

RESEARCH

Open Access

# Synergistic photothermal ablative effects of functionalizing carbon nanotubes with a POSS-PCU nanocomposite polymer

Aaron Tan<sup>1</sup>, Seyed Yazdan Madani<sup>1</sup>, Jayakumar Rajadas<sup>2</sup>, Giorgia Pastorin<sup>3</sup> and Alexander M Seifalian<sup>1,4\*</sup>

## Abstract

**Background:** The application of nanotechnology in biology and medicine represents a significant paradigm shift in the approach to the treatment of cancer. Evidence suggests that when exposed to near-infrared radiation (NIR), carbon nanotubes (CNTs) dissipate a substantial amount of heat energy. We have developed a novel nanocomposite polymer, polyhedral oligomeric silsesquioxane poly (carbonate-urea) urethane (POSS-PCU). POSS-PCU displays excellent biocompatibility and has been used in making artificial organs as well as protective coatings for medical devices.

**Results:** Functionalizing (or “coating”) CNTs with POSS-PCU confers biocompatibility and increase the amount of heat energy generated, by enhancing dispersion. Here we demonstrate that POSS-PCU-functionalized multi-walled CNTs (MWNTs) act synergistically together when exposed to NIR to thermally ablate cancer cells.

**Conclusion:** Given that POSS-PCU has already been used in human in first-in-man studies as trachea, lacrimal duct, bypass graft and other organs, our long-term goal is to take POSS-PCU coated CNTs to clinical studies to address the treatment of cancer by optimizing its therapeutic index and increasing its specificity via antibody conjugation.

**Keywords:** Carbon nanotubes, POSS-PCU, Thermal ablation, Nanotechnology, Nanocomposite polymer, Colorectal cancer

## Background

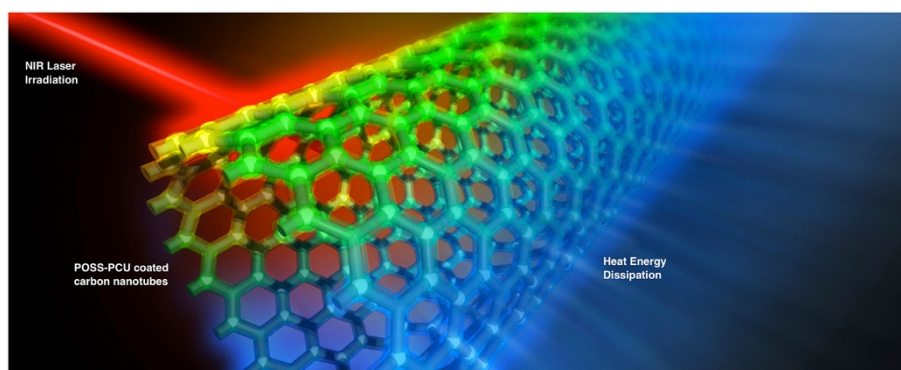
Carbon nanotubes (CNTs) were initially described as “helical microtubules of graphitic carbon” by Sumio Iijima (of NEC Corporation), in a letter to Nature in 1991 [1]. An unprecedented interest in its application has arisen since then, with wide-ranging potentials from being high-power energy storage devices [2], to therapeutic and diagnostic agents in cancer biology [3,4]. CNTs have a high optical absorbance in the near infrared (NIR) region [5] (700 nm to 1100 nm) and subsequently dissipate a high amount of heat energy. This phenomenon can thus be exploited in a biomedical setting to destroy cancer cells via photothermal ablation [6,7] (Figure 1).

Generally, CNTs can be classified into two broad categories: multi-walled carbon nanotubes (MWNTs) and single-walled carbon nanotubes (SWNTs). SWNTs can be conceptualized as a rolled-up sheet of planar  $sp^2$  hybridized carbon atoms, while MWNTs are concentric layers of SWNTs [8]. CNTs have exceptional tensile strength [9] (in the gigapascal (GPa) region), and a very high Young’s modulus [10] (in the tetrapascal (TPa) region), making them one of the strongest materials known. It has been previously reported that MWNTs display a higher thermal conductivity, compared to SWNTs, as their larger diameters allow a greater number of phonon modes, resulting in longer phonon mean free paths [11]. Due to the lack of endogenous chromophores that absorb within the NIR spectrum, biological membranes are highly transparent to NIR within wavelengths of 700 nm to 1100 nm [12]. Alluding to their electronic band structures, CNTs can attain a very high temperature (of up to 70°C) within minutes of exposure to NIR lasers [13]. Exposure of CNTs to NIR laser would

\* Correspondence: aseifalian@ucl.ac.uk

<sup>1</sup>Centre for Nanotechnology & Regenerative Medicine, UCL Division of Surgery & Interventional Science, University College London, London, NW3 2QG, UK

<sup>4</sup>Royal Free London NHS Foundation Trust Hospital, London, UK  
Full list of author information is available at the end of the article



**Figure 1** A computer graphics rendering of photothermal ablation when POSS-PCU-functionalized CNTs are exposed to near infrared (NIR) laser. CNTs are functionalized with a novel nanocomposite polymer, POSS-PCU, depicted as a translucent “coating” on the CNT wall. Intense heat energy is dissipated, depicted by blue emission, which can be exploited to thermally ablate cancer cells.

release a significant amount of vibrational energy, which consequently causes cell death via coagulative necrosis. This event is known as photothermal cancer ablation, with rupturing cell membranes and denaturation of cellular proteins.

We have previously developed a novel nanocomposite polymer called polyhedral oligomeric silsesquioxane poly (carbonate-urea) urethane (POSS-PCU, trade name UCL Nano™). POSS-PCU is biocompatible [14], resistant to degradation [15], and has potential applications for being scaffolds for artificial organs [16], and coatings for medical devices [17]. Indeed, first-in-man clinical studies have already been conducted using POSS-PCU constructed three-dimensional (3D) biomimetic scaffolds of trachea [18], lacrimal duct and bypass graft, with favorable results.

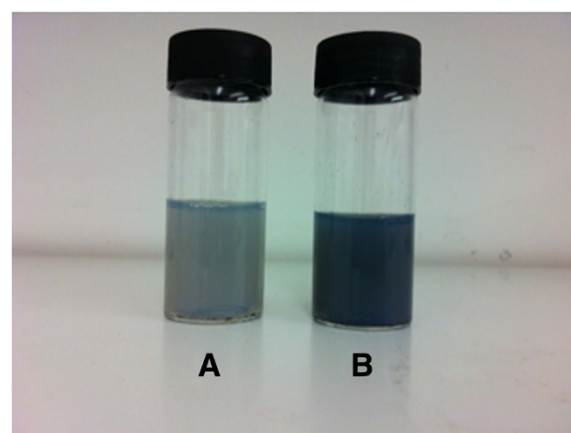
A major concern for the use of CNTs in biological systems is its insolubility and inherent toxicity. With proper functionalization, for example with polyethylene glycol (PEG), CNTs appear to be well-tolerated in animal studies [19]. Hence, we postulate the amphiphilic nature of POSS-PCU would increase dispersion of CNTs in biological systems [20], as well as conferring biocompatibility. Furthermore, we also set out to investigate if the increased dispersion of CNTs would affect temperature changes during NIR laser exposure, as well as the effects of photothermal ablation on a colorectal cancer cell line.

## Results & discussion

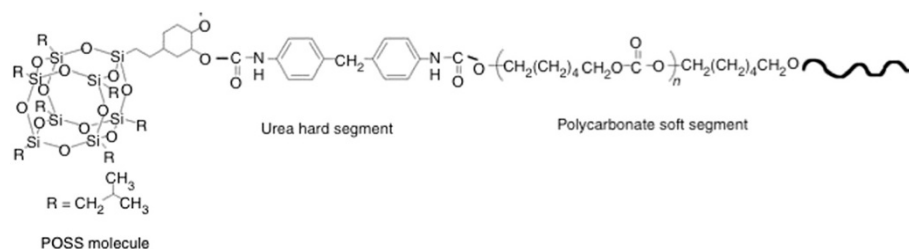
### Conferring solubility and biocompatibility via POSS-PCU functionalization

Raw CNTs appear as a black powder-like substance, which is insoluble in water. Upon functionalization with POSS-PCU via bath sonication, the final product appears as a homogenous mixture (Figure 2) suitable for biological applications. The 2 main functionalization schemes are covalent and non-covalent. Covalent functionalization

involves oxidation or cycloaddition reactions, which introduces functional groups onto the side-walls of CNTs [13]. However, the intrinsic chemical harshness of covalent functionalization disrupts the structure of CNTs, interrupting its inherent attributes like Raman scattering and photoluminescence [21]. In contrast, non-covalent functionalization preserves the structural integrity of CNTs, as it simply provides an external “coating”. Non-covalent functionalization occurs via hydrophobic interactions or  $\pi$ - $\pi$  stacking of the aromatic rings [22]. Functionalization via PEG phospholipids (PL-PEG) can render CNTs biocompatible while simultaneously increasing its circulation time in biological systems [23]. Arising from this, we postulate that POSS-PCU would similarly interface with CNTs via  $\pi$ - $\pi$  stacking of its aromatic rings of the urea hard segment (Figure 3) onto the wall of CNTs. TEM images and UV/Vis/NIR analysis (Figures 4 and 5)



**Figure 2** Visual appearance of CNTs after functionalization with POSS-PCU. **A** contains CNTs functionalized with 50% POSS-PCU, while **B** contains 100% POSS-PCU. Non-covalent functionalization with POSS-PCU via bath sonication confers solubility to CNTs.



**Figure 3 Skeletal formula of POSS-PCU.** This novel nanocomposite polymer displays superior biocompatibility and has been used in constructing scaffolds for artificial organs as well as protective coating for medical devices.

confirms the non-covalent functionalization of POSS-PCU onto the surface of CNTs.

Raw CNTs are insoluble in water and biological systems. Upon non-covalent functionalization with POSS-PCU, solubility was conferred, rendering it ready for use in biological systems. POSS-PCU is highly compatible, and its use in humans as a bioartificial trachea, bypass graft and lacrimal duct underscores its potential usability as a coating agent for nanoparticles as well. Unfunctionalized CNTs are not appropriate for use in biological systems, and we postulate that biocompatibility can be conferred via functionalization with POSS-PCU.

#### Synergistic thermal effects of POSS-PCU and CNTs

As POSS-PCU has thermoplastic behavior, does not absorb in the near infrared spectrum. Thus, when exposed to NIR laser, the temperature of POSS-PCU remains constant at room temperature. Our results reveal that the presence of

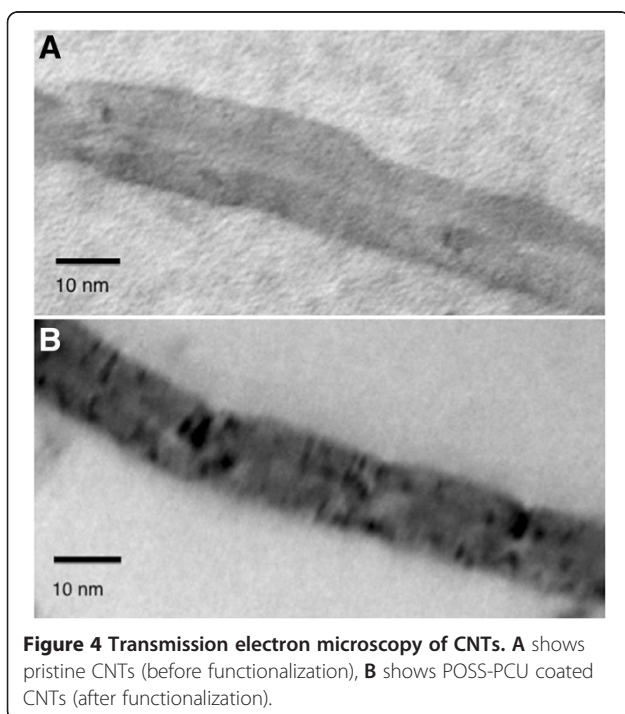
POSS-PCU does not simply maintain the temperature profile curve (as one might expect due to its absence of heating effect), but instead increases the temperature profile curve when functionalized with CNTs.

Upon functionalization with POSS-PCU, CNTs were able to attain a greater rate of temperature increase as well as a higher final temperature of 88°C within ten minutes of NIR laser exposure compared to non-functionalized CNTs ( $p = 0.0006$ ). At both wattages, synergistic thermal effects were observed, as POSS-PCU-CNT was able to attain a higher temperature than POSS-PCU and CNTs alone (Figure 6). At higher wattage, functionalization with 100% POSS-PCU shows a greater increase in temperature compared to 50% POSS-PCU. This is possibly due to the significant greater magnitude of vibrational energy, which manifests itself more synergistically at 100% POSS-PCU functionalization due to its dispersion effects. 100% POSS-PCU functionalization ensures that a greater surface area of CNTs are exposed to NIR, hence increasing its heating effects.

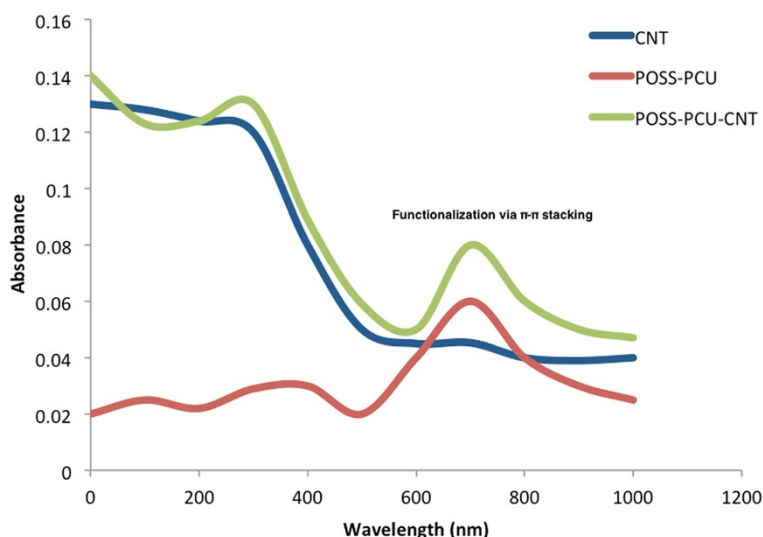
We have therefore successfully demonstrated that POSS-PCU can not only confer solubility to CNTs, but also enhance its heating effects (Figure 7). This is attributed to POSS-PCU acting like a surfactant when functionalized to CNTs, increasing its dispersion in biological systems [20]. Consequently, the surface area to volume ratio of functionalized CNTs would be increased, allowing more particles per unit area to absorb the NIR laser, thereby producing a higher rate of temperature increase as well as a higher final temperature.

#### Enhanced cell kill using POSS-PCU-CNT complex

With the observation of increased thermal response to NIR exposure, we proceeded to investigate if POSS-PCU-CNT was able to thermally ablate cancer cells. A HT-29 human colorectal cancer cell line was used as a model for photothermal ablation. There was a slight decrease in cell number after incubation with CNTs (without laser), possibly due to the inherent toxicity of CNTs. In contrast, treatment with POSS-PCU-CNT (without laser) did not result in any significant change in cell number, indicating that functionalization with POSS-PCU had conferred a high degree of



**Figure 4 Transmission electron microscopy of CNTs.** **A** shows pristine CNTs (before functionalization), **B** shows POSS-PCU coated CNTs (after functionalization).

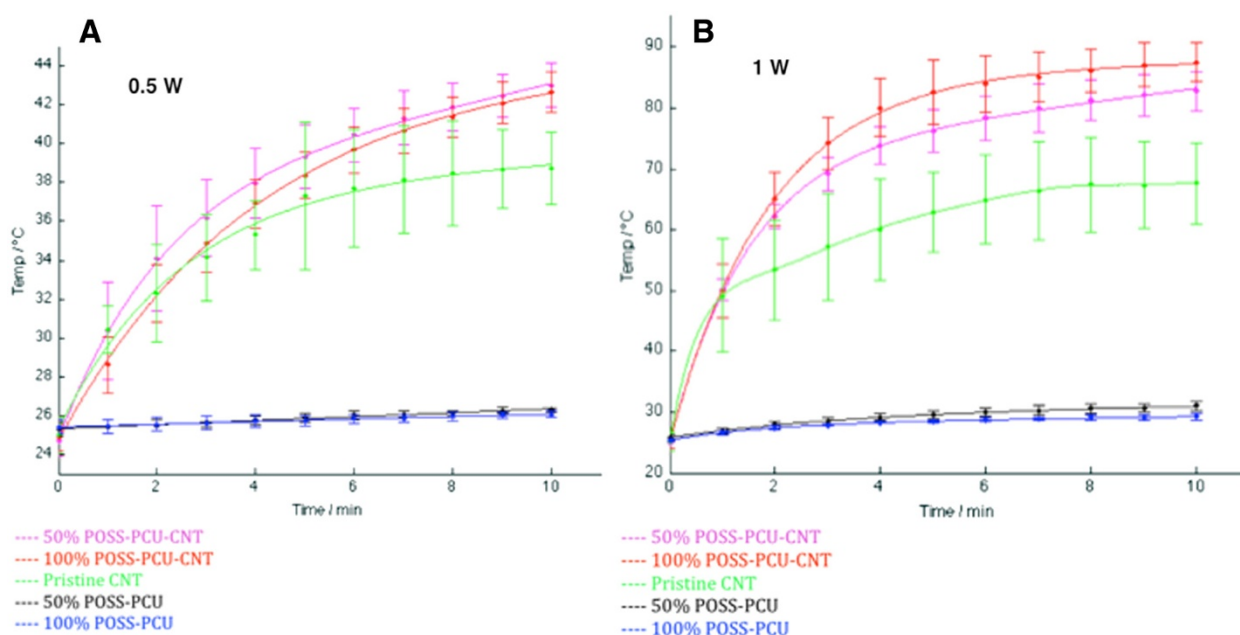


**Figure 5 Absorption spectra of POSS-PCU-CNT.** At a wavelength of 700 nm, POSS-PCU shows greatest absorbance (red). Non-covalent functionalization of POSS-PCU onto CNTs results in a higher absorbance at 700 nm (green), compared to unfunctionalized CNTs (blue).

biocompatibility to CNTs. Concurrent with established literature, a decrease in cell number (of around 50%) was observed when (unfunctionalized) CNTs were irradiated with NIR laser. More importantly, when POSS-PCU-CNT was irradiated with NIR laser, there was a dramatic decrease in cell numbers, thermally ablating 95% of cancer cells ( $p=0.0000842$ ) (Figures 8 and 9). Hence, we have

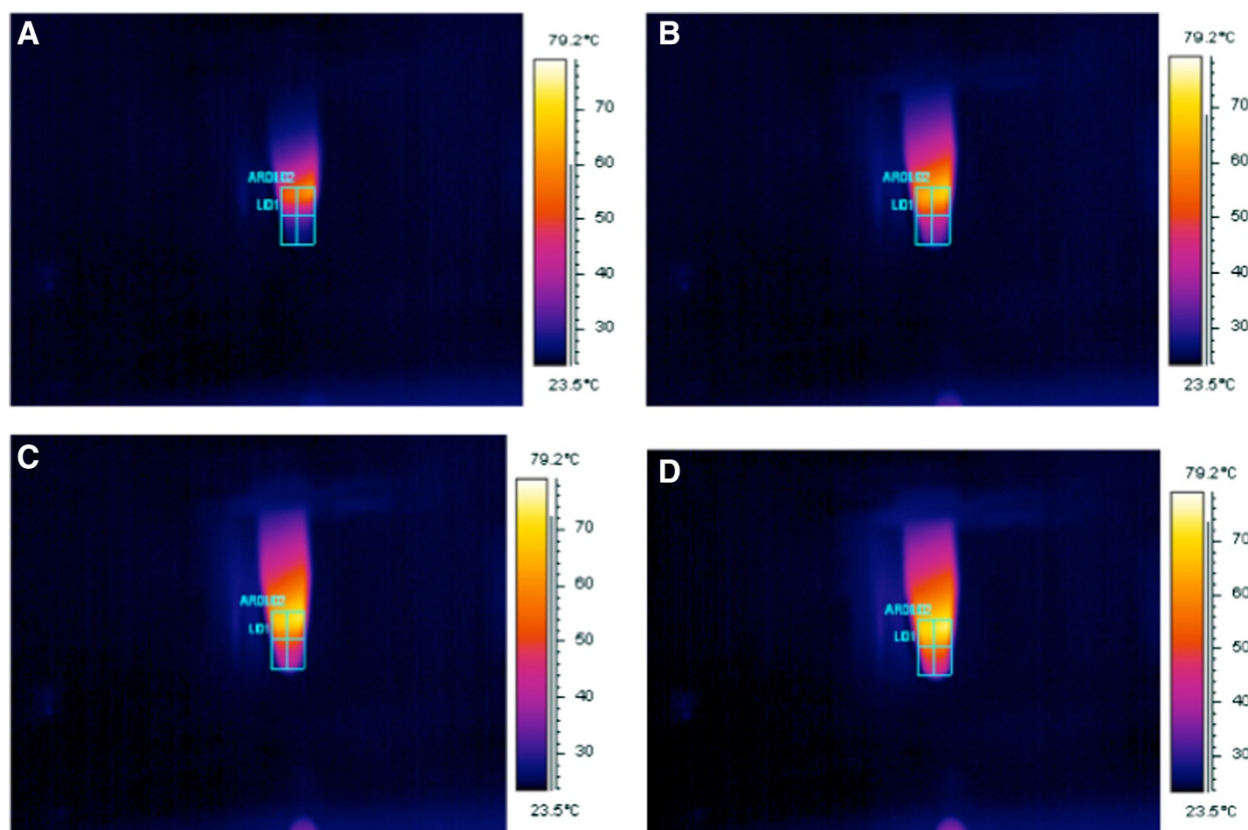
shown that POSS-PCU exerts a dual function: firstly by providing biocompatibility to CNTs, and secondly by acting synergistically with CNTs during exposure to NIR laser, resulting in a greater temperature increase and enhanced cell death.

We have successfully demonstrated that POSS-PCU-functionalized CNTs have the ability to thermally ablate



**Figure 6 Synergistic thermal effects of POSS-PCU and CNTs.** At both wattages (0.1 W and 1 W), POSS-PCU-CNT displays a higher rate of temperature increase and final temperature. POSS-PCU does not absorb in the NIR spectrum, but synergistically elevates the temperature profile of CNTs upon functionalization. At higher wattage, functionalization with 100% POSS-PCU shows a greater increase in temperature compared to 50% POSS-PCU.





**Figure 7** Thermograph of NIR irradiated POSS-PCU-CNT. POSS-PCU-CNT have a higher heating profile at 2.5 min (A), 5 min (B), 7.5 min (C), and 10 min (D), compared to unfunctionalized CNTs.

cancer cells when exposed to NIR laser. Furthermore, POSS-PCU-functionalized CNTs attain a higher maximum temperature as well as a higher cell death rate compared to unfunctionalized CNTs. This has significant implications in photothermal ablative techniques, as it shows that cell death would only occur when functionalized CNTs are exposed to NIR. It has to be noted that the depth of penetration of NIR is only effect up to a few centimeters; hence for non-superficial cancers, delivery of NIR can be achieved using optical fibers. Although the toxicology of CNTs due to its size is also a current point of contention, current scientific consensus dictates that functionalizing CNTs with biocompatible molecules can confer biocompatibility to CNTs. We have demonstrated a proof-of-concept that POSS-PCU nanocomposite polymers can render CNTs biocompatible as well as augmenting its intended clinical function of photothermal cancer ablation.

## Conclusions

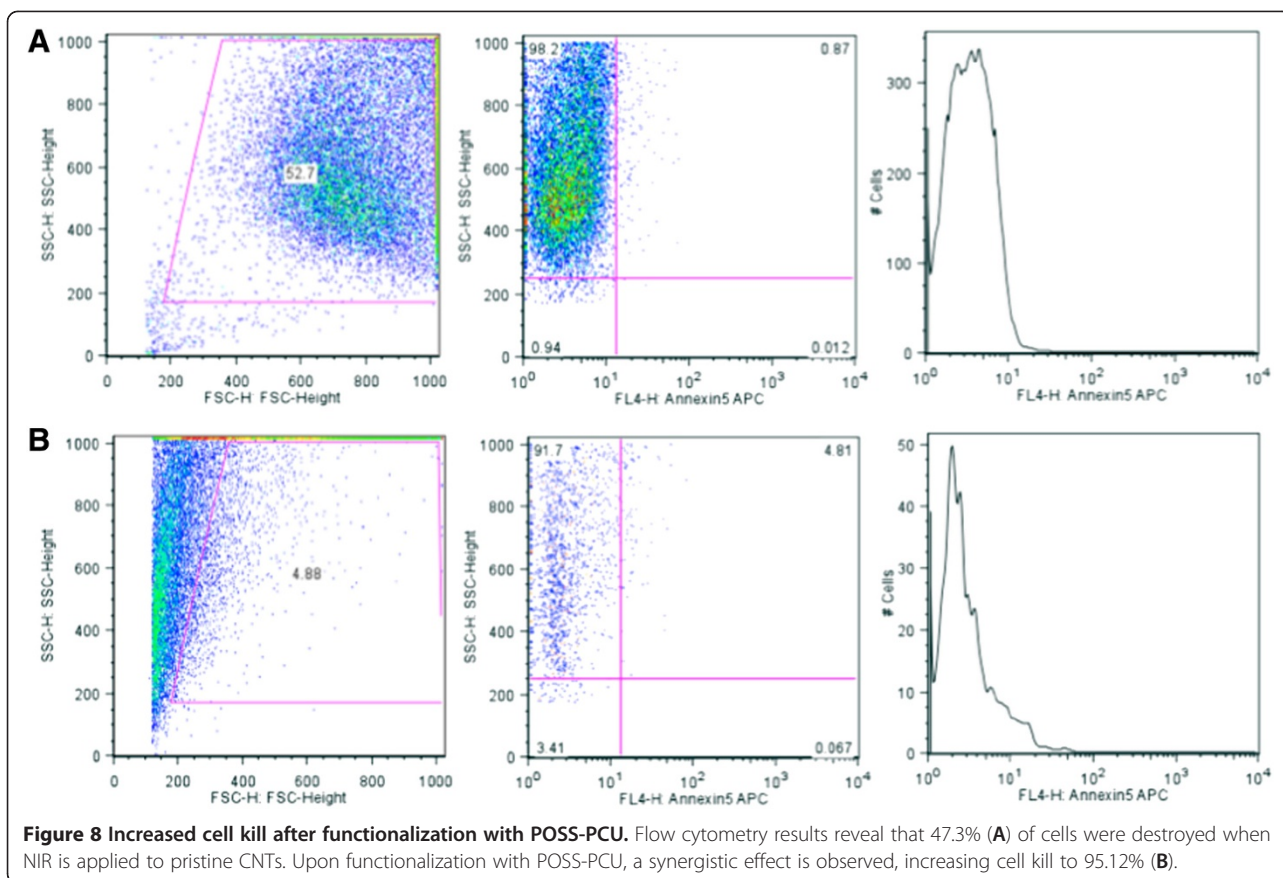
In this study, we present a model of synergism between CNTs and a novel nanocomposite polymer, POSS-PCU. This manifests itself in terms of temperature increase and the ability to thermally ablate cancer cells when

exposed to NIR laser. Furthermore, POSS-PCU confers solubility and biocompatibility to CNTs, making it an ideal functionalizing agent to address cancer. Future work regarding cancer-specific antibody conjugation for targeted therapy and *in vivo* studies would not only be imperative, but integral for elucidating the biological applications for CNTs and nanocomposite polymers. Taken together, this work underscores the dual functionality and synergistic effects of POSS-PCU functionalization, which could have implications and ramifications in a wide-ranging area of nanomedicine.

## Materials & methods

### Carbon nanotubes (CNTs) and dunctionalization with POSS-PCU

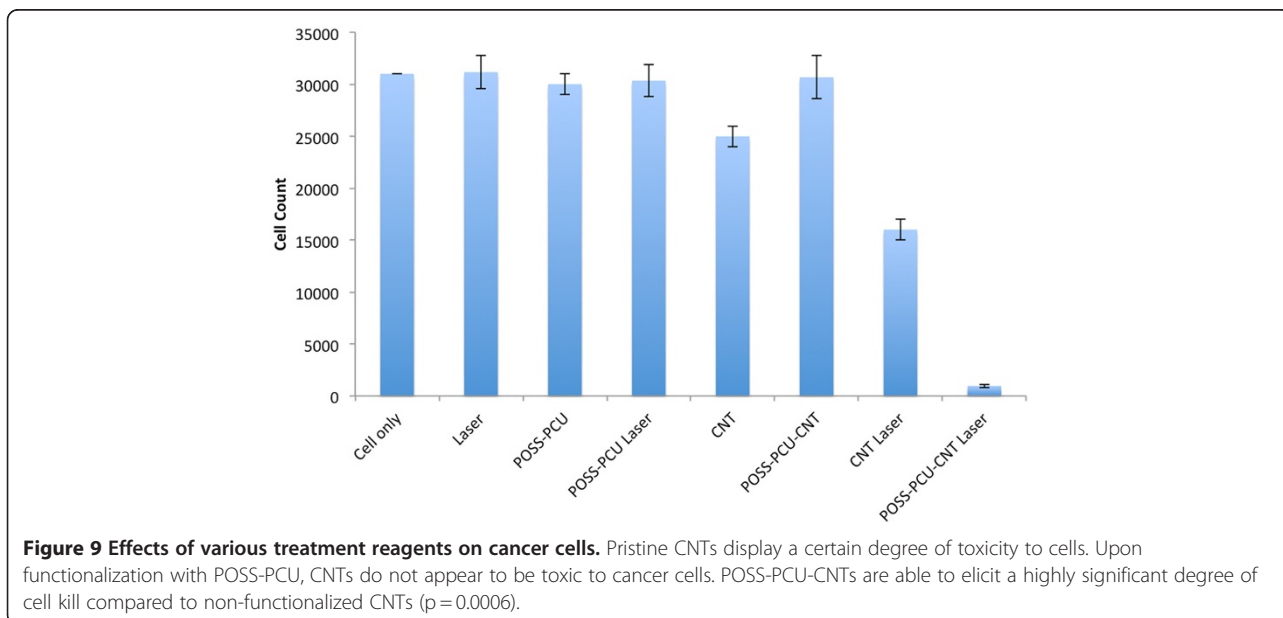
Pristine multi-walled carbon nanotubes (MWNTs) were obtained from Nanothinx S.A. (Rio-Patras, Greece). They had a mean diameter of 15 nm, and a mean length of 100 nm. The preparation of POSS-PCU has been described elsewhere[24]. Briefly, 1 mg of MWNT was mixed with 1 mg of POSS-PCU (50% and 100%) in a glass scintillation vial and 5 ml of water was added. The resultant mixture was sonicated in a sonicator bath (Grant Instruments, Cambridge, UK) for 60 minutes at room temperature.



### Characterization of POSS-PCU-CNT Complexes

Visual characterization was conducted using a JEM-2100 F transmission electron microscope (JEOL Ltd. UK). Optical absorbance of CNT, POSS-PCU, and

POSS-PCU-CNT was obtained using a LAMBDA 1050 UV/Vis/NIR (PerkinElmer, UK). Scan parameters were as follows: start (4000), number of scans (100), resolution (4), units (%T), end (400), interval (4), shuttle (interleaved).



The absorption spectrum was obtained via a second derivative and a 16-point reading.

### Near infrared radiation (NIR) laser system and temperature tracking

A NIR laser system (Thorlabs Ltd, UK) with a 808 nm laser diode was used to irradiate CNTs. The laser diode controller was set to either 2300 mA (1 W) or 1250 mA (0.5 W). To prevent over-heating, the temperature controller was set to 10 k $\Omega$ . Temperature tracking and thermal images were obtained using a Thermacam™ SC500 thermal camera (FLIR Systems, UK). CNTs were exposed to NIR irradiation for 10 minutes, and temperature changes were recorded every minute.

### Thermal ablation of cancer cells

A HT-29 human colorectal cancer cell line was used as a model for photothermal cancer ablation. Briefly, 3 million ( $3 \times 10^6$ ) cells were counted and seeded on a 96-well plate, with a theoretical cell count of 300,000 in each well. To each well, 100  $\mu$ l of treatment reagent was added (CNT, POSS-PCU, or POSS-PCU-CNT). All treatments were done in triplicates. NIR laser was then applied for 10 minutes. Cells were then incubated at 37°C with 5% CO<sub>2</sub> for 24 hours, and counted thereafter. All reagents pertaining to cell work were purchased from Invitrogen. Cell count was done using a MACSQuant Analyzer flow cytometry (Miltenyi Biotec Ltd, Surrey, UK).

### Curve fitting and statistical analyses

Curve fitting (least squares method) and statistical analyses were conducted at a 95% confidence interval using MATLAB® (MathWorks Inc, USA). Statistical significance testing was conducted using unpaired Student's *t*-test. *P* values of < 0.05 were considered statistically significant.

### Competing interests

The authors declare that they have not competing interests.

### Authors' contributions

AT, SYM, JR, GP planned, carried out, and wrote the manuscript. AMS conceived the study, participated in the design and coordination, and helped draft the manuscript. All authors read and approved the final manuscript.

### Acknowledgements

The authors would like to acknowledge funding from the Engineering and Physical Sciences Research Council (EPSRC) – Industrial CASE.

### Author details

<sup>1</sup>Centre for Nanotechnology & Regenerative Medicine, UCL Division of Surgery & Interventional Science, University College London, London, NW3 2QG, UK. <sup>2</sup>Department of Neurology & Neurological Sciences, Biomaterials & Advanced Drug Delivery Laboratory, School of Medicine, Stanford University, California, USA. <sup>3</sup>Department of Pharmacy, Faculty of Science, National University of Singapore, Singapore, Singapore. <sup>4</sup>Royal Free London NHS Foundation Trust Hospital, London, UK.

Received: 12 June 2012 Accepted: 16 July 2012  
Published: 31 July 2012

### References

1. Iijima S: **Helical microtubules of graphitic carbon.** *Nature* 1991, **354**(6348):56–58.
2. Hyder MN, Lee SW, Cebeci FC, Schmidt DJ: *Shao-Horn Y. Hammond PT: Layer-by-Layer Assembled Polyaniline Nanofiber/Multiwall Carbon Nanotube Thin Film Electrodes for High-Power and High-Energy Storage Applications.* *ACS nano*; 2011.
3. Tan A, Yildirim L, Rajadas J, De La Pena H, Pastorin G, Seifalian A: **Quantum dots and carbon nanotubes in oncology: a review on emerging theranostic applications in nanomedicine.** *Nanomedicine (Lond)* 2011, **6**(6):1101–1114.
4. Li J, Yap SQ, Yoong SL, Nayak TR, Chandra GW, Ang WH, Panczyk T, Ramaprabhu S, Vashist SK, Sheu F-S, *et al*: **Carbon nanotube bottles for incorporation, release and enhanced cytotoxic effect of cisplatin.** *Carbon* 2012, **50**(4):1625–1634.
5. Zhou F, Xing D, Ou Z, Wu B, Resasco DE, Chen WR: **Cancer photothermal therapy in the near-infrared region by using single-walled carbon nanotubes.** *J Biomed Opt* 2009, **14**(2):021009.
6. Kam NW, O'Connell M, Wisdom JA, Dai H: **Carbon nanotubes as multifunctional biological transporters and near-infrared agents for selective cancer cell destruction.** *Proc Natl Acad Sci USA* 2005, **102**(33):11600–11605.
7. Madani SY, Tan A, Dwek M, Seifalian AM: **Functionalization of single-walled carbon nanotubes and their binding to cancer cells.** *Int J Nanomedicine* 2012, **7**:905–914.
8. Dai H: **Carbon nanotubes: synthesis, integration, and properties.** *Accounts of chemical research* 2002, **35**(12):1035–1044.
9. Yu MF, Lourie O, Dyer MJ, Moloni K, Kelly TF, Ruoff RS: **Strength and breaking mechanism of multiwalled carbon nanotubes under tensile load.** *Science* 2000, **287**(5453):637–640.
10. Treacy MMJ, Ebbesen TW, Gibson JM: **Exceptionally high Young's modulus observed for individual carbon nanotubes.** *Nature* 1996, **381**(6584):678–680.
11. Aliev AE, Lima MH, Silverman EM, Baughman RH: **Thermal conductivity of multi-walled carbon nanotube sheets: radiation losses and quenching of phonon modes.** *Nanotechnology* 2010, **21**(3):035709.
12. Welscher K, Liu Z, Sherlock SP, Robinson JT, Chen Z, Daranciang D, Dai H: **A route to brightly fluorescent carbon nanotubes for near-infrared imaging in mice.** *Nat Nanotechnol* 2009, **4**(11):773–780.
13. Liu Z, Tabakman S, Welscher K, Dai H: **Carbon Nanotubes in Biology and Medicine: In vitro and in vivo Detection, Imaging and Drug Delivery.** *Nano research* 2009, **2**(2):85–120.
14. Ahmed M, Ghanbari H, Cousins BG, Hamilton G, Seifalian AM: **Small calibre polyhedral oligomeric silsesquioxane nanocomposite cardiovascular grafts: influence of porosity on the structure, haemocompatibility and mechanical properties.** *Acta Biomater* 2011, **7**(11):3857–3867.
15. Kannan RY, Salacinski HJ, Odlyha M, Butler PE, Seifalian AM: **The degradative resistance of polyhedral oligomeric silsesquioxane nanore integrated polyurethanes: an in vitro study.** *Biomaterials* 2006, **27**(9):1971–1979.
16. Chaloupka K, Motwani M, Seifalian AM: **Development of a new lacrimal drainage conduit using POSS nanocomposite.** *Biotechnol Appl Biochem* 2011, **58**(5):363–370.
17. Solouk A, Cousins BG, Mirzadeh H, Solati-Hashtjini M, Najarian S, Seifalian AM: **Surface modification of POSS-nanocomposite biomaterials using reactive oxygen plasma treatment for cardiovascular surgical implant applications.** *Biotechnol Appl Biochem* 2011, **58**(3):147–161.
18. Jungebluth P, Alici E, Baiguera S, Le Blanc K, Blomberg P, Bockoy B, Crowley C, Einarsson O, Grinnemo KH, Gudbjartsson T, *et al*: **Tracheobronchial transplantation with a stem-cell-seeded bioartificial nanocomposite: a proof-of-concept study.** *Lancet* 2011, **378**(9808):1997–2004.
19. Schipper ML, Nakayama-Ratchford N, Davis CR, Kam NW, Chu P, Liu Z, Sun X, Dai H, Gambhir SS: **A pilot toxicology study of single-walled carbon nanotubes in a small sample of mice.** *Nat Nanotechnol* 2008, **3**(4):216–221.
20. Cousins BG, Das AK, Sharma R, Li Y, McNamara JP, Hillier IH, Kinloch IA, Ulijn RV: **Enzyme-activated surfactants for dispersion of carbon nanotubes.** *Small* 2009, **5**(5):587–590.
21. Chen Z, Tabakman SM, Goodwin AP, Kattah MG, Daranciang D, Wang X, Zhang G, Li X, Liu Z, Utz PJ, *et al*: **Protein microarrays with carbon nanotubes as multicolor Raman labels.** *Nat Biotechnol* 2008, **26**(11):1285–1292.
22. Chen RJ, Zhang Y, Wang D, Dai H: **Noncovalent sidewall functionalization of single-walled carbon nanotubes for protein immobilization.** *J Am Chem Soc* 2001, **123**(16):3838–3839.

23. Shim M, Shi Kam NW, Chen RJ, Li Y, Dai H: **Functionalization of carbon nanotubes for biocompatibility and biomolecular recognition.** *Nano Lett* 2002, **2**:285–288.
24. Kannan RY, Salacinski HJ, Ghanavi JE, Narula A, Odlyha M, Peirovi H, Butler PE, Seifalian AM: **Silsesquioxane nanocomposites as tissue implants.** *Plast Reconstr Surg* 2007, **119**(6):1653–1662.

doi:10.1186/1477-3155-10-34

**Cite this article as:** Tan *et al.*: Synergistic photothermal ablative effects of functionalizing carbon nanotubes with a POSS-PCU nanocomposite polymer. *Journal of Nanobiotechnology* 2012 **10**:34.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

