

REVIEW

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Nanozymes-recent development and biomedical applications

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Abstract

Nanozyme is a series of nanomaterials with enzyme-mimetic activities that can proceed with the catalytic reactions of natural enzymes. In the field of biomedicine, nanozymes are capturing tremendous attention due to their high stability and low cost. Enzyme-mimetic activities of nanozymes can be regulated by multiple factors, such as the chemical state of metal ion, pH, hydrogen peroxide (H₂O₂), and glutathione (GSH) level, presenting great promise for biomedical applications. Over the past decade, multi-functional nanozymes have been developed for various biomedical applications. To promote the understandings of nanozymes and the development of novel and multi-functional nanozymes, we herein provide a comprehensive review of the nanozymes and their applications in the biomedical field. Nanozymes with versatile enzyme-like properties are briefly overviewed, and their mechanism and application are discussed to provide understandings for future research. Finally, underlying challenges and prospects of nanozymes in the biomedical frontier are discussed in this review.

Keywords: Nanozyme, Oxidative stress, Reactive oxygen species, Disease therapy

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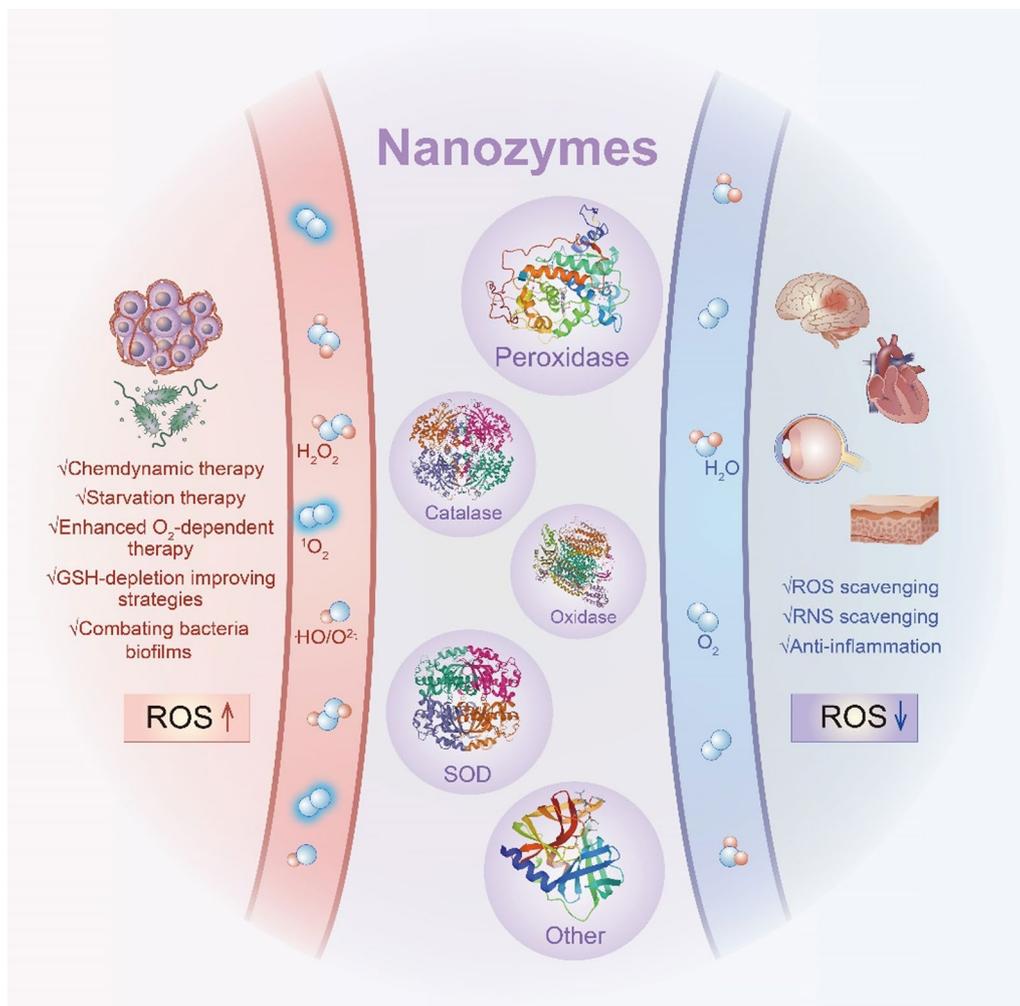
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Graphical Abstract

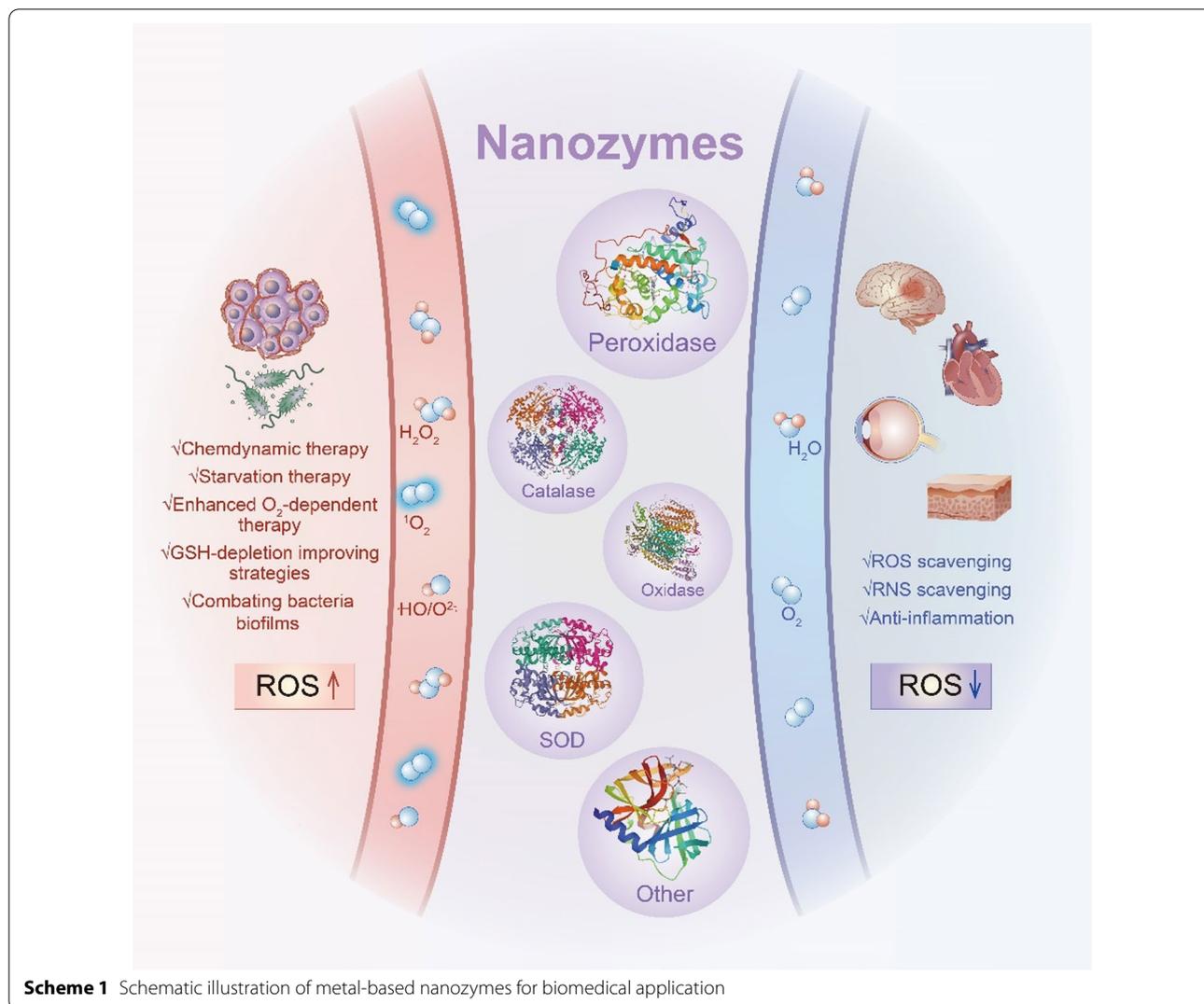


Background

Nanozymes, as artificial enzymes, are nanomaterials with enzyme mimetic activities, which have attracted considerable interest due to their relatively higher physicochemical stability against harsh environments, higher durability, and lower costs than natural enzymes [1]. In the past decades, numerous nanomaterials have been revealed to elucidate the oxidase (OXD), glucose oxidase (GOD), peroxidase (POD), catalase (CAT), superoxide dismutase (SOD), and glutathione peroxidase (GPx) mimicking activities with extensive biomedical applications [2, 3]. At present, nanozymes are mainly composed of metal and metal oxides, since the metallic active center can effectively mimic the catalytic electronic redox process enabled by natural enzymes. Specifically, the enzyme-mimicking activities are affected by various factors, such as the oxidation states of the metallic center,

reduction agent, temperature, and pH in the surrounding environment [4, 5]. Interestingly, the disease features differ from normal tissues provide typical therapeutic options for rational design and application of nanozymes in biomedicine. It is well known that the tumor microenvironment (TME) exhibits higher redox potential levels than the normal tissues. Such characteristics in the tumor can catalyze enzyme-like activities of the nanozymes [6–8]. For instance, metallic ions (such as Fe^{3+} , Cu^{2+} , and Mn^{4+} , etc.) can be reduced to lower-valent metallic ions (Fe^{2+} , Cu^+ , and Mn^{3+}) by intracellular GSH [9–11]. Hence, the POD activities and catalytic efficiency of the nanozymes could be altered remarkably in the specific pathological microenvironment.

Although numerous nanozymes having been made in the biomedicine field, it is still challenging to obtain a fundamental insight into the key factors that affect the



catalytic performance, enzymatic-like properties, as well as the substrate selectivity of nanozymes, on the basis of the interplay between intrinsic structure and extrinsic environment [12, 13]. Moreover, the catalytic mechanisms of metal oxide nanozymes are pivotal to rationally design novel nanozymes with inherent catalytic capacities and this approach has been widely applied in biomedicine as a controllable multifunctional platform [14, 15]. Recently, many types of nanoparticles with inherent catalytic properties have been reported to achieve various biomedical applications, including oxygen-dependent tumor therapy, radiotherapy, chemodynamic therapy, bacterial infection diseases, and reactive oxygen species (ROS)-related diseases, etc. [16–20]. Therefore, recent advances in the field of nanozyme's biomedical application may bring new insights into the popularization of nanoparticles in the treatment of the biomedical field.

In this review, Different metal- or metal-based nanozymes have been overviewed and described as classified according to their catalytic active center, which significantly impacts the functionalities and activities of the nanozymes during certain catalytic reactions. The versatile enzymes-like properties, mechanism of nanozymes, and the factors that affect the catalytic performance are initially summarized. Then, recently administrated strategies of nanozymes in the therapeutic frontier have been introduced (Scheme 1). Finally, the current challenges of the development of nanozymes and prospects are discussed. We hope that the present review will be of significant benefit for different biomedical fields and provide insightful ideas for the design and development of nanozymes.

Cerium-based nanozymes

Cerium (Ce)-based nanoparticles have been exploited for biomedical applications since they exhibit multiple enzyme-like activities such as catalase- (CAT), peroxidase- (POD), cytochrome c oxidase-, and superoxide dismutase-mimetic (SOD) functions [21–23]. The underlying mechanism of nanoceria-mediated enzymatic reactions was associated with the chemical state of the cerium element. The reduction state (Ce^{3+}) and oxidation state (Ce^{4+}) affect the enzyme-like performance of CeO_2 [24, 25]. Singh et al. reported that CeVO_4 nanoparticles exhibited cytochrome c oxidase (CcO) activity, which can dismutase oxygen into water at physiological pH conditions due to the electron transfer between Ce^{3+} , Ce^{4+} , and V^{5+} [26]. Interestingly, the ratio of $\text{Ce}^{3+}/\text{Ce}^{4+}$ of CeVO_4 has been shown to affect the CcO-like activity of CeVO_4 . The authors suggested that the lower ratio $\text{Ce}^{3+}/\text{Ce}^{4+}$ had higher CcO-like activity in CeVO_4 while the higher ratio of lowered valence states (Ce^{3+}) corresponds to the higher SOD-like activity. The present study demonstrated the enzyme-mimetic activities of Ce-based nanozymes associated with available oxidation states. Importantly, recent studies manifested that the surface defect characteristics of CeO_2 could partly affect the enzyme-like capability. Recently, Wang et al. revealed the SOD- and CAT mimicking mechanisms of CeO_2 by first-principles calculations [27]. Their results suggested that oxygen vacancies played critical roles to scavenge superoxide anion ($\text{O}_2^{\cdot-}$) and hydrogen peroxide (H_2O_2). The oxygen vacancies impacting enzyme-mimic activities were mainly ascribed to the reduced activation energy and the formation of the intermediate species. This research suggests that the reduction of activation energy by CeO_2 is critically important in exploring the catalytic processes. Their catalytic activity is substantially affected by the intrinsic properties (e.g., dimensions, oxygen vacancy) and physiological factors such as pH, GSH, and temperature. Meanwhile, the oxygen vacancy concentrations were highly dependent on the particle size of CeO_2 [28].

Despite the advancements made in biomedicine, nanoceria has key limitations that need to be overcome. For instance, the precise regulation of the enzyme activities remains challenging for nanoceria to meet the biomedical application. Moreover, the toxicity of nanoceria against normal tissues still presents great challenges in achieving clinical application. Previous studies have explored the toxicity effect of CeO_2 of varied shapes in RAW264.7 cell line. Compared with the cubic/octahedral morphological nanoparticles, the rod-like CeO_2 shows increasing serum concentration of tumor necrosis factor alpha (TNF- α) and lactate dehydrogenase (LDH) release, demonstrating the morphological-dependent

cytotoxicities [29]. The biocompatibility of CeO_2 nanoparticle was demonstrated in further in vivo studies in rats when administering CeO_2 of as high as 20 mg/kg [30]. These observations provide an important inspiration for other nanozyme to achieve higher therapeutic effect with minimal toxicity. To solve the low selectivity and poor therapeutic effects, it is highly appealing to design controllable nanoceria-based therapeutic systems. Noteworthy, the catalytic behavior of nanoceria is also determined by physiologically catalytic environments. By controlling enzyme activities in a TME or light stimuli, a desired therapeutic effect with low tissue damage could be achieved. Zhu et al. reported that the self-regulated nanoceria-doped poly-(cyclopentadithiophene-alt-benzothiadiazole) (SPN-C23) as smart nanoplatforms for tumor photodynamic therapy (PDT) [31]. When near-infrared (NIR) laser was irradiated against the tumor tissue with acidic microenvironment (pH=6.5), SPN-C23, acted as a ROS converter, was able to amplify PDT damage against tumor tissues through catalyzing $\text{O}_2^{\cdot-}$ to produce H_2O_2 (Fig. 1A). When exposed to the normal microenvironment (pH=7.4), Ce^{4+} of SPN-C23 could transform $\text{O}_2^{\cdot-}$ to O_2 with generated Ce^{3+} due to oxygen vacancies in the surface of nanoceria. Consequently, SPN-C23 exhibited high singlet oxygen sensor green (SOSG) fluorescence enhancement at pH=6.5 and possessed relatively low fluorescence intensity at pH=7.4 (Fig. 1B), demonstrating that pH-dependent single oxygen species ($^1\text{O}_2$) production of SPN-C23. SPN-C23 also possesses higher fluorescence at pH=6.5 with NIR laser irradiation, indicating that the generation of H_2O_2 (Fig. 1C). PDT efficiency of SPN-C23 was evaluated on in vivo 4-T1 xenograft tumor model. Under NIR laser irradiation, the growth of the tumor in SPN-C23 treated group was significantly suppressed after intravenous injection of SPN-C23 for 16 days (Fig. 1D). Tissue damage by SPN-0 and SPN-C23 were examined by histological (H&E) staining respectively. The damaged area of the healthy muscle tissues in the SPN-C23 group was significantly decreased as compared to the SPN-0 treatment group (Fig. 1E). Their study provides an effective approach by utilizing ceria-based nanozyme to regulate PDT against cancer with high biocompatibility.

ROS as the signaling messengers play an essential function in the physiological signal transduction pathway [32]. Endogenous and low concentrations of ROS can be produced from normal metabolic processes in the living cells [33, 34]. However, ROS are highly toxic at higher concentrations and can damage protein, RNA, and DNA, leading to cell death [32, 35]. Therefore, ameliorating the oxidative stress induced by ROS has been proved to be beneficial in pathological therapy, such as age-related macular degeneration (AMD), traumatic brain injury,

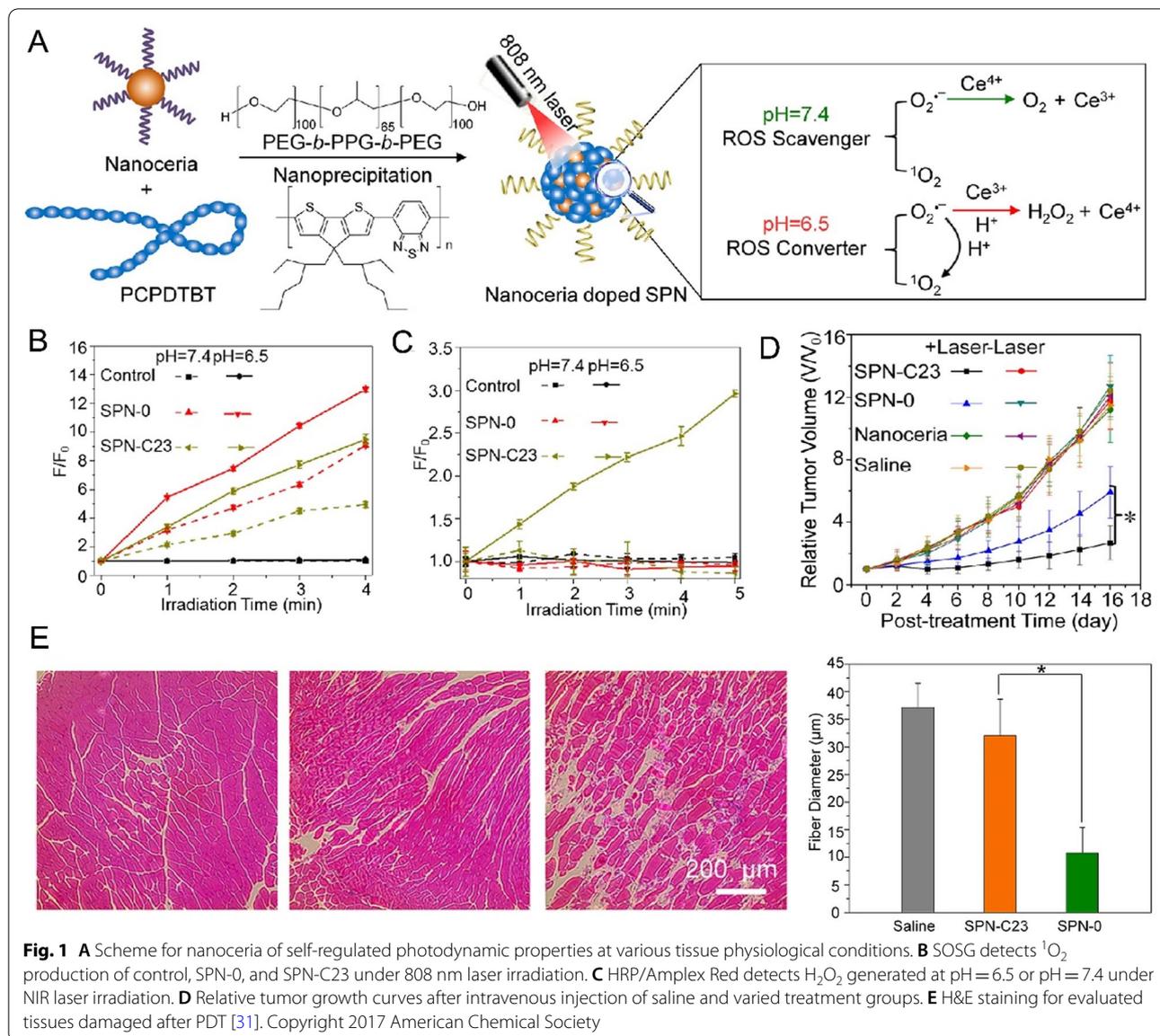


Fig. 1 **A** Scheme for nanoceria of self-regulated photodynamic properties at various tissue physiological conditions. **B** SOSG detects 1O_2 production of control, SPN-0, and SPN-C23 under 808 nm laser irradiation. **C** HRP/Amplex Red detects H_2O_2 generated at pH = 6.5 or pH = 7.4 under NIR laser irradiation. **D** Relative tumor growth curves after intravenous injection of saline and varied treatment groups. **E** H&E staining for evaluated tissues damaged after PDT [31]. Copyright 2017 American Chemical Society

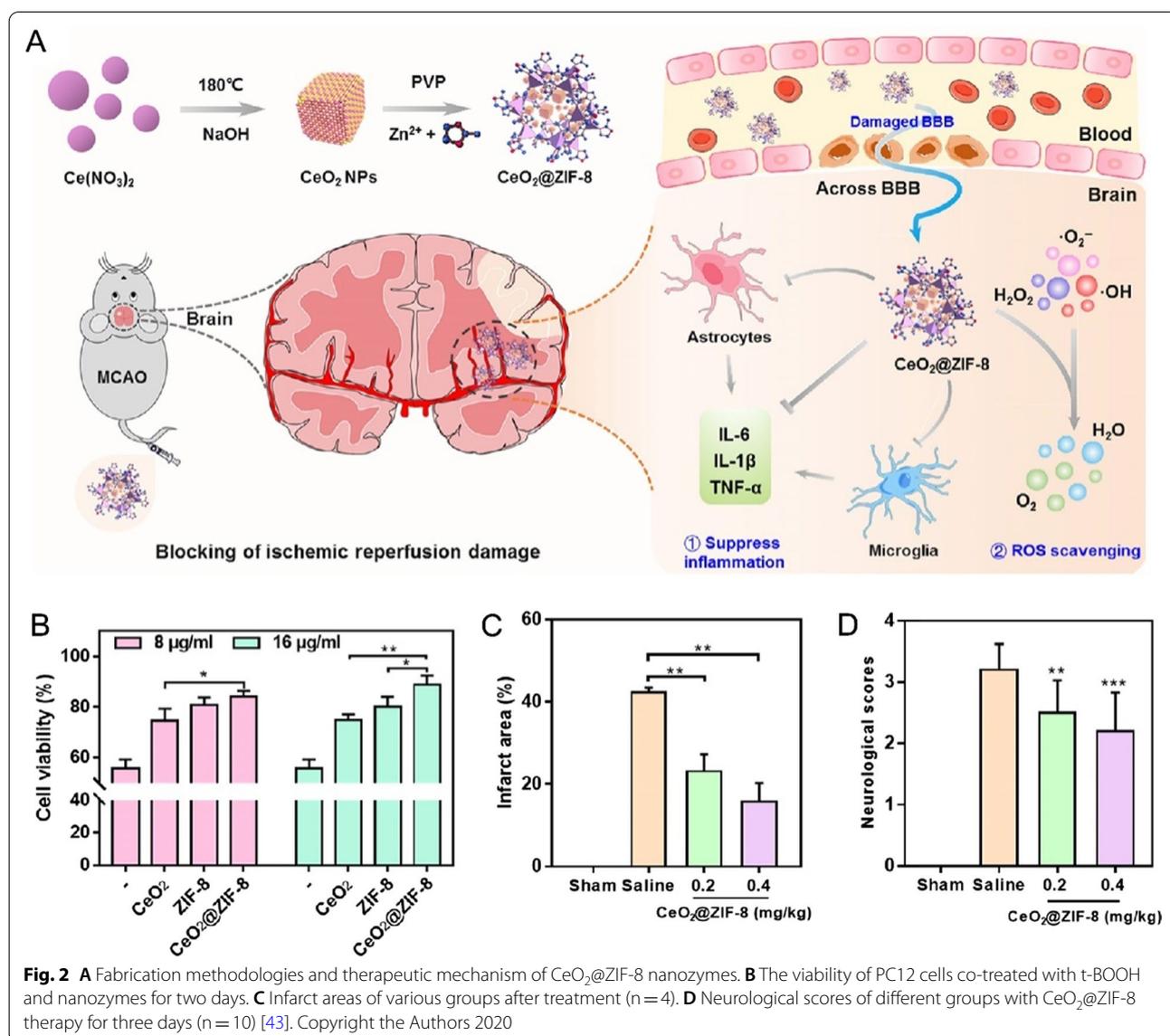
and ischemic disease [23, 36–38]. Nanoceria is often employed as an antioxidant agent to treat ROS-relevant diseases. For instance, AMD is associated with irreversible ROS damage against the macular that may lead to blindness [39, 40]. Generally, evidence showed that AMD pathology can be treated by counteracting the overproduction of ROS. Mitra et al. reported a nanoceria with dominated Ce^{3+} that can scavenge the ROS, such as H_2O_2 and $\cdot OH$, and inhibit neovascularization formation [41]. Moreover, Yan et al. developed a single-atom Pt/ CeO_2 for traumatic brain injury treatment [36]. Compared with CeO_2 , Pt/ CeO_2 exhibits highly enzymatic activity that can scavenge $O_2^{\cdot-}$, $\cdot NO$, and $\cdot OH$. Furthermore, in vivo administration of Pt/ CeO_2 to C57BL/6 mice can

significantly improve the wound recovery by up to 100%, higher than the mice in the untreated group with 50% of the wound closure. Similarly, ischemic stroke is one of the inflammations associated with excessive ROS generation. With excellent antioxidation activity, ceria nanozymes have been applied for efficient treatment of reperfusion-induced injury in ischemic stroke [42]. Although nanoceria with ROS eliminating ability can protect the cells from ROS damage, physiological stability and biocompatibility remained a challenge towards further clinical prospects. Regarding the present issue, He et al. designed a multi-functional nano-system for treating ischemic stroke with prolonged blood circulation time and higher biosafety [43]. $CeO_2@ZIF-8$ were prepared by in-situ capping of

CeO₂ with zeolitic imidazolate framework-8 (ZIF-8) (Fig. 2A). In their design, ZIF-8 was served as peroxidase to maintain the activity of CeO₂, as well as to enhance the penetration and accumulation of CeO₂ to brain tissue. In PC12 neuronal cells, CeO₂@ZIF-8 can effectively protect cells from tert-butyl hydroperoxide (t-BOOH) induced cell apoptosis (Fig. 2B). In vivo administration of CeO₂@ZIF-8 could significantly reduce the infarcted area and increased the neurological scores of mice, confirming that CeO₂@ZIF-8 can effectively treat mice with ischemic stroke (Fig. 2C, D).

Another interesting application of nanoceria was the elimination of extracellular DNA (eDNA) for anti-biofilms. Biofilms, a community of bacteria cells, prevents a majority of approaches to treat bacterial infectious

diseases in humans. eDNA is a crosslinking component of bacteria biofilm, which provides a potential survival benefit for bacterial infection. It has been established that eDNA has a profound impact on the process of biofilm formation. Therefore, eradicating eDNA is an effective strategy to treat biofilm infection. Functionalization of CeO₂ offers a versatile approach in combating biofilm formation and bacterial infection. Liu et al. designed metal-organic framework (MOF)/Ce-based nanozymes to combat biofilms [44]. The MOF/Ce nanozymes with DNase mimic activities could not only inhibit the biofilm formation but also eradicate established biofilm matrix components by hydrolyzing eDNA. As compared to primitive MOF, MOF/Ce nanozymes possessed higher bactericidal activity via co-incubation. The mechanism



of the bactericidal activity of MOF/Ce nanozymes is that two adjacent Ce^{4+} could bind to the oxygen atom of the phosphate group by withdrawing the electrons, resulting in phosphodiester linkage cleaving [45].

Ferrum-based nanozymes

Ferrum (Fe)-based nanoparticles have gained extensive attention for various biomedical applications due to their magnetic resonance imaging (MRI) performance, POD-mimetic activity, and CAT mimetic properties [46, 47]. Huo et al. reported that Fe_3O_4 nanoparticles and GOD co-encapsulated into the mesoporous silica nanoparticles could be effective for tumor catalytic therapy [48]. The intrinsic POD-mimetic activity of Fe_3O_4 could generate a considerable amount of $\cdot\text{OH}$ from H_2O_2 produced by the GOD catalysis from glucose. Importantly, the efficiency of $\cdot\text{OH}$ production efficiency involving the amount of Fe^{2+} . Compared with the reaction kinetics of Fe^{2+} with H_2O_2 ($40\text{--}80 \text{ L mol}^{-1} \text{ s}^{-1}$), the reaction rate of Fe^{3+} ($9.1 \times 10^{-7} \text{ L mol}^{-1} \text{ s}^{-1}$) with H_2O_2 is relatively low [49, 50]. In addition, several reports have established links between POD-mimetic activity and CAT mimicking property of Fe_3O_4 under various conditions. Gao et al. investigated the catalytic mechanism of $\text{Fe}^{2+}/\text{Fe}^{3+}$ proportion in Fe_3O_4 with the use of either a reducing agent (NaBH_4) or an oxidizing agent (NaIO_4) [51]. They found that increased $\text{Fe}^{2+}/\text{Fe}^{3+}$ proportion of Fe_3O_4 could be achieved by NaBH_4 treatment, with the correspondingly enhanced peroxidase-like activity of Fe_3O_4 . On the contrary, decreased proportion of $\text{Fe}^{2+}/\text{Fe}^{3+}$ treated by NaIO_4 reduces the POD-like activity of Fe_3O_4 . In addition, the authors showed that the pH, temperature, and dimension of nanoparticles could effectively influence the enzyme-like activity of the Fe_3O_4 . These pieces of evidence provide insights into the metallic species and their impacts on the enzyme activities of Fe_3O_4 . Another study by Wang et al. showed that reducing agents in the physiological environment (L-cysteine/NADPH) can restore Fe^{3+} to Fe^{2+} on the surface of Fe_2O_3 , enhancing the abilities of $\cdot\text{OH}$ generation by Fe_2O_3 [52]. To further understand the impact of the physiological environment, Chen et al. investigated the enzymes-like properties of Fe-oxide (Fe_3O_4 and $\gamma\text{-Fe}_2\text{O}_3$) nanoparticles at the cellular level by electron spin resonance (ESR) and multi-parameter water quality meter [53]. They demonstrated that Fe_3O_4 and $\gamma\text{-Fe}_2\text{O}_3$ nanoparticles exhibit higher POD-like activities at $\text{pH}=4.8$, while CAT-like activities were observed at $\text{pH}=7.4$. The controllable enzymatic activities in targeted microenvironments provide flexibility and high sensitivity for diverse biomedical applications. Owing to the role of reducing agents in enzyme-like activity, it is feasible that the Fe^{3+} is reduced to Fe^{2+} by the over-expressed GSH in tumor tissues, contributing to the

elevation of the ROS generation and resultant tumor destruction.

Fe-based nanozyme with CAT-like activity have been reported to broaden the therapy for ROS-involved cerebral malaria. Zhao et al. designed and synthesized recombinant human ferritin (HFn) modified Fe_3O_4 (Fenozyme) with blood–brain barrier crossing and ROS-scavenging activity to treat cerebral malaria [54]. From the in vivo murine cerebral malaria experiment, it has been revealed that fenozyme possessed significant ROS scavenging abilities of Fe_3O_4 and prominent blood–brain barrier crossing performance of HFn. Administration of the fenozyme can significantly ameliorate the lesion of cerebral malaria and enhance the survival rate of infected mice induced by the parasite.

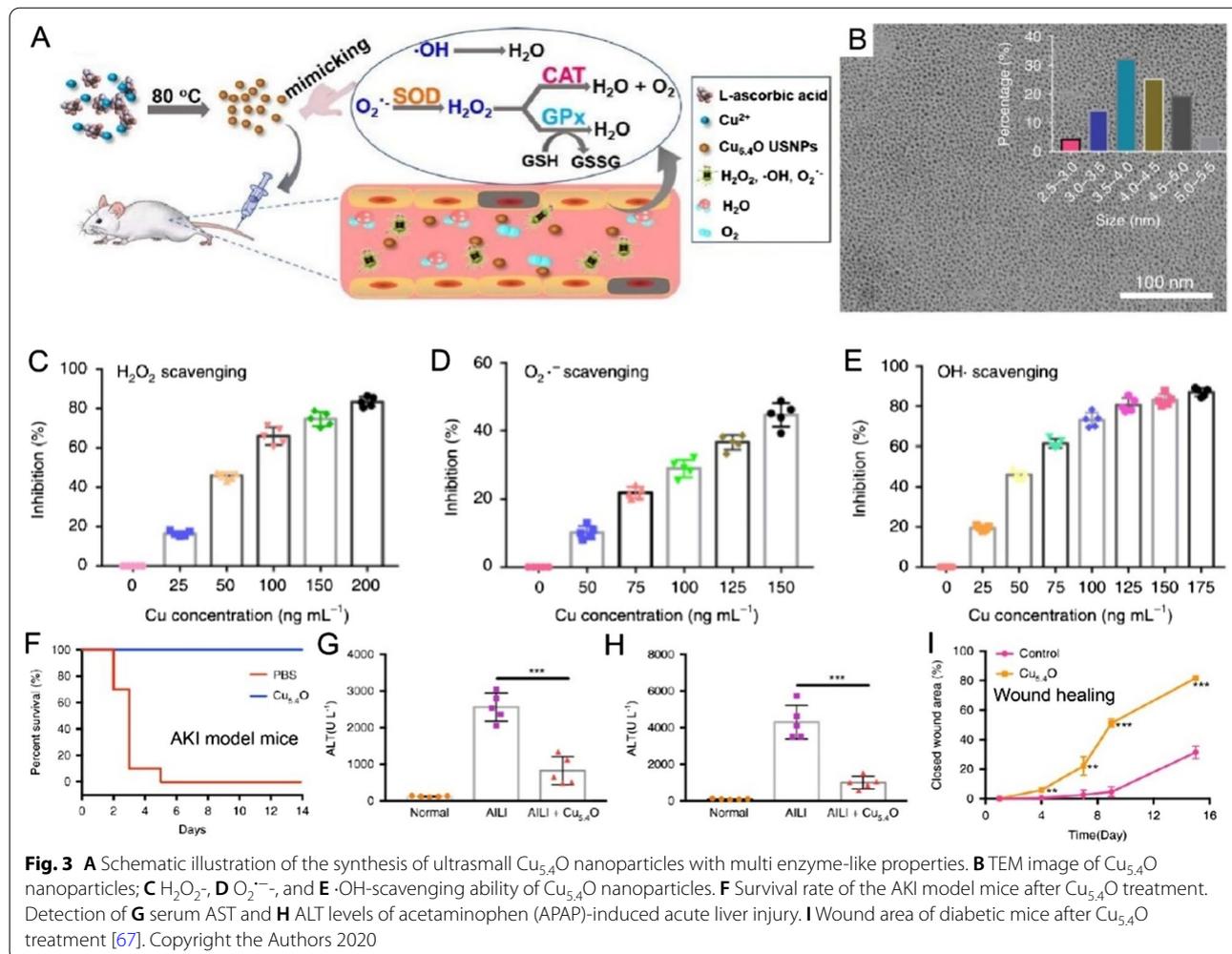
Copper-based nanozymes

Copper (Cu) oxide nanomaterials have received significant attention due to their enzyme-mimetic activity [55–57]. The POD-mimetic activity of Cu oxide nanoparticles has focused on ROS production activity by Fenton-like catalysis by Cu^+ and/or Cu^0 [58]. Besides, the reaction rate of Cu^+ with H_2O_2 was high than Fe^{2+} because the redox potential of $\text{Cu}^{2+}/\text{Cu}^+$ is lower, indicating that Cu^+ exhibited relatively higher POD-like activities than Fe^{2+} [59, 60]. Similar to Fe^{2+} , the Fenton-like activity of Cu-based nanoparticles is the potent antitumor agent. In tumor microenvironment mediated therapy, intratumoral reductive agents (such as GSH) can reduce Cu^{2+} to Cu^+ species, leading to high selectivity and efficiency. For instance, Ma et al. reported copper-amino acid mercaptide nanomaterials (Cu-Cys) with GSH depletion and Cu^+ production within the tumor microenvironment [60]. After their accumulation at the tumor sites, Cu^+ species reacted with H_2O_2 and produced sufficient ROS, initiating the tumor apoptosis via a Fenton-like reaction. Besides tumor treatment, the POD-like activity of Cu oxide nanoparticles has been employed as antibacterial treatment. Xi et al. designed Cu/carbon nanozymes that can effectively kill Gram-positive and Gram-negative bacteria [61]. Especially, they confirmed that the enzyme-like properties were dependent on the chemical state of Cu. Cu^0 exhibits high POD-like activities than Cu^{2+} and kill bacteria by Fenton-like reaction under H_2O_2 -rich environment. The research progress of Cu-based nanozymes shows their promising biomedicine applications in the targeted disease microenvironment.

Regulating the activity of nanozymes by utilizing an external stimulus may be highly desirable. It has been demonstrated that visible light could modulate the antibacterial activity of CuO. Nurul Karim et al. fabricated a CuO-nanorod that exhibits POD-mimic activities, and the enzymes-like activities were controlled by visible

light [62]. Due to the favorable band structure (1.44 eV), CuO exhibit relatively high POD-like efficiency in the presence of light to kill Gram-negative bacteria efficiently. Besides, the Cu₂O nanoparticles can mimic the cytochrome c oxidase activity [63]. It is worth noting that Cu_xO nanoparticles synthesized in the presence of the structure-directing agent phenylalanine (Phe) can mimic multienzyme activities, such as GPx, POD, superoxide dismutase (SOD), and catalase, enabling the ROS scavenging performance for Parkinson's disease amelioration [64]. The potential mechanism is that Cu_xO could reduce the intracellular ROS levels and alleviating oxidative damages. On the other hand, Korschelt et al. reported a copper hydroxide (Cu(OH)₂) nanoparticle with glycine functionalized (Gly-Cu(OH)₂) that served as SOD mimics, eliminating O₂^{•-} radicals generated while smoking [65]. The mechanism of the SOD-like activity by Gly-Cu(OH)₂ was further investigated. They found that the reduction and re-oxidized by Cu²⁺ of Gly-Cu(OH)₂ play a prominent role to eliminate O₂^{•-}. Importantly, Lin

et al. reported an interesting Cu²⁺-tannic acid (TA) complex nanozyme (Cu-TA) that exhibits SOD-like activity and catalase-like activity for ROS scavenging [66]. The high SOD-like activity of Cu-TA was dependent on the coordination of Cu²⁺ and TA, which enhance the redox potential of Cu²⁺. Additionally, the Cu-TA nanozyme can eliminate ·OH and decompose H₂O₂ to H₂O. The ROS scavenging efficiency of nanozyme was further investigated after Cu-TA being loaded into the cigarette filter, and the scavenging efficiency was calculated to be 87.0%, 68.9%, and 34.6% of O₂^{•-}, H₂O₂, and ·OH, respectively. To increase therapeutic benefit and reduce systemic toxicity, it is highly desirable to develop the Cu-based nanoparticles with higher antioxidant activity. Other groups also reported multienzymes-like activities of Cu-based nanoparticles, Liu et al. fabricated ultrasmall Cu_{5,4}O nanoparticles with extensive ROS scavenging efficiency and abilities to treat ROS-related disease (Fig. 3) [67]. They demonstrated that Cu_{5,4}O could exhibit CAT-, SOD-, and GPx-mimicking for enhanced treatment effect against



various ROS-mediated diseases at extremely magnitude such as acute kidney injury (AKI) (2 $\mu\text{g}/\text{mg}$ for treatment in vivo), liver damage (6 $\mu\text{g}/\text{mg}$ for treatment in vivo), as well as wound healing. Moreover, pharmacokinetics and biodistribution experiments revealed that $\text{Cu}_{5.4}\text{O}$ possesses highly renal clearance advances and outstanding biocompatibility.

Manganese-based nanozymes

Manganese (Mn)-oxide nanoparticles have been demonstrated with intrinsic activities of POD-, GPx-, CAT-, and SOD-like activities due to the variable Mn valent states [68, 69]. In the presence of $\text{Cl}^-/\text{HCO}_3^-$ environment, the Mn^{2+} exhibit POD-like property that can decompose H_2O_2 into $\cdot\text{OH}$ for tumor therapy [9, 70]. Similar to Fe^{3+} and Cu^{2+} , the GSH could reduce $\text{Mn}^{3+}/\text{Mn}^{4+}$ into Mn^{2+} in the tumor microenvironment, the depletion of GSH could sensitize the ROS-based therapeutic strategies such as chemodynamic therapy and photodynamic therapy [71–74]. Recently, Fu et al. constructed Mn-doped calcium phosphate nanoparticles with loaded GOD (GOD-MnCaP) for tumor therapy [70]. Under the tumor microenvironment, GOD could catalyze the intracellular glucose into H_2O_2 for Mn^{2+} -mediated $\cdot\text{OH}$ generation and gluconic acid for enhanced Mn^{2+} -mediated reaction ($\text{Mn}^{2+} + \text{H}_2\text{O}_2 \rightarrow \text{Mn}^{3+} + \cdot\text{OH} + \text{OH}^-$) [75]. Mn-containing nanomaterials have also been served as CAT-like nanozymes for O_2 generation, capturing widespread attention in O_2 mediated therapeutic strategy [72, 76–78]. The Mn-oxide nanoparticles have been established as ROS scavenging agents for the treatment of oxidation-stress mediated diseases [69, 79, 80]. Decreasing ROS levels is one of the important therapeutic mechanisms for Mn-oxidated nanoparticles. However, the targeted delivery strategies remain a great challenge for current Mn-based nanozymes to meet different disease criteria. Based on these challenges, a material design approach may provide an appropriate option to satisfy such demand. Shi et al. reported the Mn_3O_4 encapsulated erythrocyte with T7 peptides functionalization system ($\text{Mn}_3\text{O}_4@$ nanoerythrocyte-T7) for ischemic stroke protection [81]. After being accumulated into the infarcted sites via T7 peptides targeting, the $\text{Mn}_3\text{O}_4@$ nanoerythrocyte-T7 can efficiently scavenge the ROS and supply oxygen before thrombolysis stroke, with O_2 supply to the hemoglobin in the erythrocyte after thrombolysis. Selective targeting to ischemic stroke provides an attractive strategy to achieve a strong Mn-based nanozymes therapeutic effect.

Interestingly, the GOD enzyme-like activity of MnO_2 nanosheets was reported by Tang et al. [82]. They synthesized MnO_2 nanosheets (M-NSs) by a one-step wet-chemical method that had high glucose affinity and

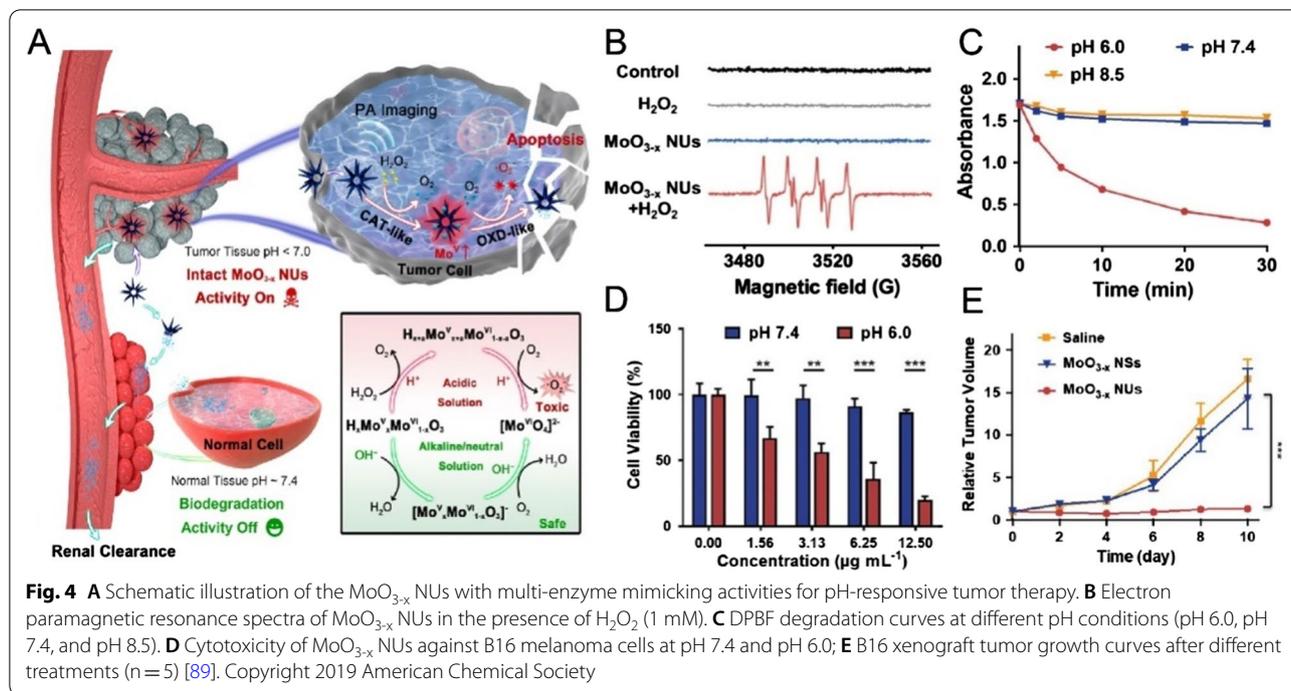
thermal stability as compared to the natural GOD. Under NIR laser irradiation, M-NSs could achieve the photothermal conversion while the glucose was gradually transformed to gluconic acid and H_2O_2 , resulting in glucose deprivation enhanced photothermal therapy. Such inorganic nanozyme with GOD-like activity provides a new strategy for the evolution of glucose deprivation and ROS-mediated cancer therapy.

Molybdenum-based nanozymes

Molybdenum (Mo) nanoparticles have attracted considerable attention as nanozymes [83–85]. There have been many reports regarding the catalytic activity of SOD, CAT, OXD, and sulfite oxidase, etc. [86, 87]. Although Mo-based nanozymes display outstanding enzyme-mimicking performance, it is still difficult to further expand their application to biomedicine. One of the main constraints for the application of Mo-based nanozymes is that these nanozymes carry both antioxidative and oxidative activities simultaneously, and may fail in their application to inhibit oxidative-mediated injury with satisfied outcome. Han et al. synthesized MoO_{3-x} nanodots with CAT- and SOD- mimic activities for Alzheimer's disease treatment [88]. However, the OXD-activities should be considered and may affect the therapeutic benefits of MoO_{3-x} . Considering these issues, new strategies should be developed to design intelligent nanozymes with controlled enzyme activity at specific microenvironments with acidity. Hu et al. constructed MoO_{3-x} nanourchins (MoO_{3-x} NUs) with pH-dependent multi-enzymatic activity for tumor-specific therapy (Fig. 4) [89]. Under normal physiological pH environment, MoO_{3-x} possessed high biocompatibility due to their stimuli-responsive biodegradation behavior. MoO_{3-x} exhibits excellent catalase enzyme activity under acidic and high H_2O_2 conditions such as reduce to the high proportion of Mo^{5+} atoms. Furthermore, MoO_{3-x} exhibits OXD-like activity that could convert O_2 by disintegrated endogenous H_2O_2 to $\text{O}_2^{\cdot-}$ for tumor-specific catalytic therapy. This research provides a new potential therapeutic strategy to reduce the toxicity of nanozymes by controlling their acidic-responsive behavior.

Cobalt-based nanozymes

Intrinsic multienzyme-like activities of cobalt (Co)-based nanoparticles have been reported [90–92]. Dong et al. reported the Co_3O_4 possesses the pH-dependent enzyme-like property and the reaction basis is similar to Fe_3O_4 [93]. The Co_3O_4 showed optimal CAT-like reactivity and SOD-like activity at higher pH conditions ($\text{pH} \geq 7.4$), and the CAT-like reactivity of Co_3O_4 was significantly high than Fe_3O_4 at the same conditions. Additionally, Co_3O_4 exhibits higher POD-like activity



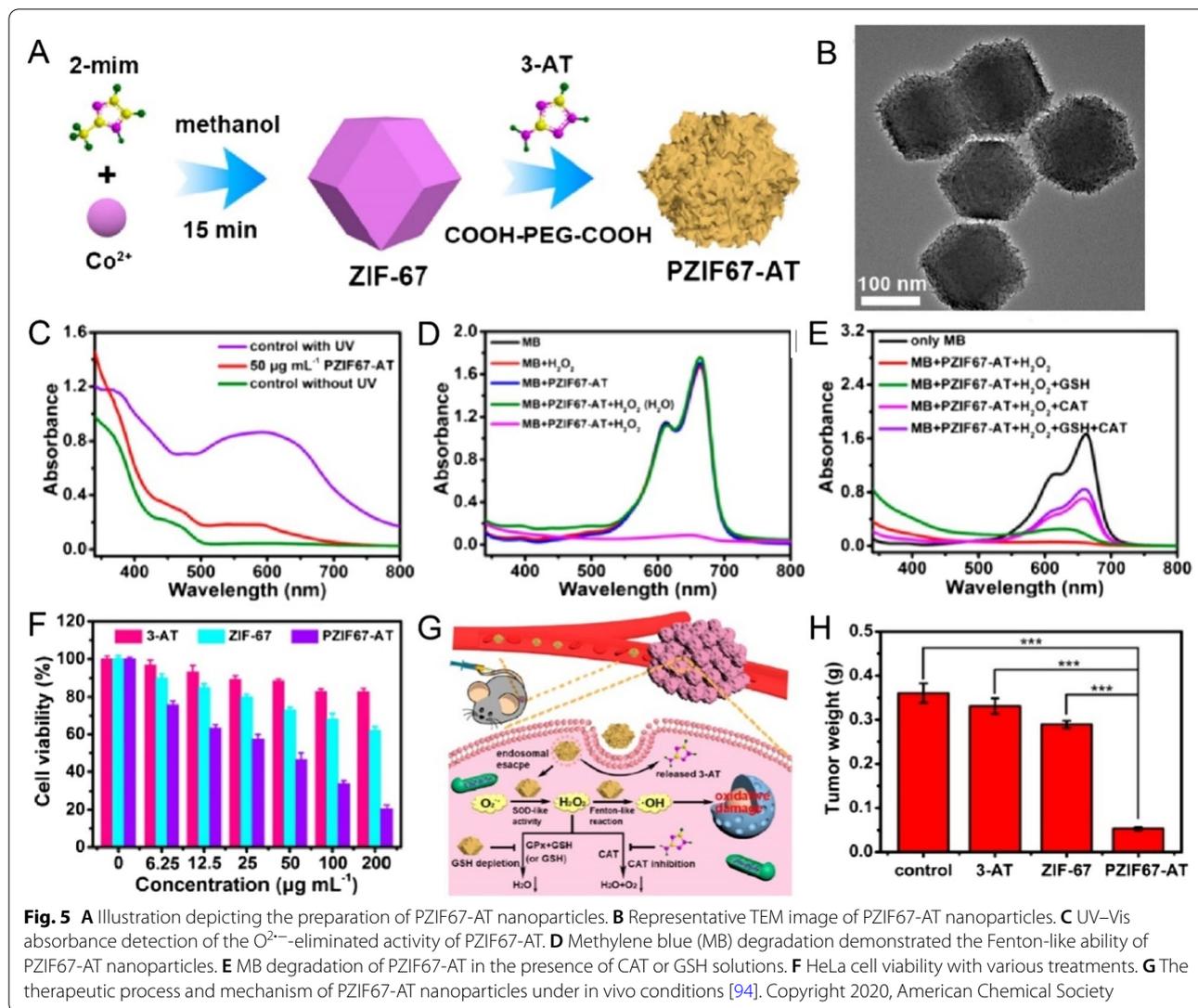
in an acidic medium (pH = 3.6). The catalyzing efficacy of Co-based nanozyme limits their biomedical application. Thus, improving the enzyme activity of nanozymes may be therapeutically attractive for better antitumor efficacy. Recently, Sang et al. developed a polyethylene glycol decorated PZIF67-AT nanoparticles by combining the multienzyme-like activities of Co-based zeolitic imidazole framework-67 and 3-amino-1,2,4-triazole (3-AT) [94]. In their design, the SOD-mimetic activity of PZIF67-AT initially converts $\text{O}_2^{\cdot-}$ to H_2O_2 (Fig. 5), subsequently, the production of H_2O_2 was converted to $\cdot\text{OH}$ by PZIF67-AT for cancer therapy. The CAT-like activity of PZIF67-AT was inhibited by 3-AT through binding of the Co-active center. In addition, the overexpressed GSH in TME could also be depleted by PZIF67-AT. This study offers an insight into nanozymes in the applications of tumor therapy.

Platinum-based nanozymes

Tumor hypoxia and overproduced H_2O_2 is the unique feature of solid tumor, which is critical to tumor proliferation and metastasis [95, 96]. Importantly, the therapeutic efficiency of current methods was limited by tumor hypoxia microenvironments, such as photodynamic therapy (PDT) and radiotherapy [97–99]. Unfortunately, oxygen-dependent PDT was severely discouraged due to the low intratumoral oxygen level [100–102]. Fortunately, nanozymes with catalase-like

activities could provide a feasible method to improve tumor oxygen-involved therapeutic methods [103, 104]. Based on the catalase-like activity, platinum (Pt) based nanomaterials have been widely applied to decompose the endogenous H_2O_2 to O_2 , thus relieving tumor hypoxia for tumor therapy, including PDT and radiotherapy [105, 106]. Zhang et al. report a Pt-PCN-224 nano-platform by decorating Pt on PCN-224 [106, 107]. In the overexpressed H_2O_2 microenvironment, Pt is capable of producing O_2 for the photosensitizer (PCN-224) during the photosensitization process to form $^1\text{O}_2$, which could remarkably enhance the outcome of tumor PDT. Besides, Li et al. synthesized porous Pt nanoparticles that can absorb X-ray and convert H_2O_2 to O_2 , improving the radiotherapy efficiency against the malignant tumor [108].

In addition, Pt-based nanozymes could also be used for ROS and inflammation associated with diseases. In another work, Lin et al. synthesized a cascade nanozyme of Pt@PCN222-Mn to realize anti-inflammatory therapy (Fig. 6) [109]. The PCN222-Mn with SOD-like activity can react with endogenous $\text{O}_2^{\cdot-}$, resulting in superoxide depletion and subsequent H_2O_2 generation. Then the Pt nanoparticles exhibited strong CAT-like performance to catalyze H_2O_2 for O_2 generation, thereby benefiting the inflammatory bowel diseases. This research provides the paradigm that the rationally designed nanozymes could have better

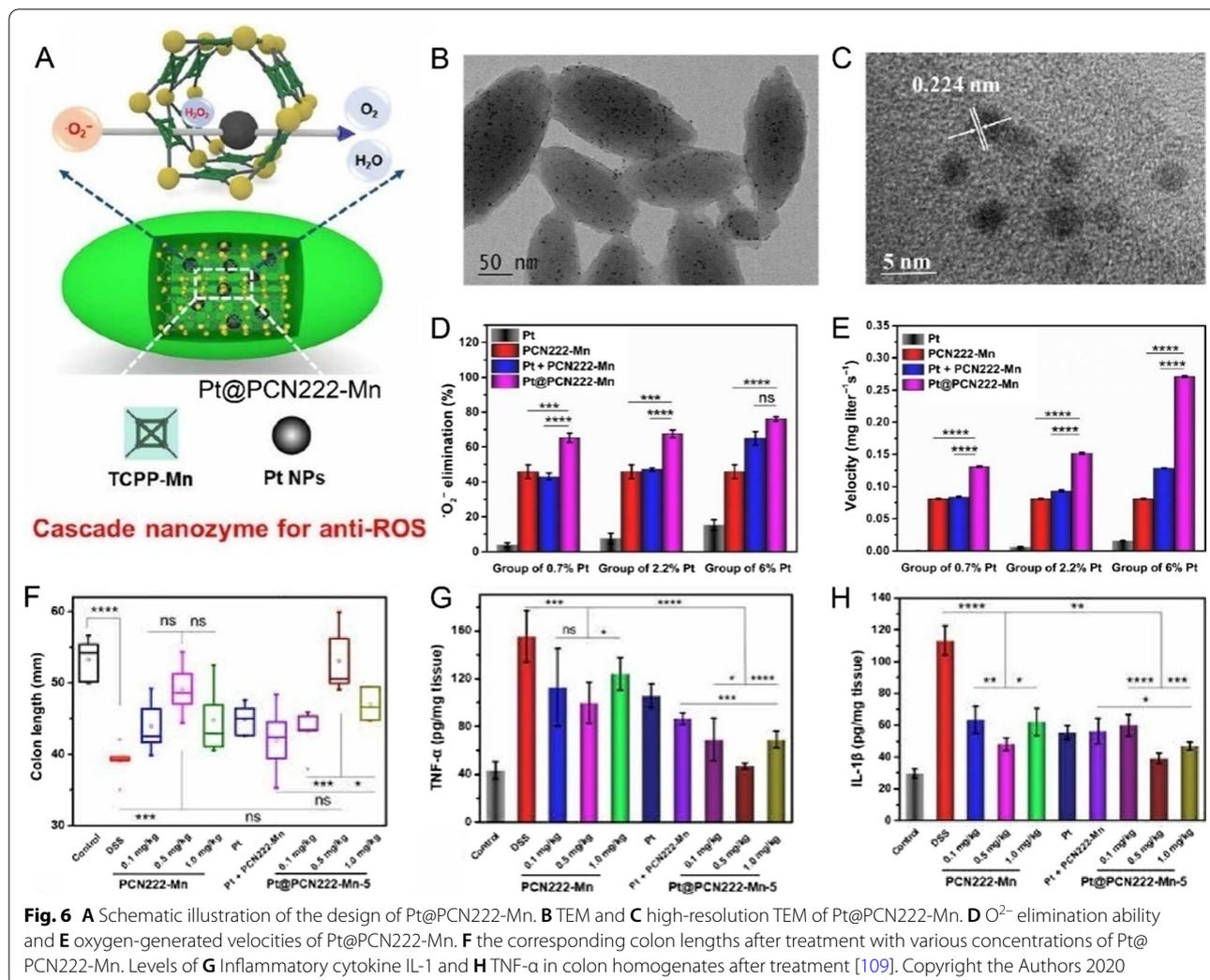


cascade enzymatic performance against pathologies in a variety.

Gold-based nanozymes

Enzymatic properties of gold nanoparticles (Au NPs) have widespread uses in biomedical applications [110]. Many studies have reported that Au NPs exhibited multiple-enzymes mimicking abilities such as peroxidase-mimetic activity and glucose oxidase (GOD) activity [111–113]. The GOD-like activity of Au nanoparticles can deplete the glucose and generate H_2O_2 , which could effectively consume glucose nutrients and inducing cell starving in tumor tissues. For example, Gao et al. synthesized the Au-containing inorganic nanozyme platform (DMSN-Au- Fe_3O_4 -PEG) [114]. Firstly, Au specifically catalyzes glucose to H_2O_2 , which was reacted with Fenton agent (Fe_3O_4) to produce highly toxic

hydroxyl radicals ($\cdot OH$) by typical Fenton reaction for tumor suppression. Additionally, the GOD-mimetic activity of Au nanoparticles has been reported to be synergized with CAT-mimetic nanomaterial enhanced tumor therapy efficiency. Liu et al. loaded Pt and Au into the porphyrin metal–organic frameworks (PCN) with folic acid decoration (P@Pt@P-Au-FA) [115]. The authors showed that endogenous H_2O_2 could be catalyzed by Pt to O_2 for enhanced PDT. The oxygen molecules act as the substrate for gold nanoparticles to convert glucose into H_2O_2 , supplying the reactant of Pt repeatedly. Within the oxygen cycle, remarkably consumption of glucose and production of gluconic acid could accelerate the catalytic efficiency and the antitumor efficiency of Au. However, the total O_2 level was not increased during this reaction cycle, low



intracellular O_2 levels still constrain tumor therapies that are oxygen-dependent.

The elevated GSH level in cancer cells enables tumor cells to maintain redox homeostasis and resistance to overexpression of ROS [116]. Depletion of GSH has been developed as a smart strategy for enhanced chemodynamic therapy, chemotherapy, photodynamic therapy, and radiotherapy [117]. However, redox homeostasis destruction has been rarely reported for cancer treatment. Based on the biochemical reactions between Au and thiol of GSH, Gong et al. synthesized single-atom Au nanoagents with GPx-like activity to amplify mitochondrial ROS for tumor therapy (Fig. 7) [118]. First, single-atom Au was incorporated into carbon dot (CAT-g) as metal centers catalysis of GSH. Then, triphenylphosphine and cinnamaldehyde were further employed to modify CAT-g (MitoCAT-g) to enable the mitochondria targeting ability and ROS generating

ability. As a result, MitoCAT-g effectively strengthened the oxidative stress in the mitochondrial of tumor cells and trigger apoptosis for cell death.

Iridium-based nanozymes

Other CAT-like nanozymes containing metal oxide-based nanomaterials, such as iridium (Ir) based nanoparticles, have been developed for biomedical applications [119, 120]. The mechanism of these metal oxide-based nanozymes was associated with the oxidation valent of metal species. For example, Su et al. investigated the connections between the CAT-like property of PVP-Ir(0) NPs and chemical state, demonstrating that the formation of IrO_2 upon exposure to H_2O_2 enables the PVP-IrNPs to exhibit CAT-like activity [121]. The POD activities of PVP-IrNPs were originated from electron transfer mediators. Zhang et al. demonstrated that PVP-IrNPs can scavenge ROS and reactive nitrogen species

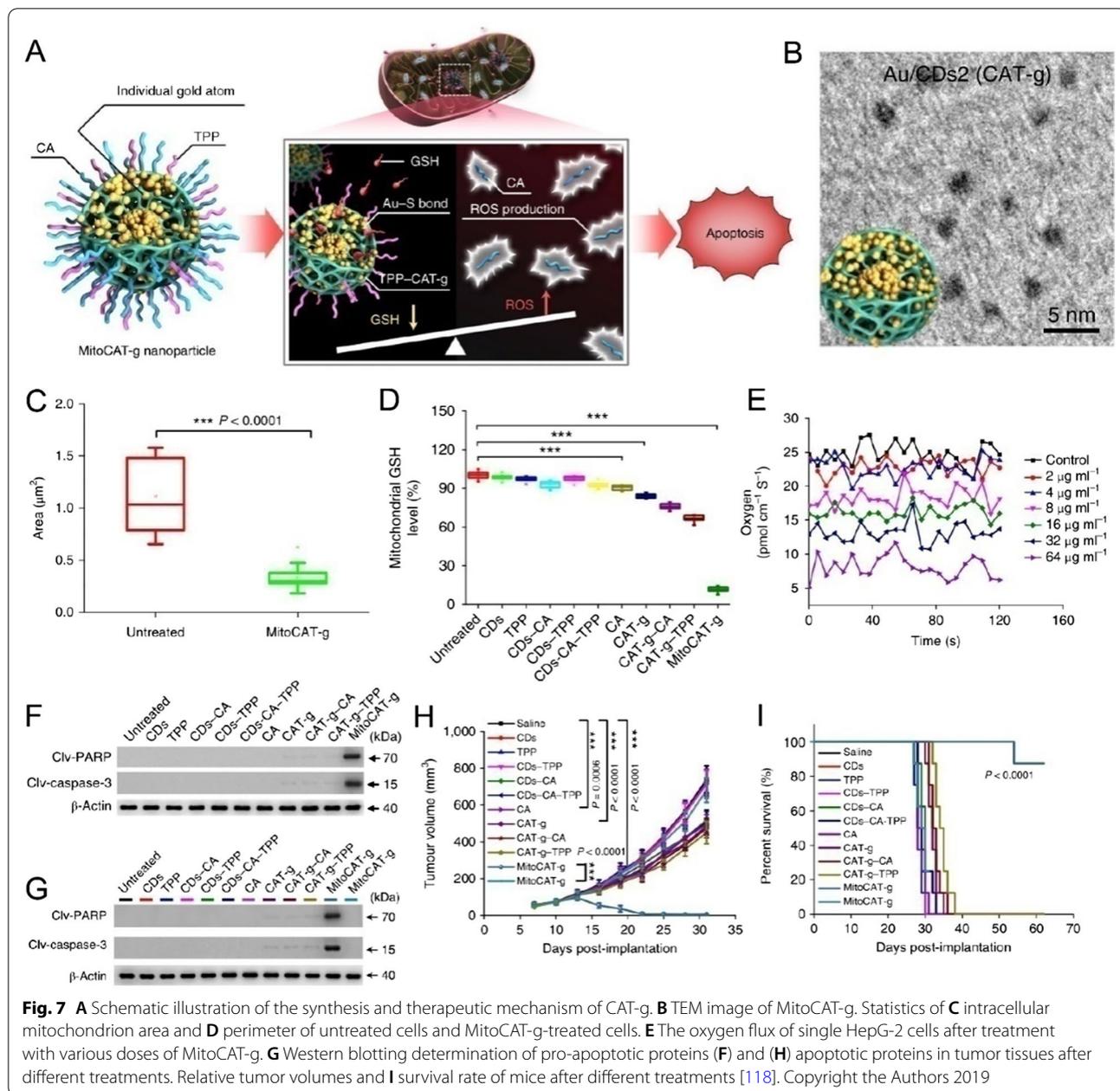


Fig. 7 **A** Schematic illustration of the synthesis and therapeutic mechanism of CAT-g. **B** TEM image of MitoCAT-g. Statistics of **C** intracellular mitochondrion area and **D** perimeter of untreated cells and MitoCAT-g-treated cells. **E** The oxygen flux of single HepG-2 cells after treatment with various doses of MitoCAT-g. **G** Western blotting determination of pro-apoptotic proteins (**F**) and (**H**) apoptotic proteins in tumor tissues after different treatments. Relative tumor volumes and **I** survival rate of mice after different treatments [118]. Copyright the Authors 2019

(RNS) to alleviate AKI [122]. In their work, ultrasmall PVP-IrNPs (1.5 nm) could rapidly accumulate to the kidney after intravenous administration, protecting ROS- or RNS-mediated cellular damage. Furthermore, PVP-IrNPs could be easily excreted to urine by the kidney and exhibit lower systemic toxicity. Besides, Ir-oxide (IrOx) has been reported that acid-activated OXD-like and pH-dependent CAT-like functions for targeted tumor therapies [123]. At neutral normal tissues, the IrOx presented dominantly CAT-like activities. While the POD-like and OXD-like activities were greatly improved along with the

gluconic acid generation by GOD catalysis. Importantly, the glutathione (GSH) can be consumed by Ir⁴⁺, dramatically reduced antioxidative species and enhanced lethality could be ultimately achieved.

Ruthenium-based nanozymes

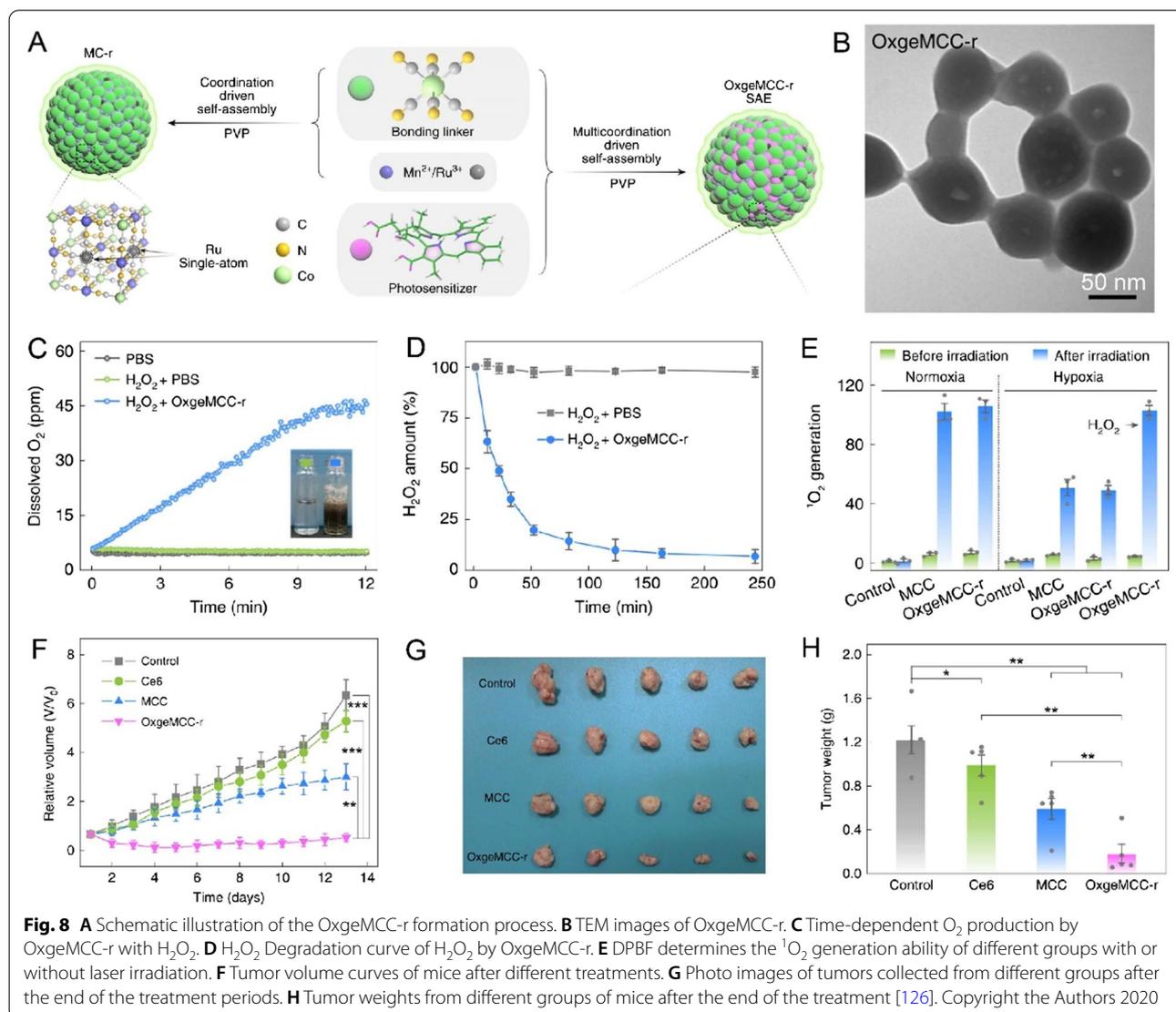
Recently, Xu et al. also discovered that ruthenium (Ru)-based nanoparticles with catalase-like activity could be constructed for highly efficient phototherapy against 4-T1 tumors [124]. In their work, RuO₂@BSA was first prepared by alkaline precipitation methods, and

photosensitizer (IR-808-Br₂) was subsequently decorated into the protein shell to form RuO₂@BSA@IR-808-Br₂. First, the RuO₂ possesses high CAT-like activity, endowing IR-808-Br₂ with highly efficient PDT activity. Second, the RuO₂ has photothermal conversion efficiency for PTT. As a result, RuO₂@BSA@IR-808-Br₂ achieves sufficient tumor inhibition by synergistically enhanced efficacy of PDT and PTT. The catalase-mimetic activity of RuO₂ was activated after being exposed to the tumor microenvironment, and immediately convert H₂O₂ to oxygenate the IR-808-Br₂ for the photodynamic process under near-infrared irradiation. Wei et al. reported a multi-functional IrRu-GOD@PEG NPs that could realize tumor starvation therapy and oxidative therapy by chemical catalysis from H₂O₂ to ¹O₂ [125]. Such an oxidative therapeutic strategy through IrRu alloy nanoparticles provides a new insight for tumor therapy.

Recently, the emerging single atom Ru has drawn extensive attention for endogenous O₂ generation. Wang et al. reported an O₂ generation single-atom Ru nano-platform (OxgeMCC-r) to enhance the therapeutic efficacy of PDT by self-assembly in the presence of PVP, Mn/Ru, and Ce6 (Fig. 8) [126]. Single-atom Ru allows effective O₂ generation at a low concentration of the Ru to overcome tumor hypoxia for ¹O₂-mediated tumor killing.

Conclusion and outlook

The fast development and revolution of nanoscience and nanotechnology has broadened extensive research interests for their application in biomedicine. Nanozymes is one of the emerging research frontiers that exhibit great prospects for disease therapy. Herein, we have summarized and discussed the most recent development of nanozymes with their intrinsic therapeutic



features for versatile biomedical applications. Despite the tremendous advantages of nanozymes in biomaterial applications, some critical issues and challenges are still needed to be considered. (1) The catalytic efficiency of most nanozymes should be further improved, with controllable enzyme-like activity. It is expected that the enzymatic reactions could be highly lesion site-specific, guaranteeing the biocompatibility and therapeutic specificity. To achieve high enzyme-like activity, introduction of the single-atom nanozymes is the most attractive strategy to achieve such issue, owing to their highly dispersion of the catalytic active sites and atomic utilization efficiency. Yet the loading efficiency of the single metal atoms is limited, challenges for the performance advancement of single-atom nanozymes are remained. In addition, rational design of cascade nanozymes may represent a promising strategy to improve the catalytic efficiency. (2) The molecular mechanism of nanozymes with multienzyme activities are still not clear. For instance, nanozymes mimicking dual-enzymes of CAT and POD, are supposed to exhibit self-competition performance during biomedical applications (as these enzymes both consume H_2O_2). The exact molecular mechanism of the electron movements within the metallic species should be further investigated under different conditions. In this regard, disease microenvironment holds great potential to regulate the desired enzymatic performance of specific nanozyme. Numerous studies have demonstrated the feasibility and responsiveness of nanozymes in broad biomedicine applications under characteristic stimuli, such as pH condition, GSH, and light, etc. Another strategy to regulate the specific enzymatic performance of the nanozyme lies in the application of specific inhibitors. (3) The selectivity and specificity of nanozymes should be further improved and optimized. During tumor therapy, although pH- or GSH-responsive nanozymes have exhibited “smart” enzyme-like activities to kill tumor cells, low selectivity and specificity have limited their further applications. To address this dilemma, rationally designed controllable nanozymes are significant to achieve high specificity of its enzyme-mimetic activity against various diseases. Recently, many studies have demonstrated that exogenous stimuli, such as light and ultrasonic, could serve as the trigger to control nanozymes activation. These modalities may provide feasible options to achieve the desired prominent site-specificity. (4) Biocompatibility and biodegradability should be considered. Overcoming the in vivo toxicity of nanozymes during therapeutics is still a barrier toward clinical application. Currently, systemic injection of nanozymes will inevitably cause adverse effect against normal tissues. For metal-based nanozymes, the toxicity is largely associated to the metallic species of the constructed metal-based nanozymes.

Although numerous studies have demonstrated the cytoprotective role and biocompatible character of nanozymes, metal ion release is still considered as the possible factor to cause the side impact against normal tissues due to the metal overload. For example, copper or iron overloaded in normal tissue/cells may trigger Fenton or Fenton-like reaction that could severely damage the biomacromolecules as well as the nucleic acids. Therefore, pharmacokinetics of the nanozymes are of critically important during the biocompatibility and biosafety evaluation. The surface tunable properties of nanozymes provide an opportunity to design biosafety agents. Taken into considerations, surface modification is one of the alternative strategies to overcome the limitation of nanozymes. Moreover, considering the ligands of nanozymes could influence therapeutic outcomes, bioavailability, clearance dynamics, and systemic toxicity. From this perspective, it is necessary to carefully choose a suitable ligand and endow nanozymes with higher biosafety.

Abbreviations

GSH: Glutathione; OXD: Oxidase; GOD: Glucose oxidase; POD: Peroxidase; CAT: Catalase; SOD: Superoxide dismutase; GPx: Glutathione peroxidase; TME: Tumor microenvironment; $O_2^{\cdot-}$: Superoxide anion; H_2O_2 : Hydrogen peroxide; ROS: Reactive oxygen species; RNS: Reactive nitrogen species; CcO: Cytochrome c oxidase; TNF- α : Tumor necrosis factor alpha; LDH: Lactate dehydrogenase; SPN-C23: Self-regulated nanoceria-doped poly(cyclopentadithiophene-alt-benzothiadiazole); PDT: Photodynamic therapy; NIR: Near-infrared; SOSG: Singlet oxygen sensor green; AMD: Age-related macular degeneration; 1O_2 : Single oxygen species; AMD: Age-related macular degeneration; ZIF-8: Zeolitic imidazolate framework-8; t-BOOH: Tert-butyl hydroperoxide; eDNA: Extracellular DNA; MOF: Metal-organic framework; MRI: Magnetic resonance imaging; ESR: Electron spin resonance; HFn: Human ferritin; Cu-Cys: Copper-amino acid mercaptide nanomaterials; Phe: Phenylalanine; TA: Tannic acid; M-NSs: MnO_2 nanosheets; $MoO_3 \cdot x$: $MoO_3 \cdot x$ nanorods; 3-AT: 3-Amino-1,2,4-triazole; PCN: Porphyrin metal-organic frameworks.

Authors' contributions

XR and DC wrote the manuscript, YW, HL, and YZ checked different sections of the manuscript. HC, XL and MH edited the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare no conflicts of interests.

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References

- Wu J, Wang X, Wang Q, et al. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II). *Chem Soc Rev*. 2019;48:1004–76.
- Wang D, Jana D, Zhao Y. Metal-organic framework derived nanozymes in biomedicine. *Acc Chem Res*. 2020;53:1389–400.
- Fan Y, Liu S, Yi Y, et al. Catalytic nanomaterials toward atomic levels for biomedical applications: from metal clusters to single-atom catalysts. *ACS Nano*. 2021;15:2005–37.
- Zhang L, Zhang L, Deng H, et al. In vivo activation of pH-responsive oxidase-like graphitic nanozymes for selective killing of *Helicobacter pylori*. *Nat Commun*. 2021;12:2002.
- Gao L, Yan X. Nanozymes: biomedical applications of enzymatic Fe₃O₄ nanoparticles from in vitro to in vivo. *Adv Exp Med Biol*. 2019;1174:291–312.
- Yang M, Li J, Gu P, et al. The application of nanoparticles in cancer immunotherapy: targeting tumor microenvironment. *Bioact Mater*. 2021;6:1973–87.
- Liu F, Lin L, Zhang Y, et al. A tumor-microenvironment-activated nanozyme-mediated theranostic nanoreactor for imaging-guided combined tumor therapy. *Adv Mater*. 2019;31:e1902885.
- Wang Z, Li Z, Sun Z, et al. Visualization nanozyme based on tumor microenvironment “unlocking” for intensive combination therapy of breast cancer. *Sci Adv*. 2020;6:eabc8733.
- Lin LS, Song J, Song L, et al. Simultaneous fenton-like ion delivery and glutathione depletion by MnO₂-based nanoagent to enhance chemodynamic therapy. *Angew Chem Int Ed*. 2018;57:4902–6.
- Cao S, Fan J, Sun W, et al. A novel Mn-Cu bimetallic complex for enhanced chemodynamic therapy with simultaneous glutathione depletion. *Chem Commun*. 2019;55:12956–9.
- Fu LH, Wan Y, Qi C, et al. Nanocatalytic theranostics with glutathione depletion and enhanced reactive oxygen species generation for efficient cancer therapy. *Adv Mater*. 2021;33:e2006892.
- Wang C, Wang H, Xu B, et al. Photo-responsive nanozymes: mechanism, activity regulation, and biomedical applications. *View*. 2020;2:20200045.
- Dong H, Fan Y, Zhang W, et al. Catalytic mechanisms of nanozymes and their applications in biomedicine. *Bioconjugate Chem*. 2019;30:1273–96.
- Ji S, Jiang B, Hao H, et al. Matching the kinetics of natural enzymes with a single-atom iron nanozyme. *Nat Catal*. 2021;4:407–17.
- Huang Y, Ren J, Qu X. Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chem Rev*. 2019;119:4357–412.
- Wang H, Wan K, Shi X. Recent advances in nanozyme research. *Adv Mater*. 2019;31:e1805368.
- Liu X, Gao Y, Chandrawati R, et al. Therapeutic applications of multifunctional nanozymes. *Nanoscale*. 2019;11:21046–60.
- Zhang L, Jiang C, Li B, Liu Z, et al. A core-shell Au@Cu_{2-x}Se heterogeneous metal nanocomposite for photoacoustic and computed tomography dual-imaging-guided photothermal boosted chemodynamic therapy. *J Nanobiotechnol*. 2021;19:1–18.
- Meng X, Li D, Chen L, et al. High-performance self-cascade pyrite nanozymes for apoptosis-ferroptosis synergistic tumor therapy. *ACS Nano*. 2021;15:5735–51.
- Liu J, Wang A, Liu S, et al. A titanium nitride nanozyme for pH-responsive and irradiation-enhanced cascade-catalytic tumor therapy. *Angew Chem Int Ed*. 2021;60:25328–38.
- Ghorbani M, Derakhshankhah H, Jafari S, et al. Nanozyme antioxidants as emerging alternatives for natural antioxidants: achievements and challenges in perspective. *Nano Today*. 2019;29:100775.
- Liu C, Yao J, Hu J, et al. Navigating nMOF-mediated enzymatic reactions for catalytic tumor-specific therapy. *Mater Horiz*. 2020;7:3176–86.
- Li F, Qiu Y, Xia F, et al. Dual detoxification and inflammatory regulation by ceria nanozymes for drug-induced liver injury therapy. *Nano Today*. 2020;35:100925.
- Fan Y, Li P, Hu B, et al. A smart photosensitizer-cerium oxide nanoprobe for highly selective and efficient photodynamic therapy. *Inorg Chem*. 2019;58:7295–302.
- Zhang S, Liu Y, Sun S, et al. Catalytic patch with redox Cr/CeO₂ nanozyme of noninvasive intervention for brain trauma. *Theranostics*. 2021;11:2806–21.
- Singh N, Mughes G. CeVO₄ nanozymes catalyze the reduction of dioxygen to water without releasing partially reduced oxygen species. *Angew Chem Int Ed*. 2019;58:7797–801.
- Wang Z, Shen X, Gao X, et al. Simultaneous enzyme mimicking and chemical reduction mechanisms for nanoceria as a bio-antioxidant: a catalytic model bridging computations and experiments for nanozymes. *Nanoscale*. 2019;11:13289–99.
- Esch F, Fabris S, Zhou L, et al. Electron localization determines defect formation on ceria substrates. *Science*. 2005;309:752–5.
- Forest V, Leclerc L, Hochepeid J, et al. Impact of cerium oxide nanoparticles shape on their in vitro cellular toxicity. *Toxicol In Vitro*. 2017;38:136–41.
- Qian Q, Zhang Y, Chen Y, et al. Assessment of pulmonary toxicity of potential antioxidant drug PEGylated nanoceria after intratracheal instillation in rats. *J Appl Toxicol*. 2021;41:941–52.
- Zhu H, Fang Y, Miao Q, et al. Regulating near-infrared photodynamic properties of semiconducting polymer nanotheranostics for optimized cancer therapy. *ACS Nano*. 2017;11:8998–9009.
- Zhang C, Wang X, Du J, et al. Reactive oxygen species-regulating strategies based on nanomaterials for disease treatment. *Adv Sci*. 2021;8:2002797.
- Lian M, Xue Z, Qiao X, et al. Movable hollow nanoparticles as reactive oxygen scavengers. *Chem*. 2019;5:2378–87.
- Stavros P, Sikha S. Therapeutic benefits of nanoparticles in stroke. *Front Neurosci*. 2015;9:182.
- Nash K, Ahmed S. Nanomedicine in the ROS-mediated pathophysiology: applications and clinical advances. *Nanomedicine*. 2015;11:2033–40.
- Yan R, Sun S, Yang J, et al. Nanozyme-based bandage with single-atom catalysis for brain trauma. *ACS Nano*. 2019;13:11552–60.
- Yildirim Z, Ucgun N, Yildirim F. The role of oxidative stress and antioxidants in the pathogenesis of age-related macular degeneration. *Clinics*. 2011;66:743–6.
- Zhang K, Tu M, Gao W, et al. Hollow Prussian blue nanozymes drive neuroprotection against ischemic stroke via attenuating oxidative stress, counteracting inflammation, and suppressing cell apoptosis. *Nano Lett*. 2019;19:2812–23.
- Abokyi S, To C, Lam T, et al. Central role of oxidative stress in age-related macular degeneration: evidence from a review of the molecular mechanisms and animal models. *Oxid Med Cell Longevity*. 2020;2020:7901270.
- Kauppinen A, Paterno JJ, Blasiak J, et al. Inflammation and its role in age-related macular degeneration. *Cell Mol Life Sci*. 2016;73:1765–86.
- Mitra RN, Gao R, Zheng M, et al. Glycol chitosan engineered autoregenerative antioxidant significantly attenuates pathological damages in models of age-related macular degeneration. *ACS Nano*. 2017;11:4669–85.
- Ni D, Wei H, Chen W, et al. Ceria nanoparticles meet hepatic ischemia-reperfusion injury: the perfect imperfection. *Adv Mater*. 2019;31:e1902956.
- He L, Huang G, Liu H, et al. Highly bioactive zeolitic imidazolate framework-8-capped nanotherapeutics for efficient reversal of reperfusion-induced injury in ischemic stroke. *Sci Adv*. 2020;6:eaay9751.

44. Liu Z, Wang F, Ren J, et al. A series of MOF/Ce-based nanozymes with dual enzyme-like activity disrupting biofilms and hindering recolonization of bacteria. *Biomaterials*. 2019;208:21–31.
45. Xu F, Lu Q, Huang PJ, et al. Nanoceria as a DNase I mimicking nanozyme. *Chem Commun*. 2019;55:13215–8.
46. Huo M, Wang L, Wang Y, et al. Nanocatalytic tumor therapy by single-atom catalysts. *ACS Nano*. 2019;13:2643–53.
47. Xu B, Cui Y, Wang W, et al. Immunomodulation-enhanced nanozyme-based tumor catalytic therapy. *Adv Mater*. 2020;32:e2003563.
48. Huo M, Wang L, Chen Y, et al. Tumor-selective catalytic nanomedicine by nanocatalyst delivery. *Nat Commun*. 2017;8:357.
49. Zhou W, Gao J, Zhao H, et al. The role of quinone cycle in Fe^{2+} - H_2O_2 system in the regeneration of Fe^{2+} . *Environ Technol*. 2017;38:1887–96.
50. Pham AL, Doyle FM, Sedlak DL. Kinetics and efficiency of H_2O_2 activation by iron-containing minerals and aquifer materials. *Water Res*. 2012;46:6454–62.
51. Gao L, Zhuang J, Nie L, et al. Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat Nanotechnol*. 2007;2:577–83.
52. Wang B, Yin JJ, Zhou X, et al. Physicochemical origin for free radical generation of iron oxide nanoparticles in biomicroenvironment: catalytic activities mediated by surface chemical states. *J Phys Chem C*. 2012;117:383–92.
53. Chen Z, Yin J, Zhou Y, et al. Dual enzyme-like activities of iron oxide nanoparticles and their implication for diminishing cytotoxicity. *ACS Nano*. 2012;6:4001–12.
54. Zhao S, Duan H, Yang Y, et al. Fenozyme protects the integrity of the blood-brain barrier against experimental cerebral malaria. *Nano Lett*. 2019;19:8887–95.
55. Shan J, Li X, Yang K, et al. Efficient bacteria killing by Cu_2WS_4 nanocrystals with enzyme-like properties and bacteria-binding ability. *ACS Nano*. 2019;13:13797–808.
56. Sun S, Chen Q, Tang Z, et al. Tumor microenvironment stimuli-responsive fluorescence imaging and synergistic cancer therapy by carbon-dot- Cu^{2+} nanoassemblies. *Angew Chem Int Ed*. 2020;59:21041–8.
57. Khoris IM, Ganganboina AB, Suzuki T, et al. Self-assembled chromogen-loaded polymeric cocoon for respiratory virus detection. *Nanoscale*. 2021;13:388–96.
58. Wang Y, Li Z, Hu Y, et al. Photothermal conversion-coordinated Fenton-like and photocatalytic reactions of Cu_{2-x}Se -Au Janus nanoparticles for tri-combination antitumor therapy. *Biomaterials*. 2020;255:120167.
59. Soltani T, Lee BK. Enhanced formation of sulfate radicals by metal-doped BiFeO_3 under visible light for improving photo-Fenton catalytic degradation of 2-chlorophenol. *Chem Eng J*. 2017;313:1258–68.
60. Ma B, Wang S, Liu F, et al. Self-assembled copper-amino acid nanoparticles for in situ glutathione and H_2O_2 sequentially triggered chemodynamic therapy. *J Am Chem Soc*. 2019;141:849–57.
61. Xi J, Wei G, An L, et al. Copper/carbon hybrid nanozyme: tuning catalytic activity by the copper state for antibacterial therapy. *Nano Lett*. 2019;19:7645–54.
62. Karim MN, Singh M, Weerathunge P, et al. Visible-light-triggered reactive-oxygen-species-mediated antibacterial activity of peroxidase-mimic CuO nanorods. *ACS Appl Mater Interfaces*. 2018;1:1694–704.
63. Chen M, Wang Z, Shu J, et al. Mimicking a natural enzyme system: cytochrome c oxidase-like activity of Cu_2O nanoparticles by receiving electrons from cytochrome c. *Inorg Chem*. 2017;56:9400–3.
64. Hao C, Qu A, Xu L, et al. Chiral molecule-mediated porous Cu_2O nanoparticle clusters with antioxidant activity for ameliorating Parkinson's disease. *J Am Chem Soc*. 2019;141:1091–9.
65. Korschelt K, Ragg R, Metzger CS, et al. Glycine-functionalized copper(II) hydroxide nanoparticles with high intrinsic superoxide dismutase activity. *Nanoscale*. 2017;9:3952–60.
66. Lin S, Cheng Y, Zhang H, et al. Copper tannic acid coordination nanosheet: a potent nanozyme for scavenging ROS from cigarette smoke. *Small*. 2019;27:e1902123.
67. Liu T, Xiao B, Xiang F, et al. Ultrasmall copper-based nanoparticles for reactive oxygen species scavenging and alleviation of inflammation related diseases. *Nat Commun*. 2020;11:2788.
68. Kuthati Y, Busa P, Goutham Davuluri VN, et al. Manganese oxide nanozymes ameliorate mechanical allodynia in a rat model of partial sciatic nerve-transection induced neuropathic pain. *Int J Nanomed*. 2019;14:10105–17.
69. Singh N, Savanur MA, Srivastava S, et al. A manganese oxide nanozyme prevents the oxidative damage of biomolecules without affecting the endogenous antioxidant system. *Nanoscale*. 2019;11:3855–63.
70. Fu L, Hu Y, Qi C, et al. Biodegradable manganese-doped calcium phosphate nanotheranostics for traceable cascade reaction-enhanced anti-tumor therapy. *ACS Nano*. 2019;13:13985–94.
71. He M, Chen Y, Tao C, et al. Mn-porphyrin-based metal-organic framework with high longitudinal relaxivity for magnetic resonance imaging guidance and oxygen self-supplementing photodynamic therapy. *ACS Appl Mater Interfaces*. 2019;11:41946–56.
72. Liu Y, Pan Y, Wei C, et al. A tumor microenvironment responsive biodegradable $\text{CaCO}_3/\text{MnO}_2$ -based nanopatform for the enhanced photodynamic therapy and improved PD-L1 immunotherapy. *Theranostics*. 2019;23:6867–84.
73. Jia Q, Ge J, Liu W, et al. A magnetofluorescent carbon dot assembly as an acidic H_2O_2 -driven oxygenator to regulate tumor hypoxia for simultaneous bimodal imaging and enhanced photodynamic therapy. *Adv Mater*. 2018;30:e1706090.
74. Fan H, Yan G, Zhao Z, et al. A smart photosensitizer-manganese dioxide nanosystem for enhanced photodynamic therapy by reducing glutathione levels in cancer cells. *Angew Chem Int Ed*. 2016;55:5477–82.
75. Ding B, Zheng P, Ma P, et al. Manganese oxide nanomaterials: synthesis, properties, and theranostic applications. *Adv Mater*. 2020;32:e1905823.
76. Kim J, Cho HR, Jeon H, et al. Continuous O_2 -evolving MnFe_2O_4 nanoparticle-anchored mesoporous silica nanoparticles for efficient photodynamic therapy in hypoxic cancer. *J Am Chem Soc*. 2017;139:10992–5.
77. Yin SY, Song G, Yang Y, et al. Persistent regulation of tumor microenvironment via circulating catalysis of MnFe_2O_4 @Metal-organic frameworks for enhanced photodynamic therapy. *Adv Funct Mater*. 2019;29:1901417.
78. Meng L, Cheng Y, Tong X, et al. Tumor oxygenation and hypoxia inducible factor-1 functional inhibition via a reactive oxygen species responsive nanopatform for enhancing radiation therapy and abscopal effects. *ACS Nano*. 2018;12:8308–22.
79. Singh N, Savanur MA, Srivastava S, et al. A redox modulatory Mn_3O_4 nanozyme with multi-enzyme activity provides efficient cytoprotection to human cells in a Parkinson's disease model. *Angew Chem Int Ed*. 2017;56:14267–71.
80. Yao J, Cheng Y, Zhou M, et al. ROS scavenging Mn_3O_4 nanozymes for in vivo anti-inflammation. *Chem Sci*. 2018;9:2927–33.
81. Shi J, Yu W, Xu L, et al. Bioinspired nanosponge for salvaging ischemic stroke via free radical scavenging and self-adapted oxygen regulating. *Nano Lett*. 2020;20:780–9.
82. Tang W, Fan W, Zhang W, et al. Wet/Sono-chemical synthesis of enzymatic two-dimensional MnO_2 nanosheets for synergistic catalysis-enhanced phototheranostics. *Adv Mater*. 2019;31:e1900401.
83. Chen Y, Chen T, Wu X, et al. Oxygen vacancy-engineered PEGylated MoO_3 nanoparticles with superior sulfite oxidase mimetic activity for vitamin B_1 detection. *Small*. 2019;15:e1903153.
84. Ni D, Jiang D, Kuttyreff C, et al. Molybdenum-based nanoclusters act as antioxidants and ameliorate acute kidney injury in mice. *Nat Commun*. 2018;9:5421.
85. Li S, Jiang D, Ehlerding EB, et al. Intrathecal administration of nanoclusters for protecting neurons against oxidative stress in cerebral ischemia/reperfusion injury. *ACS Nano*. 2019;13:13382–9.
86. Zhang Y, Li D, Tan J, et al. Near-infrared regulated nanozymatic/photothermal/photodynamic triple-therapy for combating multidrug-resistant bacterial infections via oxygen-vacancy molybdenum trioxide nanodots. *Small*. 2021;17:e2005739.
87. Ren C, Li D, Zhou Q, et al. Mitochondria-targeted TPP- MoS_2 with dual enzyme activity provides efficient neuroprotection through M1/M2 microglial polarization in an Alzheimer's disease model. *Biomaterials*. 2020;232:119752.
88. Han Q, Wang X, Liu X, et al. MoO_3 nanodots with dual enzyme mimic activities as multifunctional modulators for amyloid assembly and neurotoxicity. *J Colloid Interface Sci*. 2019;539:575–84.
89. Hu X, Li F, Xia F, et al. Biodegradation-mediated enzymatic activity-tunable molybdenum oxide nanourchins for tumor-specific cascade catalytic therapy. *J Am Chem Soc*. 2019;142:1636–44.
90. Mu J, Wang Y, Zhao M, et al. Intrinsic peroxidase-like activity and catalase-like activity of Co_3O_4 nanoparticles. *Chem Commun*. 2012;48:2540–2.

91. Wang J, Wang Y, Zhang D, et al. Intrinsic oxidase-like nanoenzyme $\text{Co}_4\text{S}_3/\text{Co}(\text{OH})_2$ hybrid nanotubes with broad-spectrum antibacterial activity. *ACS Appl Mater Interfaces*. 2020;12:29614–24.
92. Li S, Sun W, Luo Y, et al. Hollow PtCo alloy nanospheres as a high-Z and oxygen generating nanozyme for radiotherapy enhancement in non-small cell lung cancer. *J Mater Chem B*. 2021;23:4643–53.
93. Dong J, Song L, Yin JJ, et al. Co_3O_4 nanoparticles with multi-enzyme activities and their application in immunohistochemical assay. *ACS Appl Mater Interfaces*. 2014;6:1959–70.
94. Sang Y, Cao F, Li W, et al. Bioinspired construction of a nanozyme-based H_2O_2 homeostasis disruptor for intensive chemodynamic therapy. *J Am Chem Soc*. 2020;142:5177–83.
95. Wang L, Huo M, Chen Y, et al. Tumor microenvironment-enabled nanotherapy. *Adv Healthcare Mater*. 2018;7:e1701156.
96. Cao Z, Zhang L, Liang K, et al. Biodegradable 2D Fe-Al hydroxide for nanocatalytic tumor-dynamic therapy with tumor specificity. *Adv Sci*. 2018;5:1801155.
97. Jarosz-Biej M, Smolarczyk R, Cichoń T, et al. Tumor microenvironment as a “Game Changer” in cancer radiotherapy. *Int J Mol Sci*. 2019;20:3212.
98. Li X, Kwon N, Guo T, et al. Innovative strategies for hypoxic-tumor photodynamic therapy. *Angew Chem Int Ed*. 2018;57:11522–31.
99. Graham K, Unger E. Overcoming tumor hypoxia as a barrier to radiotherapy, chemotherapy and immunotherapy in cancer treatment. *Int J Nanomed*. 2018;13:6049–58.
100. Kwiatkowski S, Knap B, Przystupski D, et al. Photodynamic therapy-mechanisms, photosensitizers and combinations. *Biomed Pharmacother*. 2018;106:1098–107.
101. Zhang C, Chen W, Zhang T, et al. Hybrid nanoparticle composites applied to photodynamic therapy: strategies and applications. *J Mater Chem B*. 2020;8:4726–37.
102. Chen J, Fan T, Xie Z, et al. Advances in nanomaterials for photodynamic therapy applications: status and challenges. *Biomaterials*. 2020;237:119827.
103. Gao Z, Li Y, Zhang Y, et al. Biomimetic platinum nanozyme immobilized on 2D Metal-organic frameworks for mitochondrion-targeting and oxygen self-supply photodynamic therapy. *ACS Appl Mater Interfaces*. 2020;12:1963–72.
104. Wang D, Zhang N, Jing X, et al. A tumor-microenvironment fully responsive nano-platform for MRI-guided photodynamic and photothermal synergistic therapy. *J Mater Chem B*. 2020;8:8271–81.
105. Cao H, Yang Y, Liang M, et al. Pt@polydopamine nanoparticles as nanozymes for enhanced photodynamic and photothermal therapy. *Chem Commun*. 2021;57:255–8.
106. Yang Y, Zhu D, Liu Y, et al. Platinum-carbon-integrated nanozymes for enhanced tumor photodynamic and photothermal therapy. *Nanoscale*. 2020;12:13548–57.
107. Zhang Y, Wang F, Liu C, et al. Nanozyme decorated metal-organic frameworks for enhanced photodynamic therapy. *ACS Nano*. 2018;12:651–61.
108. Li Y, Yun KH, Lee H, et al. Porous platinum nanoparticles as a high-Z and oxygen generating nanozyme for enhanced radiotherapy in vivo. *Biomaterials*. 2019;197:12–9.
109. Liu Y, Cheng Y, Zhang H, et al. Integrated cascade nanozyme catalyzes in vivo ROS scavenging for anti-inflammatory therapy. *Sci Adv*. 2020;6:eabb2695.
110. Petree JR, Yehl K, Galior K, et al. Site-selective RNA splicing nanozyme: DNzyme and RtcB conjugates on a gold nanoparticle. *ACS Chem Biol*. 2018;13:215–24.
111. Chen J, Ma Q, Li M, et al. Glucose-oxidase like catalytic mechanism of noble metal nanozymes. *Nat Commun*. 2021;12:3375.
112. Fan L, Xu X, Zhu C, et al. Tumor catalytic-photothermal therapy with yolk-shell Gold@Carbon nanozymes. *ACS Appl Mater Interfaces*. 2018;10:4502–11.
113. Dan Q, Hu D, Ge Y, et al. Ultrasmall theranostic nanozymes to modulate tumor hypoxia for augmenting photodynamic therapy and radiotherapy. *Biomater Sci*. 2020;8:973–87.
114. Gao S, Lin H, Zhang H, et al. Nanocatalytic tumor therapy by biomimetic dual inorganic nanozyme-catalyzed cascade reaction. *Adv Sci*. 2019;6:1801733.
115. Liu C, Xing J, Akakuru OU, et al. Nanozymes-engineered metal-organic frameworks for catalytic cascades-enhanced synergistic cancer therapy. *Nano Lett*. 2019;19:5674–82.
116. Guo X, Wang L, Duval K, et al. Dimeric drug polymeric micelles with acid-active tumor targeting and FRET-traceable drug release. *Adv Mater*. 2018;30:1705436.
117. Zhang TT, Xu CH, Zhao W, et al. A redox-activated theranostic nano-agent: toward multi-mode imaging guided chemo-photothermal therapy. *Chem Sci*. 2018;9:6749–57.
118. Gong N, Ma X, Ye X, et al. Carbon-dot-supported atomically dispersed gold as a mitochondrial oxidative stress amplifier for cancer treatment. *Nat Nanotechnol*. 2019;14:379–87.
119. Feng L, Dong Z, Liang C, et al. Iridium nanocrystals encapsulated liposomes as near-infrared light controllable nanozymes for enhanced cancer radiotherapy. *Biomaterials*. 2018;181:81–91.
120. Zhen W, Liu Y, Lin L, et al. BSA-IrO₂: catalase-like nanoparticles with high photothermal conversion efficiency and a high X-ray absorption coefficient for anti-inflammation and antitumor theranostics. *Angew Chem Int Ed*. 2018;57:10309–13.
121. Xu P, Wang X, Li T, et al. Biomineralization-inspired nanozyme for single-wavelength laser activated photothermal-photodynamic synergistic treatment against hypoxic tumors. *Nanoscale*. 2020;12:4051–60.
122. Su H, Liu DD, Zhao M, et al. Dual-enzyme characteristics of polyvinylpyrrolidone-capped iridium nanoparticles and their cellular protective effect against H_2O_2 -induced oxidative damage. *ACS Appl Mater Interfaces*. 2015;7:8233–42.
123. Zhang DY, Younis MR, Liu H, et al. Multi-enzyme mimetic ultrasmall iridium nanozymes as reactive oxygen/nitrogen species scavengers for acute kidney injury management. *Biomaterials*. 2021;271:120706.
124. Zhen W, Liu Y, Wang W, et al. Specific “Unlocking” of a nanozyme-based butterfly effect to break the evolutionary fitness of chaotic tumors. *Angew Chem Int Ed*. 2020;59:9491–7.
125. Wei C, Liu Y, Zhu X, et al. Iridium/ruthenium nanozyme reactors with cascade catalytic ability for synergistic oxidation therapy and starvation therapy in the treatment of breast cancer. *Biomaterials*. 2020;238:119848.
126. Wang D, Wu H, Phua S, et al. Self-assembled single-atom nanozyme for enhanced photodynamic therapy treatment of tumor. *Nat Commun*. 2020;11:357.

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