>$  nanocomposites and the exploration of potential therapeutic mechanisms.

With increasing focus on the idea of precision medicine, theranostic platforms based on 2D  $MoS<sub>2</sub>$ still need to be optimized. In addition to the current research into the therapeutic applications of 2D  $MoS<sub>2</sub>$ , other treatment strategies that leverage its outstanding physicochemical properties, such as radiotherapy and sonodynamic therapy, are attracting signifcant interest [\[214\]](#page-30-23). Employing a more diversifed therapeutic strategy will unlock the prospects and potential for precision medicine. For instance, Cai et al. constructed a  $MoS_2/BSA-Gd_2O_3$  complex, where the inclusion of  $Gd_2O_3$  enables the platform to be well suited for MRI applications [[204](#page-30-6)]. Similarly, other metal elements, such as copper, can also act as functional reagents. This diverse combination of nanoparticles provides substantial scope for the development of innovative  $MoS<sub>2</sub>$  composites. Moreover, the heterojunction strategy should also be widely promoted within the  $2D$  MoS<sub>2</sub> platform to meet the demands of precision medicine. For example, Yu et al. used a myeloid-derived suppressor cell membrane to encapsulate Fe<sub>3</sub>O<sub>4</sub>, thereby enhancing targeting [\[215\]](#page-30-24). A neutrophil membrane was used to coat black phosphorus to improve biocompatibility  $[216]$  $[216]$ . These strategies exemplify how similar approaches could be benefcially extended to  $2D$   $MoS<sub>2</sub>$  composites. Exploring therapeutic platforms more broadly and in greater depth should be the direction of our future research to fully harness the potential of 2D  $MoS<sub>2</sub>$  in medical applications.

Despite the continuously growing list of nanodrugs in patent and clinical trials, the clinical translation of 2D nanodrugs remains largely unexplored [[217\]](#page-30-26). To date, few patents involving 2D MoS₂ have been reported (Table [2\)](#page-24-0), and there have been no related clinical trials. This underscores the need for substantial efforts to advance this feld. Achieving scalability, reducing toxicity, and optimizing therapeutic efficacy are crucial steps in the pathway to clinical translation, as mentioned above. Historically, the failure of many nanodrugs during phase II trials has been attributed predominantly to poor efficacy, suggesting that after ensuring biosafety, enhancing therapeutic efficacy is highly important  $[217]$  $[217]$ . Beyond the inherent challenges, the clinical translation process for  $2D$  MoS<sub>2</sub> is marked by uncertainty, a common issue for 2D nanodrugs, especially for borderline products combining multiple technologies. Establishing clear regulatory guidelines for the use of 2D nanodrugs in cancer treatment is essential to ensure their successful translation through clinical trials [[218,](#page-30-27) [219](#page-30-28)].



#### <span id="page-24-0"></span>**Table 2** Summary of recent patents on 2D MoS<sub>2</sub>-based cancer theranostics

Molybdenum disulfide (MoS<sub>2</sub>), polyethylene-glycol (PEG), doxorubicin (DOX), manganese dioxide (MnO<sub>2</sub>), erythrocyte (RBC), tetra (4-carboxyphenyl) porphin (TCPP), triphenylphosphine (TPP), immune adjuvant imiquimod (R837), folic acid (PVA), α-tocopheryl succinate (α-TOS), folic acid (FA), 1-methyl-tryptophan (1-MT), cisplatin (Pt), polydopamine (PDA), bovine serum albumin (BSA), aptamer (Apt), photothermal therapy (PTT), piezo-catalytic therapy (PCT), photodynamic therapy (PDT), computerized tomography (CT), photoacoustic imaging (PAI)

UV Ultraviolet

Apt Aptamer Ce6 Chlorine e6 Ag Silver

H<sub>2</sub>O<sub>2</sub> Hydrogen peroxide<br>
OPD Ortho-phenylenedia

DNA-AgNCs DNA-silver nanoclusters PA Photoacoustic ICG Indocyanine green

OH· Hydroxyl radicals ATP **Adenosine triphosphate** 

MSNs Mesoporous silica nanoparticles

Ortho-phenylenediamine

The advancement of 2D  $MoS_2$ -based theranostic applications from the laboratory to clinical settings requires a concerted effort involving multidisciplinary research teams and medical practitioners. The concept of 2D  $MoS<sub>2</sub>$  as an "all-in-one" theranostic platform is a beacon of innovation in the feld of nanomedicine, representing a signifcant stride toward the goal of precision medicine in cancer treatment, and it is poised to play a pivotal role in the future of oncology.

#### **Abbreviations** 2D Two-dimensional CT Computerized tomography EPR Enhanced permeation and retention FA Folic acid MRI Magnetic resonance imaging  $M$ oS<sub>2</sub>  $M$ olybdenum disulfide<br>NIR Near-infrared PEG Polyethylene glycol PL Photoluminescence PTT Photothermal therapy TMDs Transition metal dichalcogenides CVD Chemical vapor deposition LPE Liquid phase exfoliation Li Lithium CS Chitosan BSA Bovine serum albumin DNA Deoxyribonucleic acid RNA Ribonucleic acid ROS Reactive oxygen species LA Lipoicacid PBS Phosphate buffered saline RGD **Arginine-glycine-aspartate** HA Hyaluronic acid CD44 Cluster determinant 44 S–S Disulfde bond GSH Glutathione TME Tumor microenvironment PAI Photoacoustic imaging FI FI Fluorescence imaging PTAs Photothermal agents PAA Poly (acrylic acid) Ab Antibody EMT Epithelial-mesenchymal transition CDs Carbon dots T-BTO TETRAGONAL TETRAGONAL DATION TETRAGONAL CA Cinnamaldehyde Fe Iron MSFP MoS<sub>2</sub>@SA-Fe-PEG<br>NH<sub>2</sub>-PEG Amine-polyethyle NH<sub>2</sub>-PEG **Amine-polyethylene glycol**<br>MMI Multimodal imaging Multimodal imaging PDA Polydopamine<br>MPPF MoS<sub>2</sub>@PDA-PE  $MOS_2@PDA-PEG/peptide$ <br>
Near-infrared fluorescence Near-infrared fluorescence PDT Photodynamic therapy DOX Doxorubicin<br>
HPG Hyperbrancl Hyperbranched polyglycidyl CQ Chloroquine RBC Erythrocyte<br>BT Barium titar Barium titanate MBPF  $MoS<sub>2</sub>@ BT-PDA-FA$ <br>Gem Gem Gemcitabine Thiolated DOX SPDP Succinimidyl 3-[2-pyridyldithio] propionate  $MoS<sub>2</sub>-PEG-RGD-SPDP-DOXBiRs$ Biotin receptors PEI POLYethylenimine<br>
ROPI Polyethylenimine<br>
Ropin-RSA-PFI-I A Biotin-BSA-PFI-LA Cur Curcumin Er Erlotinib siRNA Small interfering RNA PLK1 Polo-like kinase 1 G5 Generation 5



#### **Acknowledgements**

Not applicable.

#### **Author contributions**

Xinbo Yu: Writing–review & editing, Writing–original draft, Visualization, Validation, Conceptualization. Chen Xu: Writing–review & editing, Writing– original draft, Visualization, Validation, Conceptualization. Jingxu Sun: Writing– review & editing, Writing–original draft, Visualization, Validation, Funding acquisition, Conceptualization. Hainan Xu: Writing–review & editing, Writing– original draft, Visualization, Validation, Conceptualization. Hanwei Huang: Writing–review & editing, Validation. Ziyang Gan: Writing–review & editing, Validation. Antony George: Validation. Sihui Ouyang: Writing–review & editing, Writing–original draft, Visualization, Validation, Conceptualization. Funan Liu: Writing–review & editing, Writing–original draft, Visualization, Validation, Funding acquisition, Conceptualization.

#### **Funding**

This work was supported by National Key R&D Program of China [grant numbers 2019YFC1316104, 2022YFC2403401]; National Natural Science Foundation of China [Grant Numbers 81871960, 82073368, 82373110, 82102686]; and Liaoning Revitalization Talents Program [Grant Numbers XLYC2007071, XLYC1808017].

#### **Data availability**

No datasets were generated or analysed during the current study.

#### **Declarations**

#### **Ethics approval and consent to participate**

Not applicable.

#### **Consent for publication**

All authors gave their consent for publication.

#### **Competing interests**

The authors declare no competing interests.

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# Received: 21 June 2024 Accepted: 18 August 2024

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